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(54) Title: ANTIBODIES THAT BLOCK RECEPTOR PROTEIN TYROSINE KINASE ACTIVATION, METHODS OF SCREENING FOR AND USES THEREOF

(57) Abstract: Molecules containing the antigen-binding portion of antibodies that block constitutive and/or ligand-dependent activation of a receptor protein tyrosine kinase, such as fibroblast growth factor receptor 3 (FGFR3), are found through screening methods, where a soluble dimeric form of a receptor protein tyrosine kinase is used as target for screening a library of antibody fragments displayed on the surface of bacteriophage. The molecules of the present invention which block constitutive activation can be administered to treat or inhibit skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression associated with the constitutive activation of a receptor protein tyrosine kinase.

ANTIBODIES THAT BLOCK RECEPTOR PROTEIN TYROSINE KINASE ACTIVATION, METHODS OF SCREENING FOR AND USES THEREOF

Field of the Invention

The present invention relates to: immunoglobulins (and functional fragments thereof) useful for blocking activation of receptor protein tyrosine kinases, methods for screening for such immunoglobulins, compositions comprising said immunoglobulins and methods of using the same for treating or inhibiting disease, such as skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression.

10 Background of the Invention

A wide variety of biological processes involves complex cellular communication mechanisms. One of the primary means of continual exchange of information between cells and their internal and external environments is via the secretion and specific binding of peptide growth factors. Growth factors exert pleiotropic effects and play important roles in oncogenesis and the development of multicellular organisms regulating cell growth, differentiation and migration. Many of these factors mediate their effects by binding to specific cell surface receptors. The ligand-activated receptors start an enzymatic signal transduction cascade from the cell membrane to the cell nucleus, resulting in specific gene regulation and diverse cellular responses.

20 Protein Kinases

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One of the key biochemical mechanisms of signal transduction involves the reversible phosphorylation of proteins, which enables regulation of the activity of mature proteins by altering their structure and function.

Protein kinases ("PKs") are enzymes that catalyze the phosphorylation of hydroxy groups on tyrosine, serine and threonine residues of proteins. The consequences of this seemingly simple activity are staggering; cell growth, differentiation and proliferation; e.g., virtually all aspects of cell life in one way or another depend on PK activity. Furthermore, abnormal PK activity has been related to a host of disorders, ranging from relatively non-life threatening diseases such as psoriasis to extremely virulent diseases such as glioblastoma.

The kinases fall largely into two groups, those specific for phosphorylating serine and threonine, and those specific for phosphorylating tyrosine. Some kinases, referred to as "dual specificity" kinases, are able to phosphorylate tyrosine as well as serine/threonine residues.

Protein kinases can also be characterized by their location within the cell. Some kinases are transmembrane receptor proteins capable of binding ligands external to the cell membrane. Binding the ligands alters the receptor protein kinase's catalytic activity. Others are non-receptor proteins lacking a transmembrane domain and yet others are ecto-kinases that have a catalytic domain on the extracellular (ecto) portion of a transmembrane protein or which are secreted as soluble extracellular proteins.

Many kinases are involved in regulatory cascades where their substrates may include other kinases whose activities are regulated by their phosphorylation state. Thus, activity of a downstream effector is modulated by phosphorylation resulting from activation of the pathway.

Receptor protein tyrosine kinases (RPTKs) are a subclass of transmembrane-spanning receptors endowed with intrinsic, ligand-stimulatable tyrosine kinase activity. RPTK activity is tightly controlled. When mutated or altered structurally, RPTKs can become potent oncoproteins, causing cellular transformation. In principle, for all RPTKs involved in cancer, oncogenic deregulation results from relief or perturbation of one or several of the autocontrol mechanisms that ensure the normal repression of catalytic domains. More than half of the known RPTKs have been repeatedly found in either mutated or overexpressed forms associated with human malignancies (including sporadic cases; Blume-Jensen et al., 2001). RPTK over expression leads to constitutive kinase activation by increasing the concentration of dimers. Examples are Neu/ErbB2 and epidermal growth factor receptor (EGFR), which are often amplified in breast and lung carcinomas and the fibroblast growth factors (FGFR) associated with skeletal and proliferative disorders (Blume-Jensen et al., 2001).

Fibroblast Growth Factors

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Normal growth, as well as tissue repair and remodeling, require specific and delicate control of activating growth factors and their receptors. Fibroblast Growth Factors (FGFs) constitute a family of over twenty structurally related polypeptides that are developmentally regulated and expressed in a wide variety of tissues. FGFs stimulate proliferation, cell migration and differentiation and play a major role in skeletal and limb development, wound healing, tissue repair, hematopoiesis, angiogenesis, and tumorigenesis (reviewed in Ornitz and Itoh, 2001).

The biological action of FGFs is mediated by specific cell surface receptors belonging to the RPTK family of protein kinases. These proteins consist of an extracellular ligand binding domain, a single transmembrane domain and an intracellular tyrosine kinase domain which undergoes phosphorylation upon binding of FGF. The FGF receptor (FGFR) extracellular region contains three immunoglobulin-like (Ig-like) loops or domains (D1, D2 and D3), an acidic box, and a heparin binding domain. Five FGFR genes that encode for multiple receptor protein variants have been identified to date.

Another major class of cell surface binding sites includes binding sites for heparan sulfate proteoglycans (HSPG) that are required for high affinity interaction and activation of all members of the FGF family. Tissue-specific expression of heparan sulfate structural variants confer ligand-receptor specificity and activity of FGFs.

FGFR-Related Disease

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Recent discoveries show that a growing number of skeletal abnormalities, including achondroplasia, the most common form of human dwarfism, result from mutations in FGFRs. Specific point mutations in different domains of FGFR3 are associated with autosomal 15 dominant human skeletal disorders including hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) and thanatophoric dysplasia (TD) (Cappellen et al., 1999; Webster et al., 1997; Tavormina et al., 1999). FGFR3 mutations have also been described in two craniosynostosis phenotypes: Muenke coronal craniosynostosis (Bellus et al., 1996; Muenke et al., 1997) and Crouzon syndrome with acanthosis nigricans 20 (Meyers et al., 1995). Crouzon syndrome is associated with specific point mutations in FGFR2 and both familial and sporadic forms of Pfeiffer syndrome are associated with mutations in FGFR1 and FGFR2 (Galvin et al., 1996; Schell et al., 1995). Mutations in FGFRs result in constitutive activation of the mutated receptors and increased receptor protein tyrosine kinase activity, rendering cells and tissue unable to differentiate. 25 Specifically, the achondroplasia mutation results in enhanced stability of the mutated receptor, dissociating receptor activation from down-regulation, leading to restrained chondrocyte maturation and bone growth inhibition (reviewed in Vajo et al., 2000).

There is accumulating evidence for mutations activating FGFR3 in various types of cancer. Constitutively activated FGFR3 in a large proportion of two common epithelial cancers, bladder and cervix, as well as in multiple myeloma, is the first evidence of an oncogenic role for FGFR3 in carcinomas. FGFR3 currently appears to be the most frequently mutated

oncogene in bladder cancer where it is mutated in almost 50% of the cases and in about 70% of cases having recurrent superficial bladder tumors (Cappellen, et al, 1999; van Rhijn, et al, 2001; Billerey, et al, 2001). FGFR3 mutations are seen in 15-20% of multiple myeloma cases where point mutations that cause constitutive activation directly contribute to tumor development and progression (Chesì, et al, 1997; Plowright, et al, 2000, Ronchetti, et al, 2001).

In this context, the consequences of FGFR3 signaling appear to be cell type-specific. In chondrocytes, FGFR3 hyperactivation results in growth inhibition (reviewed in Ornitz, 2001), whereas in the myeloma cell it contributes to tumor progression (Chesi et al., 2001).

In view of the link between RPTK-related cellular activities and a number of human disorders various strategies have been employed to target the receptors and/or their variants for therapy. Some of these have involved biomimetic approaches using large molecules patterned on those involved in the cellular processes, e.g., mutant ligands (US Patent 4,966,849); soluble receptors and antibodies (WO 94/10202, US 6,342,219); RNA ligands (US Patent 5,459,015) and tyrosine kinase inhibitors (WO 94/14808; US Patent 5,330,992).

Antibody therapy

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The search for new therapies to treat cancer and other diseases associated with growth factors and their corresponding cell surface receptors has resulted in the development of humanized antibodies capable of inhibiting receptor function. For example, US patents 5,942,602 and 6,365,157 disclose monoclonal antibodies specific for the HER2/neu and VEGF receptors, respectively. US patent 5,840,301 discloses a chimeric, humanized monoclonal antibody that binds to the extracellular domain of VEGF and neutralizes ligand-dependent activation.

There is an unmet need for highly selective molecules capable of blocking aberrant constitutive receptor protein tyrosine kinase activity, in particular FGFR activity, thereby addressing the clinical manifestations associated with the above-mentioned mutations, and modulating various biological functions.

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SUMMARY OF THE INVENTION

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It is an object of the present invention to provide molecules which are able to block receptor protein tyrosine kinase (RPTK) activity.

It is an object of the present invention to provide molecules which are able to block fibroblast growth factor receptor (FGFR) activity, and more preferably fibroblast growth factor receptor 3 (FGFR3) activity.

It is another object of the present invention to provide a method to screen for molecules which are able to block said receptor activity.

It is yet another object of the present invention to provide a pharmaceutical composition comprising as an active ingredient a molecule of the invention useful in treating or preventing skeletal and proliferative diseases and disorders.

It is a further object of the present invention to provide a method for inhibiting growth of tumor cells associated with ligand-dependent or constitutive activation of a receptor protein tyrosine kinase, preferably a fibroblast growth factor receptor, and more preferably FGFR3.

It is yet a further object of the present invention to provide a method for treating skeletal disorders associated with ligand-dependent or constitutive activation of a receptor protein tyrosine kinase, preferably a fibroblast growth factor receptor, and more preferably FGFR3.

It is yet a further object of the present invention to provide a method for blocking receptor protein tyrosine kinase activation in the cells of patients in need thereof by treatment with molecules capable of inhibiting receptor protein tyrosine kinase function.

It is yet another object of the present invention to provide a method for inhibiting tumor growth, tumor progression or metastases.

It is still a further object to provide molecules useful for in vivo imaging of diseased states.

It is still a further object of the invention to provide a kit containing molecules of the invention.

These and other objects are met by the invention disclosed herein.

The present invention provides a molecule that contains the antigen-binding portion of an antibody which has a specific affinity for a receptor protein tyrosine kinase and which blocks constitutive activation of a receptor protein tyrosine kinase. The present invention further

provides a molecule that contains the antigen-binding portion of an antibody which has a specific affinity for a receptor protein tyrosine kinase and which blocks ligand-dependent activation of a fibroblast growth factor receptor (FGFR), including FGFR1 and FGFR3.

Certain molecules of the present invention were found to inhibit or block constitutive, or ligand independent, activation of the FGFR3. Generation of inhibitory molecules would be useful for developing medicaments for use in treating or preventing skeletal and proliferative diseases and disorders associated with constitutive activation of receptor protein tyrosine kinases.

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Certain mutations in the genes of receptor protein tyrosine kinases result in activation of the receptor in a manner that is independent of the presence of a ligand. Such ligand-independent, or constitutive, receptor protein tyrosine kinase activation results in increased receptor activity. The clinical manifestations of certain mutations are skeletal and proliferative disorders and diseases, including achondroplasia and various cancers.

Furthermore, the present invention is directed to novel molecules comprising an antigen binding domain which binds to a receptor protein tyrosine kinase and blocks constitutive activation of said receptor protein tyrosine kinase. The molecules of the invention maybe antibodies or antigen binding fragments thereof.

A currently preferred embodiment of the present invention provides a molecule which binds to the extracellular domain of a receptor protein tyrosine kinase and blocks constitutive and ligand-dependent activation of the receptor.

A currently more preferred embodiment of the present invention provides a molecule which binds to the extracellular domain of an FGF receptor and blocks constitutive and ligand-dependent activation of the receptor.

A currently most preferred embodiment of the present invention provides a molecule which binds FGFR3 and blocks constitutive and ligand-dependent activation of the receptor, comprising V_L-CDR3 and V_H-CDR3 regions having SEQ ID NO:25 and 24, respectively and the corresponding polynucleotide sequence SEQ ID NO:51 and 50.

A currently most preferred embodiment of the present invention provides a molecule which binds FGFR3 and blocks constitutive and ligand-dependent activation of the receptor, comprising V_L-CDR3 and V_H-CDR3 regions having SEQ ID NO:13 and 12 or SEQ ID NO:9

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and 8, respectively and the corresponding polynucleotide sequence SEQ ID NO:35 and 34 or SEQ ID NO: 31 and 30.

Another currently preferred embodiment of the present invention provides a molecule herein denoted MSPRO12 comprising a light chain having SEQ ID NO:94 and a heavy chain having SEQ ID NO:105 and the corresponding polynucleotide sequences having SEQ ID NO:75 and 89, respectively.

Another currently preferred embodiment of the present invention provides a molecule herein denoted MSPRO2 comprising a light chain having SEQ ID NO:92 and a heavy chain having SEQ ID NO:103 and the corresponding polynucleotide sequences having SEQ ID NO:74 and 86,

Another currently most preferred embodiment of the present invention provides a molecule herein denoted MSPRO59 comprising a light chain having SEQ ID NO:102 and a heavy chain having SEQ ID NO:113 and the corresponding polynucleotide sequences having SEQ ID NO:76 and 91, respectively.

According to the principles of the present invention, molecules which bind FGFR and block ligand-dependent receptor activation are provided. These molecules are useful in treating disorders and diseases associated with an FGFR that is activated in a ligand-dependent manner including certain skeletal disorders, hyperproliferative diseases or disorders and non-neoplastic angiogenic pathologic conditions such as neovascular glaucoma, macular

degeneration, hemangiomas, angiofibromas, psoriasis and proliferative retinopathy including proliferative diabetic retinopathy.

A currently most preferred embodiment of the present invention provides a molecule which binds FGFR3 and blocks ligand-dependent activation of the receptor, comprising V_H -CDR3 and V_L -CDR3 regions having SEQ ID NO:20 and 21, respectively and the corresponding polynucleotide sequence SEQ ID NO:44 and 45, respectively.

Other currently preferred embodiments of the present invention provides a molecule which binds FGFR3 and blocks ligand-dependent activation of the receptor, comprising V_H -CDR3 and V_L -CDR3 regions selected from the group consisting of SEQ ID NO:10 and SEQ ID NO:11; SEQ ID NO:14 and SEQ ID NO:15; SEQ ID NO:16 and SEQ ID NO:17; SEQ ID

NO:18 and SEQ ID NO:19; SEQ ID NO:20 and SEQ ID NO:21; SEQ ID NO:26 and SEQ ID NO:27 or SEQ ID NO:28 and SEQ ID NO:29 and the corresponding polynucleotide sequences according to table 1B.

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Additional currently preferred embodiments of the present invention provide molecules having an antigen binding domain comprising a VL region and a VH region, respectively, selected from the group consisting of respectively, selected from the group consisting of SEQ ID NO: 92 and 103; SEQ ID NO: 93 and 104; SEQ ID NO: 94 and 105; SEQ ID NO:95 and 106; SEQ ID NO: 96 and 107; SEQ ID NO: 97 and 108; SEQ ID NO:98 and 109; SEQ ID NO: 99 and 110; SEQ ID NO: 101 and 112; and SEQ ID NO:102 and 113.

A currently preferred embodiment of the present invention provides a molecule comprising V_{H} -CDR3 and V_{L} -CDR3 domains having SEQ ID NO:22 and SEQ ID NO:23, which has specific affinity for FGFR1 and which blocks ligand-dependent activation of FGFR1, and the corresponding polynucleotides having SEQ ID NO:46 and SEQ ID NO:47.

A currently preferred embodiment of the present invention provides a molecule comprising domains having SEQ ID NO: 100 and 111, which has specific affinity for FGFR1 and which blocks ligand-dependent activation of FGFR1, and the corresponding polynucleotides having SEQ ID NO:73 and SEQ ID NO:82.

In addition, the present invention also relates to methods for screening for the molecules according to the present invention by using a dimeric form of a receptor protein tyrosine kinase as a target for screening a library of antibody fragments.

According to one currently preferred embodiment the screening method comprises

screening a library of antibody fragments for binding to a dimeric form of a receptor protein tyrosine kinase;

identifying an antibody fragment which binds to the dimeric form of the receptor protein tyrosine kinase as a candidate molecule for blocking constitutive activation of the receptor protein tyrosine kinase; and

determining whether the candidate molecule blocks constitutive or ligand-dependent activation of the receptor protein tyrosine kinase in a cell.

According to another currently preferred embodiment the dimeric form of the RPTK is a soluble extracellular domain of a receptor protein tyrosine kinase. Non-limiting examples of receptor protein tyrosine kinases disclosed in Blume-Jensen et al. (2001) include EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR-α, PDGFR-β, CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1,

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FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROS, Alk, Ryk, DDR, LTK and MUSK.

By using a dimeric form of the RPTK as bait in the screen, a molecule which would bind to the dimeric form of the receptor has been identified. This presents a novel concept in screening for antibodies or fragments thereof with the capacity to bind to a constitutively activated RPTK such as those associated with various disorders and diseases. It also presents an opportunity to screen for molecules which bind to a heterodimer RPTK. A further aspect of the present invention provides a pharmaceutical composition comprising as an active ingredient a molecule of the present invention useful for preventing or treating skeletal or cartilage diseases or disorders and craniosynostosis disorders associated with constitutive or ligand-dependent activation of a receptor protein tyrosine kinase.

In a currently preferred embodiment the pharmaceutical compositions of the present invention may be used for treating or preventing skeletal disorders associated with aberrant FGFR signaling, including achondroplasia, thanatophoric dysplasia, Apert or Pfeiffer syndrome and related craniosynostosis disorders.

A further aspect of the present invention provides a pharmaceutical composition comprising as an active ingredient a molecule of the present invention useful for preventing or treating cell proliferative diseases or disorders or tumor progression, associated with the constitutive or ligand-dependent activation of a receptor protein tyrosine kinase.

- In a currently preferred embodiment the pharmaceutical compositions of the present invention may be used for treating or preventing proliferative diseases associated with aberrant FGFR signaling, including multiple myeloma, transitional cell carcinoma of the bladder, mammary and cervical carcinoma, chronic myeloid leukemia and osteo- or chondrosarcoma.
- A further aspect of the invention provides molecules comprising an antigen binding domain which can be conjugated to cytotoxins useful for targeting cells expressing said antigen.
 - Another currently preferred aspect of the present invention provides molecules comprising an antigen binding domain which can be conjugated to appropriate detectable imaging moiety, useful for in vivo tumor imaging.
- A still further aspect of the present invention provides methods for treating or inhibiting the aforementioned diseases and disorders by administering a therapeutically effective amount of

a pharmaceutical composition comprising a molecule of the present invention to a subject in need thereof.

BRIEF DESCRIPTION OF THE DRAWINGS

- Figure 1 shows hFR3²³⁻³⁷⁴TDhis purification by Coommassie stained SDS-PAGE.
- Figure 2 shows hFR3²³⁻³⁷⁴TDhis binding to heparin and FGF9.
- Figure 3 shows the purification of FR3exFc and FR1exFc on SDS-PAGE.
- Figure 4 shows the neutralization effect of the hFR3²³⁻³⁷⁴TDhis and FR3exFc soluble receptors in a ligand-dependent proliferation assay.
 - Figure 5 shows the effect of MSPRO Fabs on proliferation of FGFR1 and FGFR3-expressing cells.
 - Figure 6 shows the effect of MSPRO Fabs on proliferation of FGFR3-expressing cells.
- Figures 7A and 7B show the neutralizing activity of several MSPRO Fabs in a proliferation assay using the FDCP-FR3 (C10; Fig. 7A) or the FDCP-FR1 cells (Fig. 7B).
 - Figure 8 shows the receptor specificity of MSPRO Fabs on RCJ-FR3 cells by Western blot using an anti-P-ERK antibody. Figure 8A shows different MSPRO Fabs while Figure 8B shows a dose response of MSPRO 12, 29 and 13 on RCJ-FR3 cells.
- Figures 9A-9D demonstrates the specificity and potency of MS-PRO Fabs by Western blot with anti-P-ERK antibody.
 - Figure 10 shows a diagrammatic representation of FGFR3 and of FGFR3 truncations (D2-3, D2) and isoforms (IIIb, IIIc). The isoform IIIb differs from IIIc at the carboxy terminus of the IgIII domain as indicated with a dotted line.
- Figure 11 shows that the FGFR3 neutralizing Fabs recognize IgII or IgII and III in the extracellular region of FGFR3.
 - Figure 12 shows that MSPRO29 specifically recognizes the IIIc isoform of FGFR3.
 - Figure 13 shows the results of a proliferation assay for FDCP-FR3IIIb or FDCP-FR3IIIc cells incubated with increasing dose of the indicated Fabs.
- 25 Figure 14 shows iodinated MSPRO29 binding to FGFR3.
 - Figure 15 shows results of a proliferation assay is a graph wherein iodinated MSPRO29 retained its activity against FGFR3.

Figures 16A-16F show the selective binding of radiolabelled MS-PRO29 to histological of growth plate.

- Figure 17 shows a proliferation assay of FDCP-FR3 (C10) and FDCP-FR3ach cells incubated with FGF9 and with increasing doses of the indicated Fabs.
- Figure 18B shows that MSPRO12 and MSPRO59 inhibit the ligand independent proliferation of FDCP-FR3ach cells. Fig. 18A shows analysis of the ligand-dependent FDCP-FR3wt cells.
 - Figure 19 shows the restoration of cell growth in RCS cells by MS-PRO54 and MSPRO59..
 - Figure 20 represents the growth rate of treated bone with MS-PRO 59.
- Figure 21 is a flow chart of the experimental protocol for assessing receptor activation and signaling.
 - Figure 22 shows ¹²⁵I labeled MSPRO59 localization to the FDCP-FR3ach derived tumor.
 - Figure 23 shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth after 24 and 26 days.
 - Figure 24 shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth.
- Figure 25A shows the effect of MSPRO59 on inhibiting ligand-independent tumor growth. Figure 25B shows scFv MSPRO59 blocking the proliferation of FDCP-FR3 (S375C) cells. Figure 26 shows the effect of MSPRO59 single chain antibody on inhibiting ligand-independent tumor growth.
 - Figure 27 shows binding of Fab Miniantibodies to FGFR3-Fc and FGFR1-Fc (ELISA).
- Figure 28A is an example of a Fab expression vector for use in accordance with the present invention.
 - Figure 28B is the DNA sequence of the vector according to Figure 28A
 - Figure 29A is an example of a phage display vector for use in accordance with the present invention.
- Figure 29B is the DNA sequence of the vector according to Figure 29A.
 - Figure 30 depicts the polynucleotide sequences of the VL and VH of MSPRO antibodies of the present invention.

DETAILED DESCRIPTION OF THE INVENTION

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The present invention is based on the discovery that neutralizing antibodies that block ligand-dependent and ligand-independent activation of fibroblast growth factor 3 (FGFR3), a receptor protein tyrosine kinase (RPTK), can be obtained by screening an antibody library against a dimeric form of the extracellular portion of FGFR3. Until the present invention, the present inventors are unaware of any success in obtaining neutralizing antibodies that block constitutive activation of any RPTK including FGFR or ligand-dependent FGFR activation.

The term "receptor protein tyrosine kinase" or "RPTK" as used herein and in the claims refers to a subclass of transmembrane-spanning receptors endowed with intrinsic, ligand-stimulatable tyrosine kinase activity. RPTKs comprise a large family of spatially and temporally regulated proteins that control many different aspects of growth and development. When mutated or altered structurally, RPTKs can undergo deregulation and become activated in a ligand-independent, or constitutive, manner. In certain cases they become potent oncoproteins, causing cellular transformation.

As used herein and in the claims the term "fibroblast growth factor receptor" or "FGFR" denotes a receptor specific for FGF which is necessary for transducing the signal exerted by FGF to the cell interior, typically comprising an extracellular ligand-binding domain, a single transmembrane helix, and a cytoplasmic domain having tyrosine kinase activity. The FGFR extracellular domain consists of three immunoglobulin-like (Ig-like) domains (D1, D2 and D3), a heparin binding domain and an acidic box. Alternative splicing of the FGF receptor mRNAs generates different variants of the receptors.

Molecules, including antibodies and fragments thereof, comprising an antigen binding domain to a receptor protein tyrosine kinase are highly necessary for the treatment of various pathological conditions.

In the past, the laboratory of the present inventors encountered difficulties in raising neutralizing antibodies against FGFR3. When mice were immunized with the soluble monomeric FGFR3 receptor, by the time the antibody titers begins to increase, the mice died. The experiments performed in the laboratory of the present inventors that failed to obtain anti-FGFR3 neutralizing antibodies in mice are presented in the Examples.

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By using a soluble dimeric form of the extracellular domain of the FGFR3 receptor to screen for antibodies, e.g., Fabs, that bind from a phage display antibody library, the present inventors were able to overcome a problem in the prior art for which there was yet no solution and to obtain numerous high affinity (K_D < 10 nM) antibodies (Fabs) that bind FGFR3 and interfere with ligand binding, thereby blocking ligand-dependent activation of FGFR3 signaling. Very surprisingly, from among the group of Fabs that block liganddependent activation, Fab antibodies which also block ligand-independent (constitutive) activation of FGFR3 by blocking signaling caused by constitutive dimerization of FGFR3 were identified. To the best of the present inventors' knowledge, the Fab antibodies obtained which block constitutive activation of FGFR3 are the first antibodies against any receptor protein tyrosine kinase that blocks constitutive, ligand-independent activation/signaling. Trastuzamab, an anti-human epidermal growth factor receptor 2 (HER2) antibody, was the first humanized monoclonal antibody approved for therapeutic use. Its mode of action appears to be manifold, including HER2 down regulation, prevention of heterodimer formation, prevention of HER2 cleavage and others (Baselga and Albanell, 2001), US patents 5, 677171; 5772997; 6165464 and 6,399,063 disclose the anti-HER2 invention but neither teach nor suggest that the antibody blocks ligand-independent receptor activation. One aspect of the present invention is directed to neutralizing antibodies and more generally

One aspect of the present invention is directed to neutralizing antibodies and more generally to a molecule that includes the antigen binding portion of an antibody which blocks ligand-dependent activation and constitutive, ligand-independent activation of a receptor protein tyrosine kinase, preferably an FGFR and more preferably FGFR3.

Another aspect of the present invention is directed to molecules comprising an antigen binding domain which blocks ligand-dependent activation of an FGFR, more preferably FGFR3.

The molecule having the antigen-binding portion of an antibody according to the present invention can be used in a method for blocking the ligand-dependent activation and/or ligand independent (constitutive) activation of FGFR3. Preferred embodiments of such antibodies/molecules, obtained from an antibody library designated as HuCAL® (Human Combinatorial Antibody Library) clone, is presented in Table 1 with the unique VH-CDR3 and VL-CDR3 sequences given.

In addition to sequencing of the clones, a series o biochemical assays were performed to determine affinity and specificity of the molecules to the respective receptors.

TABLE 1A

HuCAL® Clone	VH-CDR3 Sequence	VL-CDR3 equence	Framework
MSPRO2	DFLGYEFDY (SEQ ID NO: 8)	QSYDYSADY (SEQ IDNO: 9)	VH1B_L3
MSPRO11	YYGSSLYHYVFGGFIDY (SEQ ID NO: 10)	QSHHFYE (SEQ ID NO: 11)	VH1B_L2
MSPRO12	YHSWYEMGYYGSTVGYMFD (SEQ ID NO: 12)	QSYDFDFA (SEQ ID NO: 13)	VH2_L3
MSPRO21	DNWFKPFSDV (SEQ ID NO: 14)	QQYDSIPY (SEQ ID NO: 15)	VH1A_k4
MSPRO24	VNHWTYTFDY (SEQ ID NO: 16)	QQMSNYPD (SEQ ID NO: 17)	VH1A_k3
MSPRO26	GYWYAYFTYINYGYFDN (SEQ ID NO: 18)	QSYDNNSDV (SEQ ID NO: 19)	VH1B_L2
MSPRO28	GGGWVSHGYYYLFDL (SEQ ID NO: 26)	FQYGSIPP (SEQ ID NO: 27)	VH1A_k1
MSPRO29	TWQYSYFYYLDGGYYFDI (SEQ ID NO: 20)	QQTNNAPV (SEQ ID NO:21)	VH1B_k3
MSPRO54	NMAYTNYQYVNMPHFDY (SEQ ID NO: 22)	QSYDYFKL (SEQ ID NO:23)	VH1B_L3
MSPRO55	SMNSTMYWYLRRVLFDH (SEQ ID NO: 28)	QSYDMYMYI (SEQ ID NO: 29)	VH1B_L2
MSPRO59	SYYPDFDY (SEQ ID NO:24)	QSYDGPDLW (SEQ ID NO:25)	VH6_L3

VH refers to the variable heavy chain, VL refers to the variable light chain; L refers to the lambda light chain and k refers to the kappa light chain

Table 1B lists the corresponding polynucleotide sequences of the CDR domains.

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TABLE 1B

HuCAL® Clone	VH-CDR3 polynucleotide sequence	VL-CDR3 olynucleotide
		Sequence
	GATTTTCTTGGTTATGAGTTTGATTAT	CAGAGCTATGAC
MSPRO2	(SEQ ID NO:30)	TATTCTGCT GAT
		TAT (SEQ ID NO:31)
	TATTATGGTTCTTCTCTTTATCATTATGTTT	CAGTCTCATCAT
MSPRO11	TTGGTGGTTTTATTGATTAT	TTTTATGAG
	(SEQ ID NO:32)	(SEQ ID NO:33)
	TATCATTCTTGGTATGAGATGGGTTATTAT	CAGAGCTATGAC
MSPRO12	GGTTCTACTGTTGGTTATATGTTTGATTAT	TTTGATTTT GCT
	(SEQ ID NO:34)	(SEQ ID NO:35)
		CAGCAGTATGAT
MSPRO21	GATAATTGGTTTAAGCCTTTTTCTGATGTT	TCTATTCCT TAT
	(SEQ ID NO:36)	(SEQ ID NO:37)
		CAGCAGATGTCT
MSPRO24	GTTAATCATTGGACTTATACTTTTGATTAT	AATTATCCTGAT
i	(SEQ ID NO:38)	(SEQ ID NO:39)
	GGTTATTGGTATGCTTATTTTACTTAT	CAGAGCTATGAC
MSPRO26	ATTAATTATGGTTATTTT GATAAT	AATAATTCTGAT
	(SEQ ID NO:40)	GTT (SEQ ID NO:41)
	GGTGGTGGTTGGGTTTCTCATGGTTATTAT	TTTCAGTATGGT
MSPRO28	TATCTTTTGATCTT	TCTATTCCT CCT
	(SEQ ID NO:42)	(SEQ ID NO: 43)
	ACTTGGCAGTATTCTTATTTTTATTAT	CAGCAGACTAAT
MSPRO29	CTTGATGGTGGTTATTATTTTGATATT	AATGCTCCTGTT
	(SEQ ID NO:44)	(SEQ ID NO:45)
	AATATGGCTTATACTAATTATCAGTATGTT	CAGAGCTATGAC
MSPRO54	AATATGCCTCATTTTGATTAT	TATTTTAAGCTT
	(SEQ ID NO:46)	(SEQ ID NO:47)
MSPRO55	TCTATGAATTCTACTATGTATTGGTATCTT	CAGAGCTATGAC
	CGTCGTGTTCTTTTTGAT CAT	ATGTATAATTAT
	(SEQ ID NO:48)	ATT (SEQ ID NO:49)
		CAGAGCTATGAC
MSPRO59	TCTTATTAT CCTGATTTT GATTAT	GGTCCTGATCTT
	(SEQ ID NO:50)	TGG (SEQ ID NO:51)

Figure 30 provides the polynucleotide sequences of the preferred embodiments of the invention. The amino acid sequence of the VH and VL domains of the currently preferred embodiments of the present invention are presented below.

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     MS-Pro-2-VL(SEQ ID NO:92)
               DIELTQPPSV SVAPGQTARI SCSGDALGDK YASWYQQKPG QAPVLVIYDD
          1
       51
               SDRPSGIPER FSGSNSGNTA TLTISGTQAE DEADYYCQSY DYSADYVFGG
10
       101
               GTKLTVLGQ
     MS-Pro-11-VL(SEO ID NO:93)
               DIALTQPASV SGSPGQSITI SCTGTSSDVG GYNYVSWYQQ HPGKAPKLMI
15
       51
               YDVSNRPSGV SNRFSGSKSG NTASLTISGL QAEDEADYYC QSHHFYEVFG
       101
               GGTKLTVLGQ
20
     MS-PRO-12-VL (SEQ ID NO:94)
               DIELTQPPSV SVAPGQTARI SCSGDALGDK YASWYQQKPG QAPVLVIYDD
       51
               SDRPSGIPER FSGSNSGNTA TLTISGTQAE DEADYYCQSY DFDFAVFGGG
25
       101
               TKLTVLGO
     MS-Pro-21-VL(SEQ ID NO:95)
               DIVMTQSPDS LAVSLGERAT INCRS SQSVL YSSNNKNYLA WYQQKPGQPP
30
       51
               KLLIYWASTR ESGVPDRFSG SGSGTDFTLT ISSLQAEDVA VYYCQQYDSI
       101
               PYTFGQGTKV EIKRT
     MS-Pro-24-VL(SEQ ID NO:96)
35
               DIVLTQSPAT LSLS PGERAT LSCRA SQSVS SSYLAWYQQK PGQAPRLLIY
       51
               GASSRATGVP ARFSGSGSGT DFTLTISSLE PEDFATYYCQ QMSNYPDTFG
       101
               QGTKVEIKRT
40
     MS-Pro-26-VL (SEQ ID NO:97)
               DIALTQPASV SGSPGQSITI SCTGTSSDVG GYNYVSWYQQ HPGKAPKLMI
       51
               YDVSNRPSGV SNRFSGSKSG NTASLTISGL QAEDEADYYC QSYDNNSDVV
45
       101
               FGGGTKLTVL GQ
     MS-Pro-28-VL (SEQ ID NO:98)
50
       1
               DIQMTQSPSS LSASVGDRVT ITCRASQGIS SYLAWYQQKP GKAPKLLIYA
       51
               ASSLQSGVPS RFSGSGSGTD FTLTISSLQP EDFAVYYCFQ YGSIPPTFGQ
       101
               GTKVEIKRT
55
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- 17 **-**

	MS-Pro-29-	-VL (SEQ ID DIVLTQSPAT	•	LSCRA SQSVS	SSYLAW YQQK	PGQAPRLLIY
5	51	GASSRATGVP	ARFS GSGSGT	DFTLT ISSLE	PEDFAT YYCQ	QTNNAPV TFG
	101	QGTKVEIKRT				
10	MS-Pro-54-	-VL (SEQ ID	NO:100)			
10	1	DIELTQPPSV	SVAP GQTARI	SCSGD ALGDK	YASWYQ QKPG	QAPVLVI YDD
	51	SDRPSGIPER	FSGS NSGNTA	TLTIS GTQAE	DEADYY CQSY	DYFKLVF GGG
15	101	TKLTVLGQ				
	MS-Pro-55-	-VL (SEQ ID DIALTQPASV	•	SCTGT SSDVG	GYNYVS WYQQ	HPGKAPK LMI
20	51	YDVSNRPSGV	SNRF SGSKSG	NTASL TISGL	QAEDEA DYYC	QSYDMYNYIV
	101	FGGGTKLTVL	GQ			
25	MS-Pro-59-	-VL (SEQ ID DIELTQPPSV	•	SCSGD ALGDK	YASWYQ QKPG	QAPVLVI YDD
	51	SDRPSGIPER	FSGS NSGNTA	TLTIS GTQAE	DEADYY CQSY	DGPDLWV FGG
30	101	GTKLTVLGQ				
	MS-Pro-2-V 1	/H (SEQ ID 1 QVQLVQSGAE	•	SCKAS GYTFT	SYYMHW VRQA	PGQGLEWMGW
35	51	INPNSGGTNY	AQKF QGRVTM	TRDTS ISTAY	MELSSLR SED	TAVYYCAR DF
	101	LGYEFDYWGQ	GTLV TVSS			
40	MS-Pro-11	-VH (SEQ ID QVQLVQSGAE	•	SCKAS GYTFT	SYYMHW VRQA	PGQGLEWMGW
	51	INPNSGGTNY	AQKF QGRVTM	TRDTS ISTAY	MELSSL RSED	TAVYYCARYY
45	101	GSSLYHYVFG	GFID YWGQGT	LVTVSS		
	MS-Pro-12	-VH(SEQ ID I QVQLKESGPA		TCTFS GFSLS	TSGVGV GWIR	QPPGKAL EWL
50	51	ALIDWDDDKY	YSTS LKTRLT	ISKDT SKNQV	VLTMTN MDPV	DTATYYCARY
50	101	HSWYEMGYYG	STVG YMFDYW	GQGTL VTVSS		
55	MS-Pro-21	-VH (SEQ ID QVQLVQSGAE	•	SCKAS GGTFS	SYAISWVRQA	PGQGLEWMGG
<u> </u>	51	IIPIFGTANY	AQKF QGRVTI	TADES TSTAY	MELSSL RSED	TAVYYCA RDN
	101	WFKPFSDVWG	QGTLVTVSS			
60						

	MS-Pro-24 1	-VH (SEQ ID QVQLVQSGAE		SCKAS GGTFS	SYAISW VRQA	PGQGLEWMGG
5	51	IIPIFGTANY	AQKFQ GRVTI	TADEST STAY	MELSSLR SED	TAVYYCAR VN
	101	HWTYTFDYWG	QGTL VTVSS			
	MS_Dro-26-	·VH(SEQ ID 1	viO+10.8)			
10	1		-	SCKAS GYTFT	SYYMHW VRQA	PGQGLEWMGW
	51	INPNSGGTNY	AQKF QGRVTM	TRDTS ISTAY	MELSSL RSED	TAVYYCARGY
15	101	WYAYFTYINY	GYFD NWGQGT	LVTVS S		
-L-V	MS-Pro-28-	-VH(SEQ ID 1	NO:109)			
	1	QVQLVQSGAE	VKKP GSSVKV	SCKAS GGTFS	SYAISW VRQA	PGQGLEW MGG
20	51	IIPIFGTANY	AQKF QGRVTI	TADES TSTAY	MELSSL RSED	TAVYYCARGG
20	101	GWVSHGYYYL	FDLW GQGTLV	TVSS		
	MS-Pro-29-	VH(SEQ ID 1	NO:110)			
25	1	QVQLVQSGAE	VKKP GASVKV	SCKAS GYTFT	SYYMHW VRQA	PGQGLEWMGW
2.7	51	INPNSGGTNY	AQKF QGRVTM	TRDTS ISTAY	MELSSL RSED	TAVYYCARTW
	101	QYSYFYYLDG	GYYF DIWGQG	TLVTVSS		
30	MS-Pro-54-	·VH(SEQ ID 1	NO:111)			
	1	QVQLVQSGAE	VKKP GASVKV	SCKAS GYTFT	SYYMHW VRQA	PGQGLEWMGW
	51	INPNSGGTNY	AQKF QGRVTM	TRDTS ISTAY	MELSSL RSED	TAVYYCA RNM
35	101	MUVYQYVIIYA	PHFD YWGQGT	LVTVSS		
	MS-Pro-55-	·VH(SEQ ID 1	NO:112)			
	1	QVQLVQSGAE	VKKP GASVKV	SCKAS GYTFT	SYYMHW VRQA	PGQGLEWMGW
40	51	INPNSGGTNY	AQKF QGRVTM	TRDTS ISTAY	MELSSL RSED	TAVYYCA RSM
	101	NSTMYWYLRR	VLFD HWGQGT	LVTVSS		
	MS-Pro-59-	VH (SEQ ID	NO:113)			
45	1	QVQLQQSGPG	LVKP SQTLSL	TCAIS GDSVS	SNSAAWNWIR	QSPGRGL EWL
	51	GRTYYRSKWY	NDYAVSVKSR	ITINP DTSKN	QFSLQL NSVT	PEDTAVY YCA
50	101	RSYYPDFDYW	GQGT LVTVSS			

In addition to sequencing of the clones, a series of biochemical assays were performed to determine affinity and specificity of the molecules to the respective receptors. Table 1C lists the affinity of the respective molecules to FGFR3 and FGFR1 as measures by Biacore and/or

FACS. In a binding assay to FGFR3-expressing cells the IC₅₀ of the molecules was

calculated (Example 6). Domain specificity was determined as described in Example 9. The ligand-independent inhibition of FGFR3 (neutralizing activity) was determined as described in Example 11. Finally, the molecules were synthesized in a number of different formats including Fab, miniantibody (Fab-dHLX), IgG1, IgG4, IgG3 and as single chain Fv (scFv).

5 Table 1C

Clone	Affinity to	Affinity	Affinity	Koff	IC50	Domain	Ligand	Available
	FGFR3	to	to	(s ⁻¹)	FR3	Specificity	independent	formats
	(BIAcore)	FGFR3	FGFR1		(FGF9)		inhibition of	
	1	(FACS)					FGFR3	
MSPRO59	1.5nM	<1nM	-	7.1x10e-4	19 nM	2	+	Fab, Fab-dHLX
	,	i					!	IgG1, IgG4,
								mIgG3, scFv
MSPRO2	37nM	43 nM	-	2x10e-2	360 nM	2	~	Fab(+/- tags),
			!					Fab-dHLX,
			!					IgG1, IgG4,
MSPRO12	14nM	6.5 nM	-	2.3x10e-3	58 nM	2	+	Fab (+/- tag),
							į	Fab-dHLX,
								IgG1, IgG4, scFv
MSPRO11	4	4 nM	108	4 x 10e-4	220 nM	3		Fab. Fab-dHLX
MSPRO21	9 nM		-	3.6x10e-3	50 nM	3c		Fab, IgG1,
								Fab-dHLX
MSPRO24			-	5.4x10e-3	<u> </u>	3c		Fab, IgG1
	10 nM	ĺ	į	1	70 nM		Į.	1
MSPRO26	4 nM	1.4		5 x 10e-4	70 nM	3		Fab, Fab-dHLX
MSPRO28	9 nM	0.3 nM	160 nM	4 x10e-3	50 nM	3		Fab
MSPRO29	6 nM	<1nM		1.4x10e-3		3c		Fab (+/- tag), IgG1,
			29 nM		20nM			IgG4,
								Fab-dHLX, scFv
MSPRO54	3.7nM		2.5nM	2x10e-3	45nM	3c		Fab, IgG1
MSPRO55	2.9nM		-	7.4x10e-4	34nM	3c		Fab

BiaCore results for certain molecules

In Table 1D the numbers are the IC_{50} s of the dimeric dHLX format of certain binders (molecule with antigen binding site) in the FDCP-FGFR3 proliferation assay performed with FGF9. The numbers in parentheses are the IC_{50} of the monomeric Fabs in the same assay.

Table 1E presents the KD value for certain MSPRO molecules in miniantibody form, as determined in a Biacore.

Table 1D

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binder	IC ₅₀
MSPRO2	61 nM (360)
MSPRO12	26 nM (58)
MSPRO21	20 nM (50)
MSPRO26	8 nM (70)

Table 1E

10 K_D determination for certain molecules

Clone	BIAcore K _D [nM]	Number of measurements
MS-Pro-2-dHLX-MH	4.3 (37)	1
MS-Pro-11-dHLX-MH	0.7 (4)	1
MS-Pro-12-dHLX-MH	1.2 (14)	1
MS-Pro-21-dHLX-MH	2.2 (4.1)	1
MS-Pro-24-dHLX-MH	2 (10)	1
MS-Pro-26-dHLX-MH	2 (4)	1
MS-Pro-28-dHLX-MH	1.6 (9)	1

The preferred, but non-limiting, embodiments of molecules according to the present invention that block constitutive (ligand-independent) activation of FGFR3 are referred to herein MSPRO2, MSPRO12 and MSPRO59 comprising VH-CDR3 and VL-CDR3 domains having SEQ ID Nos: 8 and 9; 12 and 13; and 24 and 25, respectively. The preferred, but non-limiting, embodiments of molecules according to the present invention that block ligand-dependent activation of FGFR3 are referred to herein MSPRO11, MSPRO21, MSPRO24, MSPRO26, MSPRO29, and MSPRO54 comprising VH-CDR3 and VL-CDR3 domains

having SEQ ID Nos: 10 and 11; 14 and 15; 16 and 17, 18 and 19; 21 and 22; 23 and 24, respectively. Preferably, an antibody or a molecule of the present invention has an affinity (K_D) for binding a soluble dimeric form of FGFR3 of less than about 50 nM, preferably less than about 30 nM and more preferably less than about 10 nM, as determined by the BIAcore chip assay for affinity, by a FACS-Scatchard analysis or other methods known in the art. 5 While the specific discovery of an antibody/molecule that blocks constitutive activation was made with respect to FGFR3 using a soluble dimeric form of FGFR3 to screen a phage display antibody library, it is believed that for all, or almost all receptor protein tyrosine kinases, antibodies/molecules that block constitutive activation can be similarly obtained using a soluble dimeric form of a corresponding extracellular domain of a receptor protein 10 tyrosine kinase. Non-limiting examples of receptor protein tyrosine kinases disclosed in Blume-Jensen et al. (2001) include EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR-α, PDGFR-β, CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1. Flk1/VEGFR2, Flt4/VEGFR3, FGFR1, FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROSAlk, Ryk, DDR, 15

LTK and MUSK.

Furthermore, antibodies/molecules that block ligand-dependent or ligand independent

activation of heterodimer receptor protein tyrosine kinases are intended to be included in the scope of the invention. Heterodimerization is well documented for members of the EGFR subfamily of receptor protein tyrosine kinases and others. Non-limiting examples include EGFR/PDGFRβ, Flt1/KDR and EGFR/ErbB2 heterodimers. EGFR and PDGFRβ heterodimers have been identified as a mechanism for PDGF signal transduction in rat vascular smooth muscle cells (Saito et al., 2001) and Flt-1/KDR heterodimers are required for vinculin assembly in focal adhesions in response to VEGF signaling (Sato et al., 2000).

The present invention is also directed to a molecule having the antigen-binding portion of an antibody which binds to a dimeric form of an extracellular portion of a receptor protein tyrosine kinase (RPTK), such as a FGFR, and blocks the ligand-independent (constitutive) activation and/or ligand-dependent activation of the RPTK.

Antibodies

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Antibodies, or immunoglobulins, comprise two heavy chains linked together by disulfide bonds and two light chains, each light chain being linked to a respective heavy chain by disulfide bonds in a "Y" shaped configuration. Proteolytic digestion of an antibody yields Fv

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(Fragment variable and Fc (fragment crystalline) domains. The antigen binding domains, Fab', include regions where the polypeptide sequence varies. The term $F(ab')_2$ represents two Fab' arms linked together by disulfide bonds. The central axis of the antibody is termed the Fc fragment. Each heavy chain has at one end a variable domain (VH) followed by a number of constant domains (CH). Each light chain has a variable domain (VL) at one end and a constant domain (CL) at its other end, the light chain variable domain being aligned with the variable domain of the heavy chain and the light chain constant domain being aligned with the first constant domain of the heavy chain (CH1).

The variable domains of each pair of light and heavy chains form the antigen binding site.

The domains on the light and heavy chains have the same general structure and each domain comprises four framework regions, whose sequences are relatively conserved, joined by three hypervariable domains known as complementarity determining regions (CDR1-3). These domains contribute specificity and affinity of the antigen binding site.

The isotype of the heavy chain (gamma, alpha, delta, epsilon or mu) determines immunoglobulin class (IgG, IgA, IgD, IgE or IgM, respectively). The light chain is either of two isotypes (kappa,κ or lambda,λ) found in all antibody classes.

It should be understood that when the terms "antibody" or "antibodies" are used, this is intended to include intact antibodies, such as polyclonal antibodies or monoclonal antibodies (mAbs), as well as proteolytic fragments thereof such as the Fab or F(ab')₂ fragments. Further included within the scope of the invention are chimeric antibodies; human and humanized antibodies; recombinant and engineered antibodies, and fragments thereof. Furthermore, the DNA encoding the variable region of the antibody can be inserted into the DNA encoding other antibodies to produce chimeric antibodies (see, for example, US patent 4,816,567). Single chain antibodies fall within the scope of the present invention. Single chain antibodies can be single chain composite polypeptides having antigen binding capabilities and comprising amino acid sequences homologous or analogous to the variable regions of an immunoglobulin light and heavy chain (linked VH-VL or single chain Fv (ScFv)). Both V_H and V_L may copy natural monoclonal antibody sequences or one or both of the chains may comprise a CDR-FR construct of the type described in US patent 5,091,513, the entire contents of which are hereby incorporated herein by reference. The separate polypeptides analogous to the variable regions of the light and heavy chains are held together by a polypeptide linker. Methods of production of such single chain antibodies, particularly

where the DNA encoding the polypeptide structures of the V_H and V_L chains are known, may be accomplished in accordance with the methods described, for example, in US patents 4,946,778, 5,091,513 and 5,096,815, the entire contents of each of which are hereby incorporated herein by reference.

Additionally, CDR grafting may be performed to alter certain properties of the antibody molecule including affinity or specificity. A non-limiting example of CDR grafting is disclosed in US patent 5,225,539.

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A "molecule having the antigen-binding portion of an antibody" as used herein is intended to include not only intact immunoglobulin molecules of any isotype and generated by any animal cell line or microorganism, but also the antigen-binding reactive fraction thereof, including, but not limited to, the Fab fragment, the Fab' fragment, the F(ab')₂ fragment, the variable portion of the heavy and/or light chains thereof, Fab miniantibodies (see WO 93/15210, US patent application 08/256,790, WO 96/13583, US patent application 08/817,788, WO 96/37621, US patent application 08/999,554, the entire contents of which are incorporated herein by reference) and chimeric or single-chain antibodies incorporating such reactive fraction, as well as any other type of molecule or cell in which such antibody reactive fraction has been physically inserted, such as a chimeric T-cell receptor or a T-cell having such a receptor, or molecules developed to deliver therapeutic moieties by means of a portion of the molecule containing such a reactive fraction. Such molecules may be provided by any known technique, including, but not limited to, enzymatic cleavage, peptide synthesis or recombinant techniques.

The term "Fc" as used herein is meant as that portion of an immunoglobulin molecule (Fragment crystallizable) that mediates phagocytosis, triggers inflammation and targets Ig to particular tissues; the Fc portion is also important in complement activation.

In one embodiment of the invention, a chimera comprising a fusion of the extracellular domain of the RPTK and an immunoglobulin constant domain can be constructed useful for assaying for ligands for the receptor and for screening for antibodies and fragments thereof The "extracellular domain" when used herein refers the polypeptide sequence of the RPTKs disclosed herein which are normally positioned to the outside of the cell. The extracellular domain encompasses polypeptide sequences in which part of or all of the adjacent (Cterminal) hydrophobic transmembrane and intracellular sequences of the mature RPTK have

been deleted. Thus, the extracellular domain-containing polypeptide can comprise the extracellular domain and a part of the transmembrane domain. Alternatively, in the preferred embodiment, the polypeptide comprises only the extracellular domain of the RPTK. The truncated extracellular domain is generally soluble. The skilled practitioner can readily determine the extracellular and transmembrane domains of a RPTK by aligning the RPTK of interest with known RPTK amino acid sequences for which these domains have been delineated. Alternatively, the hydrophobic transmembrane domain can be readily delineated based on a hydrophobicity plot of the polypeptide sequence. The extracellular domain is N-terminal to the transmembrane domain.

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- The term "epitope" is meant to refer to that portion of any molecule capable of being bound by an antibody or a fragment thereof which can also be recognized by that antibody. Epitopes or antigenic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and have specific three-dimensional structural characteristics as well as specific charge characteristics.
- An "antigen" is a molecule or a portion of a molecule capable of being bound by an antibody which is additionally capable of inducing an animal to produce antibody capable of binding to an epitope of that antigen. An antigen may have one or more than one epitope. The specific reaction referred to above is meant to indicate that the antigen will react, in a highly selective manner, with its corresponding antibody and not with the multitude of other antibodies which may be evoked by other antigens.
 - A "neutralizing antibody" as used herein refers to a molecule having an antigen binding site to a specific receptor capable of reducing or inhibiting (blocking) activity or signaling through a receptor, as determined by *in vivo* or *in vitro* assays, as per the specification.

A monoclonal antibody (mAb) is a substantially homogeneous population of antibodies to a specific antigen. MAbs may be obtained by methods known to those skilled in the art. See, for example Kohler et al (1975); US patent 4,376,110; Ausubel et al (1987-1999); Harlow et al (1988); and Colligan et al (1993), the contents of which references are incorporated entirely herein by reference. The mAbs of the present invention may be of any immunoglobulin class including IgG, IgM, IgE, IgA, and any subclass thereof. A hybridoma producing an mAb may be cultivated *in vitro* or *in vivo*. High titers of mAbs can be obtained in *in vivo* production where cells from the individual hybridomas are injected intraperitoneally into pristine-primed Balb/c mice to produce ascites fluid containing high

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concentrations of the desired mAbs. MAbs of isotype IgM or IgG may be purified from such ascites fluids, or from culture supernatants, using column chromatography methods well known to those of skill in the art.

Chimeric antibodies are molecules, the different portions of which are derived from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region. Antibodies which have variable region framework residues substantially from human antibody (termed an acceptor antibody) and complementarity determining regions substantially from a mouse antibody (termed a donor antibody) are also referred to as humanized antibodies. Chimeric antibodies are primarily used to reduce immunogenicity in application and to increase yields in production, for example, where murine mAbs have higher yields from hybridomas but higher immunogenicity in humans, such that human/murine chimeric mAbs are used. Chimeric antibodies and methods for their production are known in the art (Better et al, 1988; Cabilly et al, 1984; Harlow et al, 1988; Liu et al, 1987; Morrison et al, 1984; Boulianne et al, 1984; Neuberger et al, 1985; Sahagan et al, 1986; Sun et al, 1987; Cabilly et al; European Patent Applications 125023, 171496, 173494, 184187, 173494, PCT patent applications WO 86/01533, WO 97/02671, WO 90/07861, WO 92/22653 and US patents 5,693,762, 5,693,761, 5,585,089, 5,530,101 and 5,225,539). These references are hereby incorporated by reference.

Besides the conventional method of raising antibodies *in vivo*, antibodies can be generated *in vitro* using phage display technology. Such a production of recombinant antibodies is much faster compared to conventional antibody production and they can be generated against an enormous number of antigens. In contrast, in the conventional method, many antigens prove to be non-immunogenic or extremely toxic, and therefore cannot be used to generate antibodies in animals. Moreover, affinity maturation (i.e., increasing the affinity and specificity) of recombinant antibodies is very simple and relatively fast. Finally, large numbers of different antibodies against a specific antigen can be generated in one selection procedure. To generate recombinant monoclonal antibodies one can use various methods all based on phage display libraries to generate a large pool of antibodies with different antigen recognition sites. Such a library can be made in several ways: One can generate a synthetic repertoire by cloning synthetic CDR3 regions in a pool of heavy chain germline genes and thus generating a large antibody repertoire, from which recombinant antibody fragments with

various specificities can be selected. One can use the lymphocyte pool of humans as starting material for the construction of an antibody library. It is possible to construct naive repertoires of human IgM antibodies and thus create a human library of large diversity. This method has been widely used successfully to select a large number of antibodies against different antigens. Protocols for bacteriophage library construction and selection of recombinant antibodies are provided in the well-known reference text Current Protocols in Immunology, Colligan et al (Eds.), John Wiley & Sons, Inc. (1992-2000), Chapter 17, Section 17.1.

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Another aspect of the present invention is directed to a method for screening for the antibody or molecule of the present invention by screening a library of antibody fragments displayed on the surface of bacteriophage, such as disclosed in the Example herein and described in WO 97/08320, US Patent 6,300,064, and Knappik et al. (2000), for binding to a soluble dimeric form of a receptor protein tyrosine kinase. An antibody fragment which binds to the soluble dimeric form of the RPTK used for screening is identified as a candidate molecule for blocking ligand-dependent activation and/or constitutive activation of the RPTK in a cell. Preferably the RPTK of which a soluble dimeric form is used in the screening method is a fibroblast growth factor receptor (FGFR), and most preferably FGFR3.

As a first screening method, the soluble dimeric form of a receptor tyrosine kinase can be constructed and prepared in a number of different ways. For instance, the extracellular domain of a RPTK joined to Fc and expressed as a fusion polypeptide that dimerizes naturally by means of the Fc portion of the RPTK-Fc fusion. Other suitable types of constructs of FGFR3, serving as guidance for other RPTKs, are disclosed in the Examples presented herein.

The assays for determining binding of antibody fragments to FGFR3, binding affinities, inhibition of cell proliferation, etc., are also described in the Examples herein below.

The term "cell proliferation" refers to the rate at which a group of cells divides. The number of cells growing in a vessel can be quantified by a person skilled in the art when that person visually counts the number of cells in a defined volume using a common light microscope. Alternatively, cell proliferation rates can be quantified by laboratory apparati that optically or conductively measure the density of cells in an appropriate medium.

A second screen for antibody fragments as candidate molecules can be done using cells having very high over expression of the RPTK, such as for instance RCJ-M15 cells

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overexpressing mutant (achondroplasia) FGFR3. In cells expressing very high levels of receptor some ligand-independent activation occurs even without the presence of a mutation, such as a constitutive activation mutation. It is believed that RPTK overexpression forces RPTKs to dimerize and signal even in the absence of ligand. These cells have monomeric receptors as well as dimeric receptors present on their cell surface. Using this type of cell, one of skill in the art would be able to identify all different kinds of antibodies, i.e., blocking ligand-dependent activation, blocking constitutive activation, blocking activation and binding only to monomeric form, etc.

A third screen can be carried out on a cell line expressing a RPTK carrying a mutation, such as the FDCP-FR3ach line expressing the FGFR3 achondroplasia mutation. The receptors of this line become constitutively active, e.g. are able to signal in the absence of a ligand as determined by ERK (MAPK) phosphorylation, a downstream effector.

A further aspect of the present invention relates to a method for treating or inhibiting a skeletal dysplasia or craniosynostosis disorder associated with constitutive activation of a RPTK which involves administering the molecule of the present invention to a subject need thereof. Non-limiting examples of skeletal dysplasias include achondroplasia, thanatophoric dysplasia (TDI or TDII), hypochondroplasia, and severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia. Non-limiting examples of craniosynostosis disorder are Muenke coronal craniosynostosis and Crouzon syndrome with acanthosis nigricans. The symptoms and etiology of these diseases and disorders are reviewed in Vajo et al. (Vajo et al. 2000).

The present invention also provides for a method for treating or inhibiting a cell proliferative disease or disorder associated with the action of an abnormal constitutively activated RPTK, for example, tumor formation, primary tumors, tumor progression or tumor metastasis. A molecule containing the antigen binding portion of an antibody that blocks constitutive activation of a RPTK is administered to a subject in need thereof to treat or inhibit such a cell proliferative disease or disorder.

The terms "treating or inhibiting a proliferative disease or disorder" or "treating or inhibiting a tumor" are used herein and in the claims to encompass tumor formation, primary tumors, tumor progression or tumor metastasis.

Tumor formation or tumor growth are intended to encompass solid and non-solid tumors. Solid tumors include mammary, ovarian, prostate, colon, cervical, gastric, esophageal,

papillary thyroid, pancreatic, bladder, colorectal, melanoma, small-cell lung and non-small-cell lung cancers, granulose cell carcinoma, transitional cell carcinoma, vascular tumors, all types of sarcomas, e.g. osteosarcoma, chondrosarcoma, Kaposi's sarcoma, myosarcoma, hemangiosarcoma, and glioblastomas.

Non-solid tumors include for example hematopoietic malignancies such as all types of leukemia, e.g. chronic myelogenous leukemia (CML), acute myelogenous leukemia (AML), mast cell leukemia, chronic lymphocytic leukemia (CLL) and acute lymphocytic leukemia (ALL), lymphomas, and multiple myeloma (MM).

Tumor progression is the phenomenon whereby cancers become more aggressive with time.

Progression can occur in the course of continuous growth, or when a tumor recurs after treatment and includes progression of transitional cell carcinoma, osteo or chondrosarcoma, multiple myeloma, and mammary carcinoma (one of the known RPTKs involved in mammary carcinoma is ErbB3).

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The role of the FGFR3 in tumor progression associated with transitional cell carcinoma and multiple myeloma has recently been elucidated (Cappellen, et al, 1999; Chesi, et al, 2001) In another aspect of the present invention, molecules which bind FGFR, more preferably FGFR3, and block ligand-dependent receptor activation are provided. These molecules are useful in treating hyperproliferative diseases or disorders and non-neoplastic angiogenic pathologic conditions such as neovascular glaucoma, proliferative retinopathy including proliferative diabetic retinopathy, macular degeneration, hemangiomas, angiofibromas, and psoriasis. The role of FGFs and their receptors in neo- and hypervascularization has been well documented (Frank, 1997; Gerwins et al, 2000)

In another aspect of the present invention, the pharmaceutical compositions according to the present invention is similar to those used for passive immunization of humans with other antibodies. Typically, the molecules of the present invention comprising the antigen binding portion of an antibody will be suspended in a sterile saline solution for therapeutic uses. The pharmaceutical compositions may alternatively be formulated to control release of active ingredient (molecule comprising the antigen binding portion of an antibody) or to prolong its presence in a patient's system. Numerous suitable drug delivery systems are known and include, e.g., implantable drug release systems, hydrogels, hydroxymethylcellulose, microcapsules, liposomes, microemulsions, microspheres, and the like. Controlled release preparations can be prepared through the use of polymers to complex or adsorb the molecule

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according to the present invention. For example, biocompatible polymers include matrices of poly(ethylene-co-vinyl acetate) and matrices of a polyanhydride copolymer of a stearic acid dimer and sebaric acid (Sherwood et al, 1992). The rate of release molecule according to the present invention, i.e., of an antibody or antibody fragment, from such a matrix depends upon the molecular weight of the molecule, the amount of the molecule within the matrix, and the size of dispersed particles (Saltzman et al., 1989 and Sherwood et al., 1992). Other solid dosage forms are described in (Ansel et al., 1990 and Gennaro, 1990).

The pharmaceutical composition of this invention may be administered by any suitable means, such as orally, intranasally, subcutaneously, intramuscularly, intravenously, intra-arterially, intralesionally or parenterally. Ordinarily, intravenous (i.v.) or parenteral administration will be preferred.

It will be apparent to those of ordinary skill in the art that the therapeutically effective amount of the molecule according to the present invention will depend, *inter alia* upon the administration schedule, the unit dose of molecule administered, whether the molecule is administered in combination with other therapeutic agents, the immune status and health of the patient, the therapeutic activity of the molecule administered and the judgment of the treating physician. As used herein, a "therapeutically effective amount" refers to the amount of a molecule required to alleviate one or more symptoms associated with a disorder being treated over a period of time.

Although an appropriate dosage of a molecule of the invention varies depending on the administration route, age, body weight, sex, or conditions of the patient, and should be determined by the physician in the end, in the case of oral administration, the daily dosage can generally be between about 0.01-200 mg, preferably about 0.01-10 mg, more preferably about 0.1-10 mg, per kg body weight. In the case of parenteral administration, the daily dosage can generally be between about 0.001-100 mg, preferably about 0.001-1 mg, more preferably about 0.01-1 mg, per kg body weight. The daily dosage can be administered, for example in regimens typical of 1-4 individual administration daily. Various considerations in arriving at an effective amount are described, e.g., in Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th ed., Pergamon Press, 1990; and Remington's Pharmaceutical Sciences, 17th ed., Mack Publishing Co., Easton, Pa., 1990.

The molecule of the present invention as an active ingredient is dissolved, dispersed or admixed in an excipient that is pharmaceutically acceptable and compatible with the active

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ingredient as is well known. Suitable excipients are, for example, water, saline, phosphate buffered saline (PBS), dextrose, glycerol, ethanol, or the like and combinations thereof. Other suitable carriers are well-known to those in the art. (See, for example, Ansel et al., 1990 and Gennaro, 1990). In addition, if desired, the composition can contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents. Combination therapy

The combined treatment of one or more of the molecules of the invention with an antineoplastic or anti-chemotherapeutic drug such as doxorubicin, cisplatin or taxol provides a more efficient treatment for inhibiting the growth of tumor cells than the use of the molecule by itself. In one embodiment, the pharmaceutical composition comprises the antibody and carrier with an anti-chemotherapeutic drug.

The present invention also provides for a nucleic acid molecule, which contains a nucleotide sequence encoding the molecule having the antigen binding portion of an antibody that blocks ligand-dependent activation and/or constitutive activation of a receptor protein tyrosine kinase such as FGFR3, and a host cell transformed with this nucleic acid molecule. Furthermore, also within the scope of the present invention is a nucleic acid molecule containing a nucleotide sequence having at least 90% sequence identity, preferably about 95%, and more preferably about 97% identity to the above encoding nucleotide sequence as would well understood by those of skill in the art.

The invention also provides nucleic acids that hybridize under high stringency conditions to polynucleotides having SEQ ID NOs: 8 through 29 and SEQ ID NOs: 62, 64-65, 67, 69-71, 73-76 78-80, 82-87, 89, 90-91 or the complement thereof. As used herein, highly stringent conditions are those which are tolerant of up to about 5-20% sequence divergence, preferably about 5-10%. Without limitation, examples of highly stringent (-10°C below the calculated Tm of the hybrid) conditions use a wash solution of 0.1 X SSC (standard saline citrate) and 0.5% SDS at the appropriate Ti below the calculated Tm of the hybrid. The ultimate stringency of the conditions is primarily due to the washing conditions, particularly if the hybridization conditions used are those which allow less stable hybrids to form along with stable hybrids. The wash conditions at higher stringency then remove the less stable hybrids. A common hybridization condition that can be used with the highly stringent to moderately stringent wash conditions described above is hybridization in a solution of 6 X SSC (or 6 X SSPE), 5 X Denhardt's reagent, 0.5% SDS, 100 µg/ml denatured, fragmented salmon sperm

DNA at an appropriate incubation temperature Ti. See generally Sambrook et al., Molecular Cloning: A Laboratory Manual, 2d edition, Cold Spring Harbor Press (1989)) for suitable high stringency conditions.

Stringency conditions are a function of the temperature used in the hybridization experiment and washes, the molarity of the monovalent cations in the hybridization solution and in the wash solution(s) and the percentage of formamide in the hybridization solution. In general, sensitivity by hybridization with a probe is affected by the amount and specific activity of the probe, the amount of the target nucleic acid, the detectability of the label, the rate of hybridization, and the duration of the hybridization. The hybridization rate is maximized at a Ti (incubation temperature) of 20-25°C below Tm for DNA:DNA hybrids and 10-15°C below Tm for DNA:RNA hybrids. It is also maximized by an ionic strength of about 1.5M Na⁺. The rate is directly proportional to duplex length and inversely proportional to the degree of mismatching.

Specificity in hybridization, however, is a function of the difference in stability between the desired hybrid and "background" hybrids. Hybrid stability is a function of duplex length, base composition, ionic strength, mismatching, and destabilizing agents (if any).

The Tm of a perfect hybrid may be estimated for DNA:DNA hybrids using the equation of Meinkoth et al (1984), as

$$Tm = 81.5$$
°C + 16.6 (log M) + 0.41 (%GC) - 0.61 (% form) - 500/L

and for DNA:RNA hybrids, as

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$$Tm = 79.8$$
°C + 18.5 (log M) + 0.58 (%GC) - 11.8 (%GC)² - 0.56(% form) - 820/L

where M, molarity of monovalent cations, 0.01-0.4 M NaCl,

%GC, percentage of G and C nucleotides in DNA, 30%-75%,

% form, percentage formamide in hybridization solution, and

L, length hybrid in base pairs.

Tm is reduced by 0.5-1.5°C (an average of 1°C can be used for ease of calculation) for each 1% mismatching.

The Tm may also be determined experimentally. As increasing length of the hybrid (L) in the above equations increases the Tm and enhances stability, the full-length rat gene sequence can be used as the probe.

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Filter hybridization is typically carried out at 68°C, and at high ionic strength (e.g., 5 - 6 X SSC), which is non-stringent, and followed by one or more washes of increasing stringency, the last one being of the ultimately desired high stringency. The equations for Tm can be used to estimate the appropriate Ti for the final wash, or the Tm of the perfect duplex can be determined experimentally and Ti then adjusted accordingly.

The present invention also relates to a vector comprising the nucleic acid molecule of the present invention. The vector of the present invention may be, e.g., a plasmid, cosmid, virus, bacteriophage or another vector used e.g. conventionally in genetic engineering, and may comprise further genes such as marker genes which allow for the selection of said vector in a suitable host cell and under suitable conditions.

Furthermore, the vector of the present invention may, in addition to the nucleic acid sequences of the invention, comprise expression control elements, allowing proper expression of the coding regions in suitable hosts. Such control elements are known to the artisan and may include a promoter, a splice cassette, translation initiation codon, translation and insertion site for introducing an insert into the vector.

Preferably, the nucleic acid molecule of the invention is operatively linked to said expression control sequences allowing expression in eukaryotic or prokaryotic cells.

Control elements ensuring expression in eukaryotic or prokaryotic cells are well known to those skilled in the art. As mentioned herein above, they usually comprise regulatory sequences ensuring initiation of transcription and optionally poly-A signals ensuring termination of transcription and stabilization of the transcript.

Methods for construction of nucleic acid molecules according to the present invention, for construction of vectors comprising said nucleic acid molecules, for introduction of said vectors into appropriately chosen host cells, for causing or achieving the expression are well-known in the art (see, e.g., Sambrook et al., 1989; Ausubel et al., 1994).

The invention also provides for conservative amino acid variants of the molecules of the invention. Variants according to the invention also may be made that conserve the overall molecular structure of the encoded proteins. Given the properties of the individual amino acids comprising the disclosed protein products, some rational substitutions will be recognized by the skilled worker. Amino acid substitutions, *i.e.* "conservative substitutions," may be made, for instance, on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

For example: (a) nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; (b) polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; (c) positively charged (basic) amino acids include arginine, lysine, and histidine; and (d) negatively charged (acidic) amino acids include aspartic acid and glutamic acid. Substitutions typically may be made within groups (a)-(d). In addition, glycine and proline may be substituted for one another based on their ability to disrupt α-helices. Similarly, certain amino acids, such as alanine, cysteine, leucine, methionine, glutamic acid, glutamine, histidine and lysine are more commonly found in α helices, while valine, isoleucine, phenylalanine, tyrosine, tryptophan and threonine are more commonly found in β-pleated sheets. Glycine, serine, aspartic acid, asparagine, and proline are commonly found in turns. Some preferred substitutions may be made among the following groups: (i) S and T; (ii) P and G; and (iii) A, V, L and 1. Given the known genetic code, and recombinant and synthetic DNA techniques, the skilled scientist readily can construct DNAs encoding the conservative amino acid variants.

As used herein, "sequence identity" between two polypeptide sequences indicates the percentage of amino acids that are identical between the sequences. "Sequence similarity" indicates the percentage of amino acids that either are identical or that represent conservative amino acid substitutions.

20 Conjugates

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One embodiment of the present invention provides molecules of the present invention conjugated to cytotoxins. The cytotoxic moiety of the antibody may be a cytotoxic drug or an enzymatically active toxin or bacterial or plant origin, or an enzymatically active fragment of such a toxin including, but not limited to, diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from Pseudomonas aeruginosa), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, Aleurites fordii proteins, dianthin proteins, curcin, crotin, saponin, gelonin, mitogellin, restrictocin, phenomycin, and enomycin. In another embodiment, the molecules of the present invention are conjugate to small molecule anticancer drugs. Conjugates of the antibody and such cytotoxic moieties are made using a variety of bifunctional protein coupling agents. Examples of such reagents include SPDP, IT, bifunctional derivatives of imidoesters such a dimethyl adipimidate HCl, active esters such as disuccinimidyl suberate, aldehydes such as glutaraldehyde, bis-azido compounds such as bis-

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(p-azidobenzoyl) hexanediamine, bis-diazonium derivatives, dissocyanates and bis-active fluorine compounds. The lysing portion of a toxin may be joined to the Fab fragment of the antibodies.

Additionally, the molecules of the present invention can also be detected *in vivo* by imaging, for example imaging of cells which have undergone tumor progression or have metastasized. Antibody labels or markers for *in vivo* imaging of RPTKs include those detectable by X-radiography, NMR, PET, or ESR. For X-radiography, suitable labels include radioisotopes such as barium or cesium, which emit detectable radiation but are not overfly harmful to the subject. Suitable markers for NMR and ESR include those with a detectable characteristic spin, such as deuterium, which may be incorporated into the antibody.

A specific antibody or antibody portion which has been labeled with an appropriate detectable imaging moiety, such as a radioisotope (for example, ¹³¹I, ¹¹¹In, ⁹⁹Tc), a radio-opaque substance, or a material detectable by nuclear magnetic resonance, is introduced (for example, parenterally, subcutaneously or intraperitoneally) into the mammal to be examined for a disorder. It will be understood in the art that the size of the subject and the imaging system used will determine the quantity of imaging moieties needed to produce diagnostic images. In the case of a radioisotope moiety, for a human subject, the quantity of radioactivity injected will normally range from about 5 to 20 millicuries. The labeled antibody or antibody portion will then preferentially accumulate at the location of cells which contain a specific RPTK. In vivo tumor imaging is described in Burchiel et al., (1982).

pre-packaged diagnostic test kits comprising in one or more containers (i) at least one immunoglobulin of the invention and (ii) a reagent suitable for detecting the presence of said immunoglobulin when bound to its target. A kit may be conveniently used, e.g., in clinical settings or in home settings, to diagnose patients exhibiting a disease (e.g., skeletal dysplasia, craniosynostosis disorders, cell proliferative diseases or disorders, or tumor progression), and to screen and identify those individuals exhibiting a predisposition to such disorders. A composition of the invention also may be used in conjunction with a reagent suitable for detecting the presence of said immunoglobulin when bound to its target, as well as instructions for use, to carry out one or more methods of the invention.

Having now generally described the invention, the same will be more readily understood through reference to the following examples, which are provided by way of illustration and are not intended to be limiting of the present invention.

EXAMPLES

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An important approach to control FGFR3 activity is the generation of reagents that block receptor signaling. Without wishing to be bound by theory, molecules which bind the extracellular domain of the receptor may inhibit the receptor by competing with FGF or heparin binding or, alternatively, by preventing receptor dimerization. Additionally, binding to the extracellular domain may accelerate receptor internalization and turnover. Humanized antibodies are expected to have inhibitory/neutralizing action and are of particular interest since they are considered to be valuable for therapeutic applications, avoiding the human anti-mouse antibody response frequently observed with rodent antibodies. The experiments in which the neutralizing antibodies are screened, identified and obtained using fully synthetic human antibody libraries (for discovering highly specific binders against a wide variety of antigens) and FGFR3 extracellular domain are described below.

Example 1: Attempt to generate anti-FGFR3 antibodies

One hundred micrograms of soluble FGFR3 in complete Freund's Adjuvant were injected into Balb/c 3T3 naive mice (9 animals). Two repeated injections of 20 micrograms were performed at week intervals. 10 days after the second booster injection, blood was drawn from animals and serum was tested for the presence of polyclonal antibodies both by monitoring for binding to the receptor as well as for neutralizing activity at a dilution of 1:50. No significant neutralizing activity was observed in the tested serum (20% at most in some animals). A prefusion injection of 20 micrograms of soluble receptor was administered 1-2 days later but all the mice harboring some activity of neutralizing Ab died. The experiment was repeated twice with the same results.

Example 2: Generation of the FGFR3 Antigens

Two dimeric forms of the extracellular domain of the human FGFR3 were prepared for use as antigen. One was a histidine-tagged domain with a Serine 371 to Cysteine (S371C) substitution (thanatophoric dysplasia (TD) mutation) to facilitate dimerization and the second one an Fc fusion. The S371C variant was shown to bind heparin and FGF9 coated plates and

to inhibit FGF9-dependent FDCP-FR3 proliferation. The Fc fusion was similarly effective in binding assays demonstrating its potential as an inhibitor of FGFR function and as a target for selecting FGFR3 inhibitory molecules. Both soluble receptors were employed to select neutralizing human recombinant antibodies.

- 5 The two variants of the FGFR3 extracellular domain were prepared as follows:
 - 1. A construct containing the extracellular portion of FGFR3 with a thanatophoric dysplasia (TD) mutation to facilitate dimer formation conjugated to a His-tag (histidine tag) was generated. A bluescript plasmid comprising the human FGFR3 gene (pBS-hFGFR3) was used as template for PCR with the following primers:
- 10 5'-ACGTGCTAGC TGAGTCCTTG GGGACGGAGC AG (SEQ ID NO:2).
 - 5'-ACGTCTCGAG TTAATGGTGA TGGTGATGGT GTGCATACAC <u>ACA</u>GCCCGCC TCGTC (SEQ ID NO:3),

wherein the Ser 371 Cys (S371C) substitution is bold and underlined.

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The nucleotide sequence encoding the extracellular domain of FGFR3 with the TD substitution is denoted herein SEQ ID NO:7:

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TGAGTCCTTG GGG ACGGAGC AGCG CGTCGT GGGGC GAGAG GTCC CGGGCCC AGA 60
GCCCGGCCAG CAG GAGCAGT TGG TCTTCGG CAGC GGGGAT GCTGT GGAGC TGAGCT GTCC 120
CCCGCCGGG GGTGGTCCCA TGGGGCCCAC TGTCTGGGTC AAGGATGGCA CAGGGCTGGT 180
GCCCTCGGAG CGTGTCCTGG TGGGGCCCCA GCGGCTGCAG GTGCTGAATG CCTCCCACGA 240
GGACTCCGGG GCCTACAGCT GCCGGCAGCG GCTCACGCAG CGCGTACTGT GCCACTT CAG 300
TGTGCGGGTG ACAGACGCTC CATCCTCGGG AGATGACGAA GACGGGGAGG ACGAGGCTGA 360
GGACACAGGT GTG GACACAG GGGC CCCTTA CTGGA CACGG CCCGAG CGGA TGGACAA GAA 420
GCTGCTGGCC GTG CCGGCCG CCAA CACCGT CCGCT TCCGC TGCCCA GCCG CTGGCAA CCC 480
CACTCCCTCC ATCTCCTGGC TGAA GAACGG CAGGG AGTTC CGCGGC GAGC ACCGCATTGG 540
AGGCATCAAG CTG CGGCATC AGCA GTGGAG CCTGG TCATG GAAAGC GTGG TGCCCTC GGA 600
CCGCGGCAAC TACACCTGCG TCGTGGAGAA CAAGTTTGGC AGCATCCGGC AGACGTACAC 660
GCTGGACGTG CTGGAGCGCT CCCCGCACCG GCCCATCCTG CAGGCGGGGC TGCCGGCCAA 720
CCAGACGGCG GTG CTGGGCA GCGA CGTGGA GTTCC ACTGC AAGGTG TACA GTGACGC ACA 780
GCCCCACATC CAGTGGCTCA AGCA CGTGGA GGTGA ACGGC AGCAAG GTGG GCCCGGA CGG 840
CACACCCTAC GTT ACCGTGC TCAA GACGGC GGGCG CTAAC ACCACC GACA AGGAGCT AGA 900
GGTTCTCTCC TTG CACAACG TCACCTTTGA GGACG CCGGG GAGTAC ACCT GCCTGG CGGG 960
CAATTCTATT GGG TTTTCTC ATCA CTCTGC GTGGC TGGTG GTGCTG CCAG CCGAGGA GGA 1020
GCTGGTGGAG GCTGACGAGG CGGGCTGTGT GTATGCACAC CATCACCATC ACCATTA A
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The PCR fragment was digested with XhoI and ligated into pBlueScript digested with EcoRV and XhoI. The resulting plasmid, pBsFR3²³⁻³⁷⁴Tdhis, was cleaved with NdeI and

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XhoI and the DNA fragment encoding the extracellular domain of FGFR3 was ligated into the same restriction sites in pCEP-Pu/Ac7 (Yamaguchi et al., 1999; Kohfeldt et al., 1997), generating the pCEP-hFR3²³⁻³⁷⁴TDhis plasmid construct.

To express this FGFR3 variant, 293E cells (EBNA virus transfected 293 cells) were transfected with the aforementioned plasmid, pCEP-hFR3²³⁻³⁷⁴TDhis, clones were identified and grown. Cell supernatants analyzed by Western blot with anti-His antibody demonstrated high expression of the soluble receptor. Supernatants from large scale preparations were then subjected to batch affinity purification with Ni-NTA beads and the tagged soluble receptor was eluted by a step gradient ranging from 20 mM to 500 mM imidazol. A sample from each eluate was loaded onto a 7.5% SDS-PAGE and stained with GelCode (Pierce). In parallel Western blot analysis was performed and analyzed with anti-His antibodies. SDS-PAGE (Fig. 1) and immunoblot (not shown) analyses demonstrated peak amounts of purified extracellular FGFR3 in the 2nd (2) 50 mM imidazol fraction. About 0.5 mg of pure protein was obtained following this single step purification. In Figure 1, T=total protein, D= dialysed protein, U= unbound fraction.

To assess whether hFR3²³⁻³⁷⁴TDhis (hFR3-TDhis) retained the ability to associate with heparin and heparin-FGF complex, heparin coated wells were incubated with purified (2, 4 or 10 μg, labeled as FR3 2, FR3 4 or FR3 10, respectively in Fig. 2) or unpurified (FR3 sup) hFR3²³⁻³⁷⁴TDhis with (checkered bar) or without FGF9 (200ng/well, hatched bar). The binding of hFR3²³⁻³⁷⁴TDhis to each well was determined with anti-His antibody. Mock supernatant (M sup), PBS and unpurified mouse FR3AP (FGFR3-alkaline phosphatase, labeled as FRAP sup) were included as controls. This demonstrated that, like what was reported for the wild-type receptor, hFR3²³⁻³⁷⁴TDhis binds to heparin and that this interaction is augmented by the presence of FGF9 (Fig. 2). Finally, it was demonstrated that hFR3²³⁻³⁷⁴TDhis inhibits FDCP-FR3 FGF-dependent proliferation in a dose dependent manner. hFR3²³⁻³⁷⁴TDhis had no inhibitory effect on proliferation when FDCP-FR3 cells were grown in the presence of IL-3. Taken together, hFR3²³⁻³⁷⁴TDhis proved to be a good candidate as a

2. The extracellular domain of FGFR3 and FGFR1 were prepared as Fc fusions (FR3exFc and FR1exFc). The amino acid sequence of FGFR3 (NCBI access no: NP_000133) is denoted herein SEQ ID NO:1.

target antigen for screening for FGFR3 neutralizing antibodies.

1 MGA PACALAL CVAVAIVAGA SSESLGTEQR VVGRAA EVPG PEPGQQE QLV FGSGDAVE LS

61 CPPPGGGPMG PTWW KDGTG LVPSER VLVG PQRLQVLNAS HEDSGAYS CR QRLTQRVLCH
121 FSVRVTDAPS SGDD EDGEDE AEDTG VDTGA PYWTRP ERMD KKLLAVP AAN TVRFRCPA AG
181 NPTPSISWLK NGREF RGEHR IGGIKL RHQQ WSLVMES VVP SDRGNYTC VV ENKFGSIRQ T
241 YTLD VLERSP HRPLL QAGLP ANQTAV LGSD VEFHCKV YSD AQPHLQWL KH VEVNGSKVG P
301 DGTP YVTVLK TAGAN TTDKE LEVLS LHNVT FEDAGE YTCL AGNSIGF SHH SAWLVVLP AE
361 EELV EADEAG SVYAG ILSYG VGFFLF ILVV AAVTLCR LRS PPKKGLGS PT VHKISRFPL K
421 RQVS LESNAS MSSNT PLVRI ARLSSG EGPT LANVSEL ELP ADPKWELS RA RLTLGKPLGE
481 GCFG QVVMAE AIGID KDRAA KPVTVA VKML KDDATDK DL SEMEMMK MIGKHKNIIN
541 LLGA CTQGGP LYVLV EYAAK GNLREF LRAR RPPGLDY SFD TCKPPEEQ LT FKDLVSCAY Q
661 MAPE ALFDRV YTHQS DVWSF GVLLWE IFTL GGS PYPG IPV EELFKLLK EG HRMDKPANC T
721 HDLY MIMREC WHAAP SQRPT FKQLVE DLDR VLTVTST DEY LDLSAPFE QY SPGGQDTPS S
781 SSSG DDSVFA HDLLP PAPPS SGGSRT

To construct the FR3exFc fusion, a nucleotide sequence (SEQ ID NO:4) encoding the extracellular domain of FGFR3 was PCR amplified to contain terminal KpnI and BamHI restriction sites for insertion into the KpnI and BamHI sites of pCXFc (SEQ ID NO:5). This insertion positions the extracellular domain of FGFR3 to be expressed as a fusion with the Fc amino acid sequence (SEQ ID NO:6).

SEQ ID NO:4:

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20	GCGCGCTGCC	TGA GGACGCC	GCGG CCCCG	CCCCC GCCAT	GGGCGC CCCT	GCCTGCG CCC	60
	TCGCGCTCTG	CGT GGCCGTG	GCCA TCGTGG	CCGGC GCCTC	CTCGGA GTCC	TTGGGGA CGG	120
	AGCAGCGCGT	CGT GGGGCGA	GCGG CAGAAG	TCCCG GGCCC	AGAGCC CGGC	CAGCAGG AGC	180
	AGTTGGTCTT	CGG CAGCGGG	GATG CTGTGG	AGCTG AGCTG	TCCCC CGCCC	GGGGGT GGTC	240
	CCATGGGGCC	CACTGTCTGG	GTCA AGGATG	GCACA GGGCT	GGTGCC CTCG	GAGCGTG TCC	300
25	TGGTGGGGCC	CCA GCGGCTG	CAGG TGCTGA	ATGCC TCCCA	CGAGGA CTCC	GGGGCCT ACA	360
	GCTGCCGGCA	GCG GCTCACG	CAGC GCGTAC	TGTGC CACTT	CAGTGT GCGG	GTGACAG ACG	420
	CTCCATCCTC	${\tt GGGAGATGAC}$	GAA GACGGGG	AGGA CGAGGC	TGAGG ACACA	GGTGTG GACA	480
	CAGGGGCCCC	TTA CTGGACA	CGGC CCGAGC	$\operatorname{GGATG}\operatorname{GACAA}$	GAAGCT GCTG	GCCGTGC CGG	540
	CCGCCAACAC	CGT CCGCTTC	CGCT GCCCAG	CCGCT GGCAA	CCCCACTCCC	TCCATCT CCT	600
30	GGCTGAAGAA	CGG CAGGGAG	TTCC GCGGCG	AGCAC CGCAT	$\mathtt{TGGAG}\mathtt{GCATC}$	AAGCTG CGGC	660
	ATCAGCAGTG	GAG CCTGGTC	ATGG AAAGCG	TGGTG CCCTC	GGACCG CGGC	AACTACA CCT	720
	GCGTCGTGGA	GAA CAAGTTT	GGCA GCATCC	GGCAG ACGTA	CACGCT GGAC	GTGCTGG AGC	780
	GCTCCCGCA	CCGGCCCATC	CTGC AGGCGG	GGCTG CCGGC	CAACCA GACG	GCGGTGC TGG	840
	GCAGCGACGT	GG AGTTCCAC	${\tt TGCAAGGTGT}$	ACAG TGACGC	ACAGC CCCAC	ATCCAG TGGC	900
35	TCAAGCACGT	GGA GGTGAAC	GGCA GCAAGG	TGGGC CCGGA	CGGCAC ACCC	TACGTTA CCG	960
	TGCTCAAGAC	GGC GGGCGCT	AACA CCACCG	ACAAG GAGCT	AGAGGT TCTC	TCCTTGC ACA	1020
	ACGTCACCTT	TGA GGACGCC	GGGG AGTACA	CCTGC CTGGC	$\operatorname{GGGCAA}\operatorname{TTCT}$	ATTGGGT TTT	1080
	CTCATCACTC	TGC GTGGCTG	GTGG TGCTGC	CAGCC GAGGA	GGAGCT GGTG	GAGGCTG ACG	1 140

AGGCGGG 1147

SEQ ID NO:5:

	GACGGATCGG GAG ATCTCCC GATC CCCTAT GGTCG ACTCT CAGTAC AATC TGCTCTG ATG	60
	CCGCATAGTT AAG CCAGTAT CTGC TCCCTG CTTGT GTGTT GGAGGT CGCT GAGTAGT GCG	120
5	CGAGCAAAAT TTA AGCTACA ACAA GGCAAG GCTTG ACCGA CAATTG CATG AAGAATC TGC	180
	TTAGGGTTAG GCGTTTTGCG CTGCTTCGCG ATGTA CGGGC CAGATA TACG CGTTGAC ATT	240
	GATTATTGAC TAGTTATTAA TAGTAATCAA TTACGGGGTC ATTAGTTCAT AGCCCATATA	300
	TGGAGTTCCG CGTTACATAA CTTACGGTAA ATGGCCCGCC TGGCTGACCG CCCAACGACC	360
	CCCGCCCATT GACGTCAATA ATGACGTATG TTCCCATAGT AACGCCAATA GGGACTTTCC	420
10	ATTGACGTCA ATGGGTGGAC TATTTACGGT AAACTGCCCA CTTGGCAGTA CATCAAGTGT	480
	ATCATATGCC AAGTACGCCC CCTATTGACG TCAATGACGG TAAATGGCCC GCCTGGCATT	540
	ATGCCCAGTA CATGACCTTA TGGGACTTTC CTACTTGGCA GTACATCTAC GTATTAGTCA	600
	TCGCTATTAC CATGGTGATG CGGTTTTGGC AGTACATCAA TGGGCGTGGA TAGCGGTTTG	660
	ACTCACGGGG ATTTCCAAGT CTCCACCCCA TTGACGTCAA TGGGAGTTTG TTTTGGCACC	720
15	AAAATCAACG GGA CTTTCCA AAAT GTCGTA ACAAC TCCGC CCCATT GACG CAAATGG GCG	780
	GTAGGCGTGT ACGGTGGGAG GTCTATATAA GCAGAGCTCT CTGGCTAACT AGAGAACCCA	840
	CTGCTTACTG GCTTATCGAA ATTAATACGA CTCACTATAG GGAGACCCAA GCTGGCTAGC	900
	GTTTAAACTT AAGCTTGGTA CCGAGCTCGG ATCC CCGTCG TGCAT CTATC GAAGGT CGTG	960
20	GA GAT CCC GAG GAG CCC AAA TCT TGT GAC AAA ACT CAC ACA TGC CCA	1007
	ASP PRO GLU GLU PRO LYS SER CYS ASP LYS THR HIS THR CYS PRO 15	
		1055
25	PRO CYS PRO ALA PRO GLU LEU LEU GLY GLY PRO SER VAL PHE LEU PHE 31	
		1103
	PRO PRO LYS PRO LYS ASP THR LEU MET ILE SER ARG THR PRO GLU VAL 47	
2.0		1151
30	THR CYS VAL VAL VAL ASP VAL SER HIS GLU ASP PRO GLU VAL LYS PHE 63	
		1199
	ASN TRP TYR VAL ASP GLY VAL GLU VAL HIS ASN ALA LYS THR LYS PRO 79	
35	CGG GAG GAG CAG TAC AAC AGC ACG TAC CGG GTG GTC AGC GTC CTC ACC	1247
	ARG GLU GLU GLN TYR ASN SER THR TYR ARG VAL VAL SER VAL LEU THR 95	
	GTC CTG CAC CAG GAC TGG CTG AAT GGC AAG GAG TAC AAG TGC AAG GTC VAL LEU HIS GLN ASP TRP LEU ASN GLY LYS GLU TYR LYS CYS LYS VAL 111	1295
40		
	TCC AAC AAA GCC CTC CCA GCC CCC ATC GAG AAA ACC ATC TCC AAA GCC	1343
	SER ASN LYS ALA LEU PRO ALA PRO ILE GLU LYS THR ILE SER LYS ALA 127	
	AAA GGG CAG CCC CGA GAA CCA CAG GTG TAC ACC CTG CCC CCA TCC CGG	1391
45	LYS GLY GLN PRO ARG GLU PRO GLN VAL TYR THR LEU PRO PRO SER ARG 143	

- 40 -

					TGC CTG GTC AAA GGC CYS LEU VAL LYS GLY 15	1439 9
5					AGC AAT GGG CAG CCG SER ASN GLY GLN PRO 17	1487 5
					GAC TCC GAC GGC TCC ASP SER ASP GLY SER 19	1535 1
10					AGC AGG TGG CAG CAG SER ARG TRP GLN GLN 20	1583 7
15					GCT CTG CAC AAC CAC ALA LEU HIS ASN HIS 22	1631 3
				TCT CCG GGT SER PRO GLY	AAA TGATCTAGAG LYS 23	1677 5
	GGCCCGTTTA	AAC CCGCTGA	TCAG CCTCG	A CTGTG CCTTC	TAGTTG CCAG CCATCTG TTG	1737
20	TTTGCCCCTC	CCC CGTGCCT	TCCT TGACCO	C TGGAA GGTGC	CACTCC CACT GTCCTTT CCT	1797
	AATAAAATGA	GGA AATTGCA	TCGC ATTGT	C TGAGT AGGTG	TCATTC TATT CTGGGGG GTG	1857
	GGGTGGGGCA	GGA CAGCAAG	GGGG AGGAT	r gggaa gacaa	TAGCAG GCAT GCTGGGG ATG	1917
	CGGTGGGCTC	TAT GGCTTCT	GAGG CGGAA	A GAACC AGCTG	GGGCTC TAGG GGGTATC CCC	1977
	ACGCGCCCTG	TAG CGGCGCA	TTAA GCGCGG	G CGGGT GTGGT	GGTTAC GCGC AGCGTGA CCG	2037
25	CTACACTTGC	CAG CGCCCTA	GCGC CCGCT	C CTTTC GCTTT	CTTCC CTTCC TTTCTCGCCA	2097
	CGTTCGCCGG	CTT TCCCCGT	CAAG CTCTAA	A ATCGG GGCAT	CCCTTT AGGG TTCCGAT TTA	2157
					TGATGG TTCA CGTAGTG GGC	2217
					GTCCACGTTC TTTAATAGTG	2277
2.0					GGTCT ATTCT TTTGAT TTAT	2337
30					GCTGAT TTAA CAAAAAT TTA	2397
					GGAAAG TCCC CAGGCTC CCC	2457
					AGCAA CCAGG TGTGGA AAGT	2517
					TCTCAA TTAG TCAGCAA CCA GCCCAG TTCC GCCCATT CTC	2577
35					CGAGGC CGCC TCTGCCT CTG	2637 2697
33					AGGCTTTTGC AAAAAGCTCC	2757
					GTTGACAATT AATCATCGGC	2817
					TAAACCATGG CCAAGTTGAC	2877
					GCGGTC GAGT TCTGGAC CGA	2937
40					GCCGGT GTGG TCCGGGA CGA	2997
	CGTGACCCTG	TTC ATCAGCG	CGGT CCAGG	A CCAGG TGGTG	CCGGAC AACA CCCTGGC CTG	3057
	GGTGTGGGTG	CGC GGCCTGG	ACGA GCTGTA	A CGCCG AGTGG	TCGGAG GTCG TGTCCAC GAA	3117
	CTTCCGGGAC	GCC TCCGGGC	CGGC CATGA	C CGAGA TCGGC	GAGCAG CCGT GGGGGCG GGA	3177
	GTTCGCCCTG	CGC GACCCGG	CCGG CAACTO	G CGTGC ACTTC	GTGGCC GAGG AGCAGGA CTG	3237

	ACACGTGCTA	CGA GATTTCG	ATTC CACCGC	CGCCT TCTAT	GAAAGG TTGG	GCTTCGG AAT	3297
	CGTTTTCCGG	GAC GCCGGCT	GGAT GATCCT	CCAGC GCGGG	GATCT CATGC	TGGAGT TCTT	3357
	CGCCCACCC	AAC TTGTTTA	TTGC AGCTTA	TAATG GTTAC	AAATAA AGCA	ATAGCAT CAC	3417
	AAATTTCACA	AAT AAAGCAT	TTTT TTCACT	GCATT CTAGT	TGTGGT TTGT	CCAAACT CAT	3477
5	CAATGTATCT	TAT CATGTCT	GTAT ACCGTC	GACCT CTAGC	TAGAGC TTGG	CGTAATC ATG	3537
	GTCATAGCTG	TTTCCTGTGT	GAA ATTGTTA	TCCG CTCACA	ATTCC ACACA	ACATAC GAGC	3597
	CGGAAGCATA	AAG TGTAAAG	CCTG GGGTGC	CTAAT GAGTG	AGCTAA CTCA	CATTAAT TGC	3657
	GTTGCGCTCA	CTGCCCGCTT	TCCA GTCGGG	AAACC TGTCG	TGCCAG CTGC	ATTAATG AAT	3717
	CGGCCAACGC	GCG GGGAGAG	GCGGTTTGCG	TATTG GGCGC	$\mathtt{TCTTCC}\mathtt{GCTT}$	CCTCGCTCAC	3777
10	TGACTCGCTG	CGC TCGGTCG	${\tt TTCGGCTGCG}$	GCGAG CGGTA	TCAGCT CACT	CAAAGGC GGT	3837
	AATACGGTTA	TCC ACAGAAT	CAGG GGATAA	CGCAG GAAAG	AACATG TGAG	CAAAAGG CCA	3897
	GCAAAAGGCC	AGG AACCGTA	AAAA GGCCGC	GTTGC TGGCG	$\mathtt{TTTTTC}\mathtt{CATA}$	GGCTCCG CCC	3957
	CCCTGACGAG	CAT CACAAAA	ATC GACGCTC	AAGT CAGAGG	TGGCG AAACC	CGACAG GACT	4017
	ATAAAGATAC	CAG GCGTTTC	CCCC TGGAAG	CTCCC TCGTG	CGCTCT CCTG	TTCCGAC CCT	4077
15	GCCGCTTACC	GGA TACCTGT	CCGC CTTTCT	CCCTT CGGGA	AGCGTG GCGC	TTTCTCA ATG	4137
	CTCACGCTGT	AGG TATCTCA	GTTC GGTGTA	GGTCG TTCGC	TCCAAG CTGG	GCTGTGT GCA	4197
	CGAACCCCCC	GTT CAGCCCG	ACCG CTGCGC	CTTAT CCGGT	AACTAT CGTC	TTGAGTC CAA	4257
	CCCGGTAAGA	CAC GACTTAT	CGCC ACTGGC	AGCAG CCACT	GGTAAC AGGA	TTAGCAG AGC	4317
	GAGGTATGTA	GGC GGTGCTA	CAGA GTTCTT	GAAGT GGTGG	CCTAAC TACG	GCTACAC TAG	4377
20	AAGGACAGTA	TTT GGTATCT	GCGC TCTGCT	GAAGC CAGTT	ACCTTC GGAA	AAAGAGT TGG	4437
	TAGCTCTTGA	TCC GGCAAAC	AAAC CACCGC	TGGTA GCGGT	$\operatorname{GGTTTT}\operatorname{\mathbf{TTTG}}$	TTTGCAA GCA	4497
	GCAGATTACG	CGC AGAAAAA	AAGG ATCTCA	AGAAG ATCCT	${\tt TTGATCTTTT}$	CTACGGG GTC	4557
	TGACGCTCAG	TGG AACGAAA	ACTC ACGTTA	AGGGA TTTTG	GTCATG AGAT	TATCAA AAAG	4617
•	GATCTTCACC	TAGATCCTTT	$\mathtt{TAAA}\mathtt{TTAAAA}$	ATGAA GTTTT	AAATCA ATCT	AAAGTAT ATA	4677
25	TGAGTAAACT	${\tt TGGTCTGACA}$	GTTA CCAATG	CTTAA TCAGT	GAGGCA CCTA	TCTCAGC GAT	4 737
	CTGTCTATTT	CGTTCATCCA	TAGT TGCCTG	ACTCC CCGTC	GTGTAG ATAA	CTACGAT ACG	4797
	GGAGGGCTTA	CCATCTGGCC	CCAGTGCTGC	AATG ATACCG	CGAGA CCCAC	GCTCAC CGGC	4857
	TCCAGATTTA	TCA GCAATAA	ACCA GCCAGC	CGGAA GGGCC	GAGCGC AGAA	GTGGTCC TGC	4917
	AACTTTATCC	GCCTCCATCC	AGTC TATTAA	TTGTT GCCGG	GAAGCT AGAG	TAAGTAG TTC	4977
30	GCCAGTTAAT	AGT TTGCGCA	ACGT TGTTGC	CATTG CTACA	GGCATC GTGG	TGTCACGCTC	5037
	GTCGTTTGGT	ATG GCTTCAT	TCAG CTCCGG	TTCCC AACGA	TCAAGG CGAG	TTACATGATC	5097
	CCCCATGTTG	TGC AAAAAAG	CGGT TAGCTC	CTTCG GTCCT	$\mathtt{CCGATC}\mathtt{GTTG}$	TCAGAAG TAA	5157
	GTTGGCCGCA	$\mathtt{GTG}\mathtt{TTATCAC}$	TCAT GGTTAT	GGCAG CACTG	CATAAT TCTC	TTACTGT CAT	5217
	GCCATCCGTA	AGA TGCTTTT	CTG TGACTGG	TGAG TACTCA	ACCAA GTCAT	TCTGAG AATA	5277
35	GTGTATGCGG	CGA CCGAGTT	GCTC TTGCCC	GGCGT CAATA	CGGGAT AATA	CCGCGCC ACA	5337
	TAGCAGAACT	TTA AAAGTGC	TCAT CATTGG	AAAAC GTTCT	TCGGGG CGAA	AACTCTC AAG	5397
	GATCTTACCG	CTG TTGAGAT	CCAG TTCGAT	GTAAC CCACT	CGTGCA CCCA	ACTGATC TTC	5457
	AGCATCTTTT	ACT TTCACCA	GCGT TTCTGG	GTGAG CAAAA	ACAGGA AGGC	AAAATGC CGC	5517
	AAAAAAGGGA	ATA AGGGCGA	CACG GAAATG	TTGAA TACTC	ATACTC TTCC	TTTTTCAATA	5577
40	TTATTGAAGC	ATT TATCAGG	${\tt GTTATTGTCT}$	CATGA GCGGA	TACATA TTTG	AATGTAT TTA	5637

GAAAAATAAA CAAATAGGGG TTCCGCGCAC ATTTCCCCGA AAAGTGCCAC CTGACGTC

5695

SEQ ID NO:6

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ASP PRO GLU GLU PRO LYS SER CYS ASP LYS THR HIS THR CYS PRO PRO 16 CYS PRO ALA PRO GLU LEU LEU GLY GLY PRO SER VAL PHE LEU PHE PRO 32 5 PRO LYS PRO LYS ASP THR LEU MET ILE SER ARG THR PRO GLU VAL THR 48 CYS VAL VAL VAL ASP VAL SER HIS GLU ASP PRO GLU VAL LYS PHE ASN 64 TRP TYR VAL ASP GLY VAL GLU VAL HIS ASN ALA LYS THR LYS PRO ARG 80 GLU GLU GLN TYR ASN SER THR TYR ARG VAL VAL SER VAL LEU THR VAL 96 LEU HIS GLN ASP TRP LEU ASN GLY LYS GLU TYR LYS CYS LYS VAL SER 112 10 ASN LYS ALA LEU PRO ALA PRO ILE GLU LYS THR ILE SER LYS ALA LYS 128 GLY GLN PRO ARG GLU PRO GLN VAL TYR THR LEU PRO PRO SER ARG ASP 144 GLU LEU THR LYS ASN GLN VAL SER LEU THR CYS LEU VAL LYS GLY PHE 160 TYR PRO SER ASP ILE ALA VAL GLU TRP GLU SER ASN GLY GLN PRO GLU 176 ASN ASN TYR LYS THR THR PRO PRO VAL LEU ASP SER ASP GLY SER PHE 192 15 PHE LEU TYR SER LYS LEU THR VAL ASP LYS SER ARG TRP GLN GLN GLY 208 ASN VAL PHE SER CYS SER VAL MET HIS GLU ALA LEU HIS ASN HIS TYR 224 THR GLN LYS SER LEU SER LEU SER PRO GLY LYS 235

Both FR3exFc and FR1exFc soluble receptors were demonstrated to be expressed to a high level in transiently transfected 293T cells (T-cell antigen infected human embryonic kidney 293 cells). The observation that both soluble receptors remain bound to heparin-coated wells even following extensive washes led the laboratory of the present inventors to try to purify the proteins with the commercial heparin-SepharoseTM resin (Pharmacia). One hundred ml volume supernatants, harvested 48 hours post transfection with either FR3exFc or FR1exFc coding plasmids, were incubated overnight at 4°C with 1 ml heparin-SepharoseTM resin. The resin was washed and then subjected to PBS supplemented with increasing concentration of NaCl. Aliquots of each fraction were analyzed by 7.5% SDS-PAGE stained with GelCode (Pierce) demonstrating a purification profile of more than 90% homogeneity and a peak elution at 400 mM NaCl for FR3exFc (Fig. 3; T=total protein, U=unbound fraction, W=wash). In contrast, FR1exFc was hardly retained on the resin. This result was confirmed by Western analysis of the same fractions with anti-FGFR1ex antibodies demonstrating that most of FR1exFc is in the unbound fraction (not shown).

Functional analysis of FR3exFc and FR1exFc showed that both compete efficiently for FGF9 binding and stimulating FGFR3, thus, demonstrating their potential as inhibitors of FGFRs function and as a target (FR3exFc) for selecting FGFR3 inhibitory molecules.

Neutralizing effect of soluble receptors

The ability of hFR3-TDhis and FR3exFc to inhibit FGF-dependent FDCP-R3 cell proliferation was compared. Both soluble receptors inhibited FDCP-R3 cell proliferation, however, FR3exFc was about 60 times more potent than hFR3TDhis (Fig. 4; legend: ◆
5 FDCP-FR3²³⁻³⁷⁴TDhis on FDCP-FR3 cells + FGF9, ■-FR3exFc on FDCP-FR3 cells + FGF9, ▲- FDCP-FR3²³⁻³⁷⁴TDhis on FDCP-FR3 cells + IL, X- FR3exFc on FDCP-FR3 cells + IL3). Neither had an effect on FDCP cells stimulated with IL3. The fact that FR3exFc is entirely in dimeric form whereas only a small proportion (1/10) of hFR3²³⁻³⁷⁴TDhis is in a dimeric form might explain, at least in part, this difference.

Example 3: Screening for Antibodies

Panning and first screening of Ab Binding Characterization

The screening strategies to identify Fabs from the Human Combinatorial Antibody Library (HuCAL®), developed at MorphoSys, Munich, Germany and disclosed in WO 97/08320, US patent 6,300,064, and Knappik et al., (2000), the entire contents of which are incorporated herein by reference, using soluble dimeric forms of the extracellular domain of the FGFR3 receptor are shown in Table 2.

TABLE 2
Panning Strategies

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	Panning Round 1	Panning Round 2	Panning Round 3
Screen 1	FR3-TDhis	нек293	FR3-TDhis
Screen 2	FR3exFc captured with mouse anti-human IgG	RCJ-FR3ach	FR3exFc captured with mouse anti-human IgG
Screen 3	FR3-TDhis (Round 1 of panning 1)	RCJ-FR3ach & RCJ-FR3wt	FR3exFc Captured with mouse anti-human IgG

The screening was carried out, for example in Screen 1, by coating the wells of a 96 well plate with hFR3²³⁻³⁷⁴TDhis (FR3-TDhis), panning with the bacteriophage library and

selecting the positive clones. The positive clones were then tested on HEK293 (293, human embryonic kidney) cells, expressing endogenous FGFR3. The positive clones were selected and rescreened on FR3-TDhis. Two additional similar screenings were carried out as shown in Table 2. In screen 2 the first and third pannings were carried out with the FR3exFc antigen and the second panning carried out with RCJ cells expressing a mutant (achondroplasia) form of FGFR3.

An overview of the number of initial hits and of the candidate clones is shown in Table 3.

Table 3

Overview of Screenings 1, 2 and 3 on FGFR3

	screened clones	primary hits	sequenced clones	consolidated candidate clones (ELISA & FACS)
Screen 1	1076	208	69	15 MSPRO 1-15
Screen 2	864	300	32	22 MSPRO 20-33 and 52-59
Screen 3	768	487	52	11 MSPRO 40-50

10 Sequence and Vector Data

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A plasmid map and sequence (SEQ ID NO:52) of the dHLX-MH vector are presented in Fig. 28A and 28B.

Figure 29A shows the plasmid map of the phage display vector used in accordance with the present invention. Figure 29B is the corresponding sequence and restriction digest map (SEQ ID NO:53).

Figure 30 displays the polynucleotide sequences of the specific VL and VH domains of MSPRO2 (SEQ ID NO:74 and 84); MSPRO11 (SEQ ID NO:70 and 85), MSPRO12 (SEQ ID NO:75 and 89); MSPRO21 (SEQ ID NO:67 and 78); MSPRO24 (SEQ ID NO:64 AND 79); MSPRO26 (SEQ ID NO:71 AND 86); MSPRO28 (SEQ ID NO:62 AND 80); MSPRO29 (SEQ ID NO:65 AND 87); MSPRO54 (SEQ ID NO:73 AND 82); MSPRO55 (SEQ ID NO:69 AND 83); MSPRO59 (SEQ ID NO:76 AND 91). The sequences include the framework domains 1-4 and the CDR domains 1-3. SEQ ID NO:61, 63, 66, 68, and 73

denote the polynucleotide sequences of the parent VL (kappa or lambda) strands. SEQ ID NO:77, 81, 88 and 90 denote the polynucleotide sequences of the VH parent strands.

Example 4: Analysis of Fabs identified by first screening.

Specificity of Antibody recognition

The first screening yielded 15 different Fabs that specifically recognize FGFR3 *in vitro* and on the cell surface. Fourteen of these were produced and sent to ProChon for further analysis. LY6.3, an anti-lysosyme antibody, was isolated from the same library and serves as a control. ELISA analysis, according to the following protocol was carried out to determine the specificity of the isolated Fabs for FGFR3 or FGFR1.

10 Fab-FR3/Fc Binding Assay

MaxiSorp ELISA plates were coated with 100 μl anti-human Fc (10 μg/ml) in bicarbonate overnight at 4°C. Wells were washed five consecutive times with a PBS solution containing 0.1% Tween 20 (PBST). The well surface was blocked with 250 μl PBST+3%BSA (blocking solution) for 1 hour at 37°C. This was followed by capturing 1 μg of FGFR/Fc for 1 hour at room temperature. To assess the antibody binding to the captured FGFR/Fc, 1 μg each of the tested Fabs was incubated in 100 μl blocking solution per well 1 hour at room temperature Wells were washed 5 times with PBST. Reaction was initiated with the addition of 100 μl of 0.8μg/ml goat anti-human Fab-HRP diluted in blocking solution, subsequently washed and detected with TMB substrate (Pierce). The absorbance was measured at 450 nm. A comparison of ELISA analyses done in the laboratory of the present inventors (Prochon) and at MorphoSys is presented in the following Table 4.

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TABLE 4

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ProChon	<u>MorphoSys</u>				
	FR1/Fc	FR3/Fc	FR1/Fc	FR3/Fc	
MS-PRO1	+-}-	++	+/-	+	
MS-PRO2	-	++	-	++	
MS-PRO3	+	++	-	+-1-	
MS-PRO4	-	+	-	++	
MS-PRO5	-	++	+/-	+	
MS-PRO6	-	++	-	+	
MS-PRO7	-	++	-	+	
MS-PRO8	+	1-1-	_	+	
MS-PRO9	-	+/-	+/-	+	
MS-PRO10	+	++	_	+-}-	
MS-PRO11	-	+/-	+	+-+	
MS-PRO12	-	+/-	-	1-1-	
MS-PRO13	-	+/-	+/-	+	
MS-PRO14	- .	-	-	+	
LY6.3 (control)	-	-			

In most cases, the data generated at Morphosys and in the laboratory of the present inventors are in agreement. However, some Fabs behave differently. For example, MS-PRO3 and 10 were found to be completely FGFR3 specific under Morphosys conditions. In the laboratory of the present inventors, both show considerable cross-reaction with FGFR1. The FACS analysis, done at Morphosys, supports the Prochon results for MS-PRO3 but not for MS-PRO10. Taking into account the potency and specificity of the Fabs, MS-PRO2 has the highest score according to these preliminary data.

Example 5: Affinity of Fab to FGFR3

The affinity measurements were performed by BIAcore according to the standard procedure recommended by the supplier (Pharmacia). The anti-Fc antibody was coupled via the EDC/NHS chemistry to the chip and subsequently FGFR3 was captured. The Fabs of the invention were then bound to this surface.

Table 5 shows a comparison of affinities of Fabs candidates to FGFR3 as determined by BIAcore and by FACS-scatchard.

Table 5

Comparison of Antibody Affinities to FGFR3

determined by BIAcore and FACS-Scatchard

Fab clone	BIAcore [nM]	Indirect FACS-Scatchard [nM]
MSPRO2	37 ± 10	43
MSPRO11	4 ± 2	4
MSPRO12	14 ± 2	6.5
MSPRO21	9 ± 2	0.6
MSPRO24	10 ± 2	0.3
MSPRO26	4 ± 1	1.4
MSPRO28	9 ± 0.4	0.3
MSPRO29	6 ± 4	0.4

Table 1E shows the affinity as determined by BIAcore for the Fab candidates shown in Table 5 converted into the Fab mini-antibody format, Fab-dHLX-MH, where a dimer of the Fab monomer is produced after insertion into an expression vector as a fusion protein.

Table 6 shows the results of a competition assay wherein each MSPRO Fab was bound to the FGFR3 at a concentration of 500nM or 1,000 nM and coinjected in pairs with the other MSPRO Fabs. The (-) indicates binding to the same or nearby epitope while (+) indicates binding to different epitope. The results show that MSPRO2 and 12 bind to the same or nearby epitope while MSPRO11, 21, 24, 26, 28 and 29 bind to an epitope different from that of MSPRO2 or 12.

Example 6: Specific Neutralizing Activity of the Antibodies

A: FDCP Cell Proliferation Assay

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The FDCP cell line is a murine immortalized, interleukin 3 (IL3) dependent cell line of myelocytic bone marrow origin, which does not express endogenous FGF Receptors (FGFR).

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Upon transfection with FGFR cDNA, the FDCP cell line exhibits an FGF dose dependent proliferative response that can replace the dependence on IL3. FDCP cell lines, transfected with FGFR cDNAs can therefore be used to screen for specific inhibitors or activators of FGFR, as well as for analyzing FGFR signaling. The FDCP cell response to various ligands was quantitated by a cell proliferation assay with XTT reagent (Cell Proliferation Kit, Biological Industries Co.). The method is based on the capability of mitochondrial enzymes to reduce tetrazolium salts into soluble colored formazan compounds which can be quanititated and is indicative of cell viability. Specifically, FDCP cells expressing FGFR3IIIb, FGFR3IIIc or FGFR1 were grown in "full medium" (Iscove's Medium containing 2ml glutamine, 10% FCS, 100ug/ml penicillin, 100ug/ml streptomycin) supplemented with 5ug/ml heparin and 10ng/ml FGF9. Cells were split every 3 days and kept in culture no more than one month. One day prior to the experiment, the cells were split. Before the experiment, the cells were washed 3 times (1000 rpm, 6 min) with full medium. Later, the cells were resuspended and counted with Trypan Blue. Twenty thousand (20,000) cells /well were added to wells in a 96-well plate in 50ul in full medium containing 5 ug/ml heparin. Conditioned medium was added in an additional volume of 50ul full medium containing FGF9 at varying concentrations to a final volume of 100ul. A primary stock solution (usually 2x the higher concentration) of the antibody (or Fabs) was prepared in Iscove's+++ containing 5µg/ml heparin and 2.5ng/ml FGF9 or IL-3 (final concentration 1.25 ng/ml). Dilutions were filtered in a 0.2 µm syringe nitrocellulose filter blocked first with 1mg/ml BSA and washed then with Iscove's+++. Aliquots of requested serial dilutions were prepared. Dilutions were kept on ice until use, 50 ul of the corresponding 2x final concentration was added to each well and the plate was incubated at 37°C for either 40 hours or either 64 hours. After incubation, the reaction was developed as follows: 100 µl of activator solution was added to 5 ml XTT reagent and mixed gently. 50 µl of mixture was added to each well. Optical density (OD) at 490 nm at this point gave the zero time reading. Cells were then incubated at 37°C for 4 hours (in the case of a 40 hour incubation) or 2 hours (in the case of a 64 hour incubation) and proliferation was measured by O.D. at 490 nm (A490).

It is noted that the assay is successful when the O.D. of untreated control growing with saturated amounts of FGF (10 and 20 ng/ml) is at least 1.3 O.D. units. Furthermore, it is noted that the background of wells with no cells should be 0.2-0.35 O.D. units and that the

O.D. absorbance of 1.25 ng/ml FGF9 should not be less than 40% of the O.D. absorbance achieved with saturated FGF 9 concentration (10 and 20 ng/ml). Specific inhibition of FGF and FGF receptor mediated proliferation should always be accompanied with lack of any inhibition of the same antibody concentration on IL-3 dependent cell proliferation.

- 5 The following FDCP cell lines were used:
 - *FDCP-C10: FDCP cells transfected with the human wild-type FGF receptor 3IIIc.
 - *FDCP-R3: FDCP cells transfected with the human wild-type FGF receptor 3IIIb.
 - *FDCP-R1: FDCP cells transfected with the human wild-type FGFR1.
- *FDCP-F3Ach: FDCP cells infected with human FGFR3 mutated at amino acid Glycine 380 to Arginine (G380R), analogous to the most common human achondroplasia mutation.

B: Neutralizing activity

The neutralizing activity of the antibodies was measured by the aforementioned cell proliferation analysis in FDCP-FR3 and FDCP-FR1 cell lines. Increasing amounts of the indicated Fabs (MSPRO 2, 3 and 4) were added to FDCP-FR3 (closed triangle ▲(2), star * (3), and circle ● (4)) or FDCP-FR1 (diamond ♦ (2), square ■ (3) and open triangle Δ(4)) grown in the presence of FGF9 (Fig. 5). Two days later, an XTT proliferation assay was performed. While none of the Fabs inhibited FDCP-FR1 cell proliferation, MSPRO2 and 3 inhibited FDCP-FR3 proliferation with a similar IC50 of about 1.0 μg/ml (Fig. 5). In contrast, MS-PRO4 had no inhibitory effect on FDCP-FR3 proliferation. These data are in agreement with those generated at Morphosys. The rest of the Fabs were similarly analyzed on FDCP-FR3 expressing cells. Increasing amounts of the indicated Fabs were added to FDCP-FR3 grown in the presence of FGF9 (Fig. 6). The results of the proliferation assay done at Morphosys and at Prochon are compared in Table 6. (NA- data not available)

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Table 6

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	Prochon		Morphosys		
	FDCP-FR1	FDCP-FR3	FDCP-FR1	FDCP-FR3	
MSPRO1	-	++	NA	NA	
MSPRO2	-	++	NA	++	
MSPRO3	-	++	NA	++	
MSPRO4	-	-	NA	-	
MSPRO5	-	+	NA	+	
MSPRO6	-	-	NA	+/-	
MSPRO7	-	++	NA	+	
MSPRO8	-	+/-	NA	+/-	
MSPRO9	-	+	NA	+	
MSPRO10	-	+	NA	NA	
MSPRO11	_	+++	NA	++	
MSPRO12	-	+++	NA	+++	
MSPRO13	-	-	NA	NA	
MSPRO14	-	-	NA	NA	
LY6.3	-	-	NA	NA	

As shown in Table 6, there is an excellent agreement between the Prochon and Morphosys data. About half of the Fabs show considerable neutralizing activity, MSPRO12 being the most potent. Most of the inhibitory Fabs performed well in the binding assay (Table 4), with MSPRO11 and MSPRO12 being the exception to the rule, however, clearly remain good candidates to pursue. None of the Fabs (including those that crossreact with FGFR1) inhibited FGF-dependent FDCP-FR1 proliferation. In addition, FDCP-FR3 grown in the presence of IL3 were not affected by any of the Fabs.

An additional 20 new Fabs were selected from the second panning done at Morphosys. Three of these new Fabs (MSPRO52, MSPRO54 and MSPRO55) were subjected to the FDCP cell proliferation test and all were found to neutralize the receptor (Fig. 7A). Interestingly (and in

accord with MorphoSys affinity data), one Fab (MSPRO54) showed strong neutralizing activity against FGFR1 (Fig. 7B).

Example 7: Receptor Expression and Activation in RCJ Cells

RCJ cell assay

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RCJ cells (fetal rat calvaria-derived mesenchymal cells, RCJ 3.1C5.18; Grigoriadis, 1988) were generated to express various FGF Receptors an inducible manner, in the absence of tetracycline. The RCJ-M14 line (RCJ-FR3ach) expresses FGFR3-ach380 mutant upon induction by the removal of tetracycline. The cells were incubated in low serum after which FGF was added to stimulate receptor function and signaling. The cells were lysed and the receptor level, receptor activation and signaling are assessed by Western with anti-FGFR3 (Santa Cruz), anti-phospho-tyrosine (Promega), and anti-active ERK (or JNK) (Promega) respectively.

RCJ-M14 cells were grown in α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, 1x glutamine, 600 µg/ml neomycin, 2 µg/ml tetracycline, 50 µg/ml hygromycin B to subconfluence. The medium was aspirated off and the cells washed with trypsin, 1 ml/10 cm dish, then trypsinized with 0.5 ml/10 cm dish. The cells were resuspended in 10 ml α -MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, 1x glutamine, 600 µg/ml neomycin, and 2 µg/ml tetracycline.

Sixty thousand (6x10⁵) cells/well were seeded in a 6-well dish. Alternatively, twice that number may be seeded. The cells were washed thrice 24 hours later (or 8 hours later if twice the amount of cells are seeded) with 1 ml α-MEM, and then incubated with α-MEM supplemented with 15% fetal calf serum, 1x penicillin/streptomycin/nystatin, and 1x glutamine (induction medium) for 16 hours. Cells were washed thrice with 1 ml α-MEM and allowed to grow for 4 additional hours in 1 ml of 0.5% exhausted serum (prepared by diluting the induction medium X30 with α-MEM).

FGF9 (1 ng/ml) was added for 5 minutes and cells are then placed on ice. The cells were washed twice with ice-cold PBS and then lysed with 0.5 ml lysis buffer. The cells are scraped into an eppendorf tube, vortexed once and placed on ice for 10 minutes. The lysate was microcentrifuged 10 minutes at 4°C and the cleared lysate transferred into a fresh Eppendorf tube.

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The protein content was determined by Bradford or DC protein assay (Bio-Rad, cat# 500-0116 - see manufacture instructions). Total protein aliquots, supplemented with 1/5 volume of 5x sample buffer, were boiled for 5 minutes and stored at -20°C until ready to load on gel. In parallel an immunoprecipitation (IP) assay was performed, 10 µl anti- FGFR3 antibodies were added to the rest of the lysates and incubated for 4 hours at 4°C. 40 µl protein A-Sepharose was added and incubated for 1 hour at 4°C with continuous shaking. Afterwards, the mixture was microcentrifuged 15 seconds, and the fluid was aspirated, carefully leaving a volume of ~30 µl above the beads. The beads were washed 3 times with 1 ml lysis buffer. At this step, the protease inhibitor mix is omitted from the buffer.

- After the final wash, 15 μl of 5x sample buffer was added, samples were boiled 5 minutes and stored at -20⁰C until ready to load onto gel. Samples were loaded on 7.5% SDS-PAGE, cast on a Mini-PROTEAN II electrophoresis cell, and run at 100 V through the upper gel and at 150 V through the lower gel. Proteins were transferred onto nitrocellulose sheet using the Mini trans-blot electrophoretic transfer cell at 100 V for 75 minutes or at 15 V overnight.
- The lower part of the total lysate Western blots was probed with anti-active MAPK (ERK) and the upper part is probed with anti-phosphotyrosine, both diluted 5x10³. The IP lysate Western blots were probed with anti-anti-phosphotyrosine (R&D Systems). Hybridization was detected by ECL following the manufacturer's instructions.
- BIAcore and proliferation analyses done at MorphoSys showed that among the new Fabs,

 MS-PRO54 is highly cross reactive with FGFR1. To further test the cross reactivity of the
 new Fabs, RCJ cells expressing either FGFR3ach (RCJ-M14; M14 on figure 9A) FGFR3
 wild type (W11 on figure 9B), FGFR1 (R1-1 on figure 9C) or FGFR2 (R2-2 on figure 9D)
 were incubated with increasing amount of MS-PRO54 and MS-PRO59 for 1 hour. FGF9 was
 added for 5 minutes and cell lysates were analyzed by Western for pERK activation (Figs.
- 8A-B, 9A-9D). Figure 8A shows that MSPRO2 and MSPRO12 block FGFR3 receptor activation in W11 and RCJ-FR3ach expressing cells. Furthermore MSPRO13 was able to block FGFR1 activation while none of the Fabs blocked FGFR2 activation. Figures 8B and 9A-9D show the results of several Fabs on RCJ expressing wildtype FGFR3 (8B) or the different FGFR types. MSPRO29 appeared as the best FGFR3 blocker and was also effective in blocking FGFR1 (Fig. 9c); however, MSPRO54 was the most effective Fab against
 - FGFR1. None of the Fabs significantly inhibited FGFR2 activity. There are only a few amino acid residues, within the third Ig domain, that are shared by FGFR3 and FR1 but not by FR2.

Making mutants at these sites should clarify their role in Fab-receptor binding. Figure 8B depicts the dose effect of MS-PRO12, 29 and 13, stimulated with FGF9 and analyzed by Western blot using anti-ERK antibodies. MSPRO29 strongly inhibits FGFR3 activation (5ug), MSPRO12 has an inhibitory effect but at a higher concentration (50 ug).

5 Example 9: Epitope mapping of selected Fabs

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Constructs containing cDNAs that code for segments of the extracellular domain of FGFR3 were generated (Fig. 10). D2 comprises Ig domain 2, D2-3 comprises Ig domains 2 and 3 and D1-3 comprises Ig domains 1, 2 and 3. These include pChFR3^{D2}Fc that codes for Ig-like domain 2 of FGFR3 and pChFR3^{D2,3}Fc that codes for domain 2 and 3, both as human Fc fusions. The corresponding chimeric proteins, as well as the control hFR3exFc (containing domains 1, 2 and 3) were anchored to an ELISA plate coated with α human Fc antibody. A panel of 8 best Fabs, MSPRO2, 11, 12, 21, 24, 26, 28 and 29, were added, and bound Fab was determined with HRP-α human Fab (Fig. 11). The results in Fig. 11 demonstrate that MSPRO2 (speckled bar) and MSPRO12 (hatched bar) differ from the other tested Fabs. Both bind to the Ig like domain 2 while the others require domain 3 for binding. It was then tested whether or not Fabs that belong to the second group would distinguish the FGFR3IIIc isoform from the FGFRIIIb from. FDCP-FR3IIIb or FDCP-FR3IIIc cells were incubated in the presence of 1.25 ng/ml FGF9 with increasing doses of either MSPRO12 or MSPRO29. Ly6.3 was included as control. After 2 days in culture, cell proliferation was measured with the XTT reagent. Clearly, MSPRO29 (open triangle) was completely ineffective against the IIIb isoform (Fig. 12). In contrast, MSPRO12 (square on hatched or solid lines) was equally effective against both isoforms. These data suggest that residues that differ between the two isoform are critical for MSPRO29 (and probably also for the other Fabs in the same group) FGFR3 binding.

25 <u>Domains in FGFR3 recognized by the new Fabs.</u>

In agreement with data generated at Morphosys, MSPROs can be divided into 2 groups, one that includes Fabs that bind the FGFR3 Ig II domain (MSPRO2 and 12) and a second with members that require the Ig III domain for binding (MSPRO11, 21, 24, 26, 28, and 29). To classify the new Fabs obtained from the last screen performed at Morphosys, as well as some previously obtained Fabs, a proliferation assay of FDCP cells expressing either FR3IIIb or FR3IIIc was performed. The cells were incubated in the presence of 10 (IIIb) or 5 (IIIc)

ng/ml FGF9 with increasing doses of the indicated Fabs. After 2 days in culture, cell proliferation was measured with the XTT reagent.

In agreement with Morphosys data, MSPRO59 efficiently inhibited both FDCP-FR3IIIb (Fig. 13A) and FDCP-FR3IIIc cells (Fig. 13B) while MSPRO21, 24, 26, 28, 29 and 54 inhibited FDCP-FR3IIIc proliferation only.

Example 10: Bone culture

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Radiolabeled MSPRO29 was used to determine whether or not MSPRO Fabs can enter the bone growth plate.

To determine the effect of iodination on Fab activity, 50 μg of MSPRO29 was first labeled with cold iodine using Pierce IodoGen coated tubes. The process was carried out either without iodine, with 0.04 mM or with 1 mM NaI. MSPRO29 was then purified through a sephadex G-50 column. The ability of the modified Fab to bind FGFR3 was determined by ELISA. MaxiSorp wells were coated with anti-human Fc. FGFR3/Fc (checkered bars) was then anchored to the wells. In parallel, a similar set of wells was left in blocking buffer only (no FR3/Fc, hatched bars). The unmodified (no I) or the modified MSPRO29 (low for that labeled with 0.04 mM NaI (low) and high for that labeled at 1 mM NaI (high); 2 G-50 fractions each) were added at approximately 5 μg/well and binding was measured with anti-human Fab. Fresh MSPRO29 and buffer alone were included as controls (Fig. 14)...

MSPRO29 labeled in the presence of 0.04 mM NaI showed equal binding to the receptor as compared to the control unmodified Fab MSPRO29 labeled in the presence of 1 mM NaI (high I) also bound the receptor, however, the noise level of this sample was as high as the signal itself suggesting that at the high Iodide concentration the Fab was inactivated.

The neutralizing activity of the modified Fab was tested in a proliferation assay using FDCP-FR3 (C10) (Fig. 15). FDCP-FR3 (C10) cells were treated with the indicated amount of labeled or unlabeled (without I) MSPRO29. The proliferation rate of the cells was determined by XTT analysis. The Fab was labeled at either 0.04 mM (Low) or 1 mM NaI (High). Two G-50 fraction (I and II) were analyzed. Fresh MSPRO29 and buffer alone (mock) were included as controls.

This experiment showed that MSPRO29, labeled at 0.04 mM NaI, kept its activity almost entirely while that labeled at 1 mM NaI lost its activity completely. MS-PRO29 was labeled with 1 mCi ¹²⁵I. The specific activity of the Fab was 17 µCi/µg.

Ex vivo distribution of ¹²⁵I MSPRO29 in bone culture

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Femora prepared from newborn mice were incubated with 2 μg ¹²⁵I-MSPRO29 (17 μCi/μg) or ¹²⁵I-Ly6.3 (20 μCi/μg) for 1, 3 or 5 days in culture. Then, sections were processed for radiomicroscopy. After 3 days in culture, MSPRO29 was predominantly visualized at the higher hypertrophic zone and to a lesser extent at the secondary ossification region (Figs. 16A-16F). Hematoxylin-eosin staining of growth plate treated with radiolabelled MS-PRO29 or Ly6.3 (Figs. 16A and 16D, respectively) x100 magnification. Radiomicoscopic sections of growth plate treated with radiolabelled MS-PRO29 or Ly6.3 (Figs. 16B and 16E) at X100 magnification. Figs. 16C and 16F are the same as Figs. 16B and 16E but at x400 magnification. The arrow in figure 16C indicates the location of the specific binding of the radiolabelled MS-PRO29 to the higher hypertrophic zone of the growth plate.

As compared to MSPRO29, the control Ly6.3 Fab was weakly and evenly distributed throughout the whole growth plate. At day 1 in culture, the signal was weaker but with similar distribution pattern. This distribution also holds at 5 days in culture with a less favorable signal to noise ratio (data not shown). This clearly demonstrates that MSPRO29 binds FGFR3 in our target organ.

Example 11: Neutralizing Activity on Constitutively Activating Receptors

The inhibitory activity of MSPRO antibodies on ligand-dependent and ligand-independent FDCP proliferation expressing FGFR3 Achondroplasia mutation was tested.

- A proliferation assay was carried out using FDCP-FR3wt (C10) or FDCP-FR3ach cells incubated with 1.25 or 5 ng/ml FGF9 respectively and with increasing amounts of MSPRO54 or MSPRO59. As shown in Fig. 17, both MSPRO54 (diamond) and 59 (square) antibodies neutralize the mutant receptor. Few of the FDCP-FR3ach acquired ligand independent cell proliferation due to the high expression of the FGFR3ach mutation.
- FDCP cells that express the achondroplasia FGFR3 (FDCP-FR3ach) and proliferate independently of ligand were incubated with the indicated amount of MSPRO12, 29, 59 or the control Ly6.3. Two days later, cell proliferation was determined by an XTT analysis. When inhibition of cell proliferation by the MS-PRO 12, 29, 54 and 59 were tested, only the antibodies 12 and 59 (the only Ab which recognized D2 domain) inhibited the ligand-independent cell proliferation (Figs. 18A and 18B). Previously, the activity of MSPRO Fabs
 - independent cell proliferation (Figs. 18A and 18B). Previously, the activity of MSPRO Fabs generated in the first and second screens (MSPRO1-15 and MSPRO21-31, respectively) by XTT analysis of FDCP-FR3ach cells were tested. These cells, when generated, show ligand-

dependent proliferation. With time, however, they acquired a ligand-independent ability to proliferate. Accordingly, neutralizing Fabs were able to block the ligand-dependent, but not the ligand-independent, proliferation of these cells. To show whether this is also true for the new batch of Fabs, FDCP-FR3ach cells, which is the FDCP-dervied cell line that expresses a constitutive FGFR3-G380R (Ach), were subjected to XTT analysis in the presence of MSPRO59 and MSPRO29. Surprisingly, and in contrast to the ineffective MSPRO29 (triangle), MSPRO59 (diamond) completely blocked cell proliferation (Fig. 18B). Whether other Fabs that, like MSPRO59, bind to the second Ig like domain would also inhibit FDCP-FR3ach cell proliferation was tested next. Indeed, it was found that MSPRO12 strongly inhibits the constitutive cell proliferation. However, the third member in this family, MSPRO2, had no effect on either the constitutive or the ligand-dependent cell growth, suggesting that the Fab may have lost its neutralizing activity (not shown).

Example 12: RCS Chondrocyte Culture

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Effect of Fabs on growth arrest of RCS Chondrocytes

RCS is a rat chondrosarcoma derived cell line expressing preferentially high levels of FGFR2 and FGFR3 and low levels of FGFR1 (Sahni, 1999). In this cell line FGFR functions as an inhibitor of cell proliferation similar to its expected role in the achondroplasia phenotype. Analysis of RCS cell proliferation mediated by the addition of different molecules of the invention, showed that MSPRO54 and MSPRO59 were able to restore cell proliferation.
The screening was performed on RCS parental cells in 96 wells plates. Cells were seeded at a concentration of 2,000 cells/well. The following day 10ng/ml FGF-9 and 5μg/ml heparin were added to the cells. 50ug/ml of the antibodies were added. Positive and negative controls

were added to the cells. 50ug/ml of the antibodies were added. Positive and negative controls for cell proliferation are included in this assay at the same concentrations as the tested molecules. On the fourth day of incubation, plates were observed under the microscope. If all cells were viable, no quantitative assay to measure the effect of the variants was performed. If cell death was observed, the Cy-Quant assay kit is used to measure the amount of the cells. The results are measured in a fluoro ELISA reader. Figure 19 shows the ELISA results in bar graph form. Untraeted cells are shown speckled, ligand treated cells are shown in gray, control antibody (LY6.3)treated cells are in blak while MSPRO54 and MSPRO59 treated cells are shown in hatched otr checkered bars, respectively.

Example 13: Ex vivo Bone Culture

The femoral bone cultures were performed by excising the hind limbs of 369-mice, heterozygous or homozygous mice for the achondroplasia G369C mutation (age P0). The limbs were carefully cleaned up from the surrounding tissue (skin and muscles) and the femora exposed. The femora were removed and further cleared from tissue remains and ligaments. The femora were measured for their initial length, using a binocular with an eyepiece micrometer ruler. The bones were grown in 1 ml of medium in a 24 well tissue culture dish. The growing medium is α-MEM supplemented with penicillin (100 units/ml), streptomycin (0.1 mg/ml) and nystatin (12.5 units/ml). In addition, the medium contains BSA (0.2%), α-glycerophosphate (1 mM) and freshly prepared ascorbic acid (50 μg/ml). The bones were cultured for 15 days. Measurements of bone length and medium replacement were performed every three days.

At the end of the experiment, the growth rate of the bones was determined. The growth rate of bones is calculated from the slope of a linear regression fit on the length measurements obtained from day 3 to 9.

The results shown in Fig. 20 demonstrate a dose dependent increase in the growth rate of bones treated with MS-PRO 59 in comparison to non-relevant control LY6.3 Fab. The LY6.3-treated control femurs, marked with a circle, grew at the slowest rate. The MSPRO59 treated femurs exhibited a higher growth rate, with the optimal rate achieved at MSPRO59 concentration of 100ug/ml (square) while the higher cocentration (400ug/ml, triangle) showed inhibition. Moreover, the growth rates achieved by 400 microgram/ml of MSPRO59 doubled in comparison to the control Ab (3.55 U/day as compared to 1.88 U/day, respectively). This experiment shows the neutralizing effect of the MSPRO59 antibody on constitutively active FGFR3, in an *ex vivo* model.

25 Example 14: *In-vivo* trials

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FDCP-FR3ach cells, but not FDCP (control) cells, were found to be tumorigenic when injected into nude mice. Each of 9 mice received two sub-cutaneous injections with different amount of transfected cells. Fourteen days after injection, progressively growing tumors started to appear at the site of FDCP-FR3ach injection but not at the FDCP site of injection. External examination of the tumors showed a high vascular capsule. ¹²⁵I-labeled MSPRO59 and LY6.3 were injected I.P. into nude mice carrying the FDCP-FR3ach derived tumor. The

tumors were dissected 4 and 24 hrs later and radioactivity was measured. Concentration of MSPRO59 Abs in FDCP-FR3ach derived tumors is shown in Fig. 22.

Example 15: Animal Model for Bladder Carcinoma

Recent studies have shown that the IIIb isoform of FGFR3 is the only form expressed in bladder carcinoma, in particular an FGFR3 with an amino acid substitution wherein Serine 249 is replaced by Cysteine (S249C). The progression of the cancer is believed to be a result of the constitutive activation resulting from this amino acid substitution. In order to create the FGFR3 IIIb form, we isolated the IIIb region of FGFR3 from HeLa cells and generated a full length FGFR3IIIb isoform in pLXSN. Retroviruses, expressing either normal FGFR3 (FR3wt) or mutant FGFR3 (FR3-S249C) were produced and used to infect FDCP cells. Stable pools were generated and further used for *invitro* and *in-vivo* experiments.

A. MSPRO59 reduces tumor size in mice

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Twelve nude mice were injected with 2x10⁶ FDCP-S249C cells subcutaneous at 2 locations, one on each flank. A week later MSPRO59 was administered i.p. at 400ug per mouse (3 mice in total), followed by 3 injections of 275 ug each, in 2 to 3 days intervals. Following 24 and 26 days the tumor size was measures. Figure 23 shows the inhibitory effect of MSPRO59 on tumor size.

B. Treating FDCP-S249C-derived tumors with MSPRO59

Nude mice (3 in each group), were injected subcutaneous at 2 locations, one on each flank, with 2x10⁶ FDCP-S249C cells each. A week later, 400 or 80 µg MSPRO59 were injected IP. Three days later, mice were injected with 400 µg followed by 5 additional injections with 275 µg MSPRO59, each, every 3 or 4 days. Mice initially treated with 80 µg MSPRO59 were similarly given an additional 80 µg MSPRO59 followed by 5 injections with 50 µg MSPRO59 at the same schedule. Mice injected with PBS were used as control. Tumors typically appeared three weeks post injection of the cells. Tumor volume was estimated from measurements in 3 dimensions at 16,20, 23 or 32 days post cell injection.

As shown in Figure 24 there is both a delay in tumor appearance and an inhibitory effect on tumor progression in the treated mice. This indicates that these FGFR3 inhibitors are potent *in-vivo*.

These data may also help us understand the mechanism by which the S249C-derived tumors were developed. Since we are using pools of cells, treatment with MSPRO59 inhibited the

susceptible cells, leading to delay in tumor appearance. However, over time, the resistant cells survived and proliferated, giving rise to a solid tumor.

C. MSPRO59 inhibits FDCP-FR3ach380 derived tumor growth.

Nude mice were injected subcutaneously in the flank with 2x10⁶ FDCP-FR3ach380 cells, each. Treatment with MSPRO59 began at the day of tumor appearance. Three mice were treated with a known tyrosine kinase inhibitor (TKI -50 mg/Kg/injection) and three with 400 µg followed by 3 additional injections with 300 µg MSPRO59, every 3 or 4 days. Three mice were treated with PBS alone as control. The tumor size was estimated as before at the indicated days after cell injection. The dose schedule is shown in Table 7 below.

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Table 7

	Days After FDCP-FR3 ^{ach380} Cell Injection					
	21	25	28	31		
MSPRO59 (μg)	400µg	300µg	300µg	300µg		
PBS (µl)	50	50	50	50		

Results are shown in bar graph format in Figure 25A.

D. MSPRO59 inhibits FDCP-S249C induced tumor growth

To overcome the instability of the FDCP-derived pools, clones from each pool FDCP-S249C clone #2) were isolated and characterized. These clones were tested in an XTT proliferation assay and were shown to be inhibited by MSPRO59. 2x10⁶ cells from each clone were injected into nude mice. Tumors appeared 18-30 after injection.

FDCP-S249C clone #2 was injected subcutaneously on the flank. A week later mice were injected with 280 μg MSPRO59 single chain (SC) I.P. every day. Mice injected with PBS were used as control. Tumor volume was estimated from measurements in 3 dimensions at 18 or 24 days post cell injection. An apparent inhibition of tumor growth by MSPRO59(SC) was observed in tumors derived from clone 2 (figure 26). Figure 25B shows the inhibition effected by MSPRO59scFv and MSPRO59 Fab compared to the control. Both inhibit growth of the tumor resulting from constitutively activated cells.

Having now fully described this invention, it will be appreciated by those skilled in the art that the same can be performed within a wide range of equivalent parameters, concentrations, and conditions without departing from the spirit and scope of the invention and without undue experimentation.

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While this invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications. This application is intended to cover any variations, uses, or adaptations of the inventions following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth as follows in the scope of the appended claims.

All references cited herein, including journal articles or abstracts, published or corresponding U.S. or foreign patent applications, issued U.S. or foreign patents, or any other references, are entirely incorporated by reference herein, including all data, tables, figures, and text presented in the cited references. Additionally, the entire contents of the references cited within the references cited herein are also entirely incorporated by references.

Reference to known method steps, conventional methods steps, known methods or conventional methods is not in any way an admission that any aspect, description or embodiment of the present invention is disclosed, taught or suggested in the relevant art.

The foregoing description of the specific embodiments will so fully reveal the general nature of the invention that others can, by applying knowledge within the skill of the art (including the contents of the references cited herein), readily modify and/or adapt for various applications such specific embodiments, without undue experimentation, without departing from the general concept of the present invention. Therefore, such adaptations and modifications are intended to be within the meaning and range of equivalents of the disclosed embodiments, based on the teaching and guidance presented herein. It is to be understood that the phraseology or terminology herein is for the purpose of description and not of limitation, such that the terminology or phraseology of the present specification is to be interpreted by the skilled artisan in light of the teachings and guidance presented herein, in combination with the knowledge of one of ordinary skill in the art.

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CLAIMS

1. A molecule comprising the antigen binding portion of an isolated antibody which has specific binding affinity for a receptor protein tyrosine kinase and which blocks constitutive activation of said receptor protein tyrosine kinase.

- 2. The molecule according to claim 1, wherein said molecule binds to the extracellular domain of the receptor protein tyrosine kinase.
 - 3. The molecule according to claim 1 wherein the antibody binds the dimeric form of the receptor.
- 4. The molecule according to claim 1, wherein the receptor protein tyrosine kinase is selected from the group consisting of EGFR/ErbB1, ErbB2/HER2/Neu, ErbB/HER3, ErbB4/HER4, IGF-1R, PDGFR-α, PDGFR-β, CSF-1R, kit/SCFR, Flk2/FH3, Flk1/VEGFR1, Flk1/VEGFR2, Flt4/VEGFR3, FGFR1, FGFR2/K-SAM, FGFR3, FGFR4, TrkA, TrkC, HGFR, RON, EphA2, EphB2, EphB4, Axl, TIE/TIE1, Tek/TIE2, Ret, ROS, Alk, Ryk, DDR, LTK and MUSK, and heterodimeric combinations thereof.
 - 5. The molecule according to claim 4, wherein said receptor protein tyrosine kinase is a fibroblast growth factor receptor (FGFR).
 - 6. The molecule according to claim 5, wherein said FGFR is FGFR3.
 - 7. A pharmaceutical composition, comprising, as an active ingredient, the molecule according to any one of claims 1 through 6 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.
 - 8. A molecule comprising the antigen-binding portion of an antibody which has specific binding affinity for a fibroblast growth factor receptor (FGFR) and which blocks ligand-dependent activation of said FGFR.
- 9. The molecule according to claim 8, wherein said molecule binds to the extracellular domain of the FGFR.
 - 10. The molecule according to claim 9, wherein the FGFR is FGFR3.
 - 11. A pharmaceutical composition, comprising the molecule according to any one of claims 8-10 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.

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12. A kit comprising a molecule of any one of claims 1-6 and 8-10 and at least one reagent suitable for detecting the presence of said molecule when bound to said receptor protein tyrosine kinase and instructions for use.

- 13. A method for treatment of bone and cartilage related disorders, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 7 or 11 to a subject in need thereof.
- 14. The method according to claim 13 wherein the skeletal disorder is a skeletal dysplasia or a craniosynostosis disorder.
- 15. The method according to claim 14 wherein said craniosynostosis disorder is Muenke coronal craniosynostosis or Crouzon syndrome with acanthosis nigricans.
- 16. The method according to claim 13 wherein the skeletal dysplasia is selected from achondroplasia, thanatophoric dysplasia (TD), hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia.
- 17. The method according to claim 16, wherein the skeletal dysplasia is achondroplasia.
- 18. The method according to claim 13 for treating or inhibiting a malignant cell proliferative disease or disorder associated with abnormal RPTK activity.
- 19. The method according to claim 18 wherein the malignant cell proliferative disease or disorder is a hematopoietic malignancy.
- 20. The method according to claim 19 wherein the hematopoietic malignancy is multiple myeloma.
- 21. The method according to claim 18 for the treatment or inhibition of solid tumors.
- 22. The method according to claim 21 wherein the solid tumors are selected from mammary, colon, cervical, bladder, colorectal, chondrosarcoma or osteosarcoma.
- 23. The method according to claim 18 for treating or inhibiting tumor formation, primary tumors, tumor progression or tumor metastasis.
- 24. The method according to claim 23 wherein tumor progression is the progression of transitional cell carcinoma.

25. The method according to claim 19 wherein the disorder is associated with the action of a constitutively activated receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 7.

26. The method according to claim 19, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 11.

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- 27. The method according to claim 18 for treatment of hyperproliferative diseases and disorders associated with ligand dependent fibroblast growth factor receptor signaling.
- 28. The method according to claim 27 wherein the hyperproliferative diseases and disorders are vision disorders such as neovascular glaucoma, macular degeneration and proliferative retinopathy including diabetic retinopathy.
- 29. The method according to claim 27 wherein the hyperproliferative diseases are non-neoplastic angiogenic pathologic conditions such as hemangiomas, angiofibromas and psoriasis
 - 30. The method according to claim 18, wherein the disorder is associated with constitutive activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 7.
 - 31. The method according to claim 18, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 11.
- 32. A method for treating or inhibiting a cell proliferative disease or disorder, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 7 or 11 to a subject in need thereof.
- 33. The method according to claim 19, wherein the cell proliferative disease or disorder is tumor progression.

34. The method according to claim 20, wherein the tumor progression is the progression of transitional cell carcinoma.

- 35. The method according to claim 20, wherein the tumor progression is the progression of osteo or chondrosarcoma.
- 36. The method according to claim 20, wherein the tumor progression is the progression of multiple myeloma.

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- 37. The method according to claim 19 wherein the receptor protein tyrosine kinase is FGFR3 and the tumor progression is the progression of mammary carcinoma.
- 38. A method for screening a molecule comprising the antigen-binding portion of an antibody which blocks ligand-dependent activation of a receptor protein tyrosine kinase, comprising:

 screening a library of antibody fragments for binding to a dimeric form of a receptor protein tyrosine kinase;
 - identifying an antibody fragment which binds to the dimeric form of the receptor protein tyrosine kinase as a candidate molecule for blocking ligand-dependent activation of the receptor protein tyrosine kinase; and determining whether or not the candidate molecule can block ligand-dependent activation of the receptor protein tyrosine kinase in a cell.
 - 39. The method according to claim 38, wherein the receptor protein tyrosine kinase is a fibroblast growth factor receptor
 - 40. The method according to claim 39, wherein the fibroblast growth factor receptor is FGFR3.
 - 41. A molecule according to claim 1 comprising V_H -CDR3 and V_L -CDR3 regions, selected from the group consisting of SEQ ID NO: 8 and 9; SEQ ID NO: 12 and 13; and SEQ ID NO: 24 and 25.
 - 42. The molecule according to claim 41, comprising V_L region and V_H regions, selected from the group consisting of SEQ ID NO: 92 and 103; SEQ ID NO: 94 and 105 and SEQ ID NO: 102 and 113.
 - 43. A pharmaceutical composition, comprising, as an active ingredient, the molecule according to any one of claims 41 or 42 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.

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44. An isolated nucleic acid molecule, comprising a sequence selected from SEQ ID NO: 30, 31, 34, 35, 50 or 51 or a nucleotide sequence hybridizing under high stringency conditions thereto.

- 45. An isolated nucleic acid molecule, comprising a sequence selected from SEQ ID NO: 74, 75, 76, 84, 89 or 91 and 87or a nucleotide sequence hybridizing under high stringency conditions thereto.
- 46. The isolated nucleic acid molecule of claim 44, comprising nucleotides encoding a V_L-CDR3 DNA region and a V_H-CDR3 DNA region, respectively, selected from the group consisting of SEQ ID NO: 30 and 31; SEQ ID NO:34 and 35; SEQ ID NO: 50 and 51.
- 47. The isolated nucleic acid molecule of claim 45 comprising nucleotides encoding a V_L region and a V_H region, respectively, selected from the group consisting of SEQ ID NO: 74 and 84; SEQ ID NO:75 and 89; and SEQ ID NO: 76 and 91.
- 48. A vector comprising a nucleic acid molecule according to claim 46 or 47.
- 49. The vector according to claim 48 wherein the vector is an expression vector.
- 50. The host cell transformed with the vector according to claim 48 or 49.
- 51. A molecule according to claim 8 comprising the combination of V_H-CDR3 and V_L-CDR3 amino acid sequences selected from the group consisting of SEQ ID NO:10 and SEQ ID NO:11; SEQ ID NO:14 and SEQ ID NO:15; SEQ ID NO:16 and SEQ ID NO:17; SEQ ID NO:18 and SEQ ID NO:19; SEQ ID NO:20 and SEQ ID NO:21; SEQ ID NO:22 and SEQ ID NO:23, SEQ ID NO:26 and SEQ ID NO:27 or SEQ ID NO:28 and SEQ ID NO:29.
- 52. The molecule according to claim 51, comprising a V_L region and a V_H region, respectively, selected from the group consisting of respectively, selected from the group consisting of SEQ ID NO: 92 and 103; SEQ ID NO: 93 and 104; SEQ ID NO: 94 and 105; SEQ ID NO:95 and 106; SEQ ID NO: 96 and 107; SEQ ID NO: 97 and 108; SEQ ID NO:98 and 109; SEQ ID NO: 99 and 110; SEQ ID NO: 100 and 111; SEQ ID NO: 101 and 112; and SEQ ID NO:102 and 113.
- 53. A pharmaceutical composition, comprising the molecule according to any one of claims 51-52 and a pharmaceutically acceptable carrier, excipient, or auxiliary agent.

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54. An isolated nucleic acid molecule, comprising SEQ ID NO: NO:32, 33, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 48, and 49 or a nucleotide sequence hybridizing under high stringency conditions thereto.

- 55. An isolated nucleic acid molecule, comprising SEQ ID NO: 62, 64, 65, 67, 69, 70, 76, 78, 79, 80, 83, 85, 86, and 87or a nucleotide sequence hybridizing under high stringency conditions thereto.
- 56. An isolated nucleic acid molecule, comprising nucleotides encoding a V_L -CDR3 DNA region and a V_H -CDR3 DNA region, respectively, selected from the group consisting of SEQ ID NO: 32 and 33; SEQ ID NO:36 and 37; SEQ ID NO: 38 and 39, SEQ ID NO:40 and 41, SEQ ID NO: 42 and 43, SEQ ID NO: 44 and 45, SEQ ID NO: 48 and 49.
- 57. An isolated nucleic acid molecule, comprising nucleotides encoding a V_L region and a V_H region, respectively, selected from the group consisting of SEQ ID NO: 70 and 85; SEQ ID NO:67 and 78; SEQ ID NO:64 and 79; SEQ ID NO:71 and 86; SEQ ID NO:62 and 80; SEQ ID NO:65 and 87; SEQ ID NO:69 and 83.
- 58. A vector comprising a nucleic acid molecule according to claim 56 or 57.
- 59. The vector according to claim 58 which is an expression vector.
- 60. The host cell transformed with the vector according claim 58 or 59.
- 61. A method for treatment of bone and cartilage related disorders, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 43 or 53 to a subject in need thereof.
- 62. The method according to claim 61 wherein the skeletal disorder is a skeletal dysplasia or a craniosynostosis disorder.
- 63. The method according to claim 62 wherein said craniosynostosis disorder is

 Muenke coronal craniosynostosis or Crouzon syndrome with acanthosis nigricans.
- 64. The method according to claim 63 wherein the skeletal dysplasia is selected from achondroplasia, thanatophoric dysplasia (TD), hypochondroplasia, severe achondroplasia with developmental delay and acanthosis nigricans (SADDAN) dysplasia.
- 30 65. The method according to claim 64, wherein the skeletal dysplasia is achondroplasia.

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66. The method according to claim 61 for treating or inhibiting a malignant cell proliferative disease or disorder associated with abnormal RPTK activity.

- 67. The method according to claim 66 wherein the malignant cell proliferative disease or disorder is a hematopoietic malignancy.
- 68. The method according to claim 67 wherein the hematopoietic malignancy is multiple myeloma.
- 69. The method according to claim 61 for the treatment or inhibition of solid tumors.
- 70. The method according to claim 69 wherein the solid tumors are selected from mammary, colon, cervical, bladder, colorectal, chondrosarcoma or osteosarcoma.
- 71. The method according to claim 61 for treating or inhibiting tumor formation, primary tumors, tumor progression or tumor metastasis.
 - 72. The method according to claim 71 wherein tumor progression is the progression of transitional cell carcinoma.
 - 73. The method according to claim 61 wherein the disorder is associated with the action of a constitutively activated receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 43.
 - 74. The method according to claim 61, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 53.
 - 75. The method according to claim 74 for treatment of hyperproliferative diseases and disorders associated with ligand dependent fibroblast growth factor receptor signaling.
 - 76. The method according to claim 75 wherein the hyperproliferative diseases and disorders are vision disorders such as neovascular glaucoma, macular degeneration and proliferative retinopathy including diabetic retinopathy.
 - 77. The method according to claim 75 wherein the hyperproliferative diseases are non-neoplastic angiogenic pathologic conditions such as hemangiomas, angiofibromas and psoriasis

78. The method according to claim 73, wherein the disorder is associated with constitutive activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 43.

5 79. The method according to claim 74, wherein the disorder is associated with ligand-dependent activation of a receptor protein tyrosine kinase, and wherein the administered pharmaceutical composition is the pharmaceutical composition according to claim 53.

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- 80. A method for treating or inhibiting a cell proliferative disease or disorder, comprising administering a therapeutically effective amount of the pharmaceutical composition according to claim 43 or 53 to a subject in need thereof.
- 81. The method according to claim 80, wherein the cell proliferative disease or disorder is tumor progression.
- 82. The method according to claim 81, wherein the tumor progression is the progression of transitional cell carcinoma.
- 83. The method according to claim 81, wherein the tumor progression is the progression of osteo or chondrosarcoma.
- 84. The method according to claim 81, wherein the tumor progression is the progression of multiple myeloma.
- 85. The method according to claim 81 wherein the receptor protein tyrosine kinase is FGFR3 and the tumor progression is the progression of mammary carcinoma.

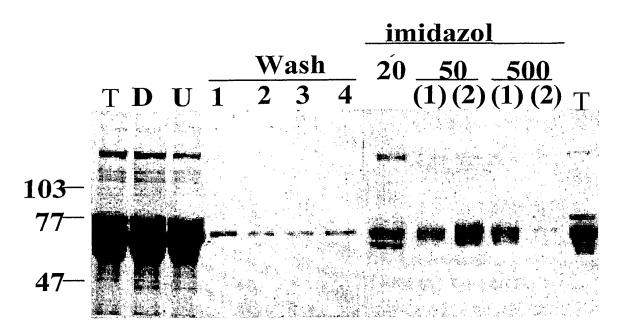
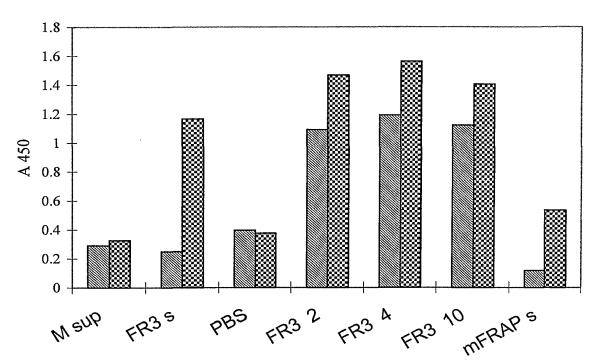
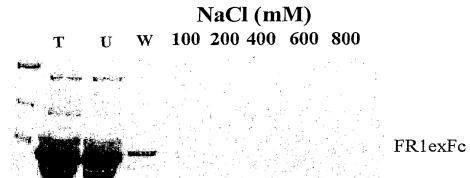


Figure 1

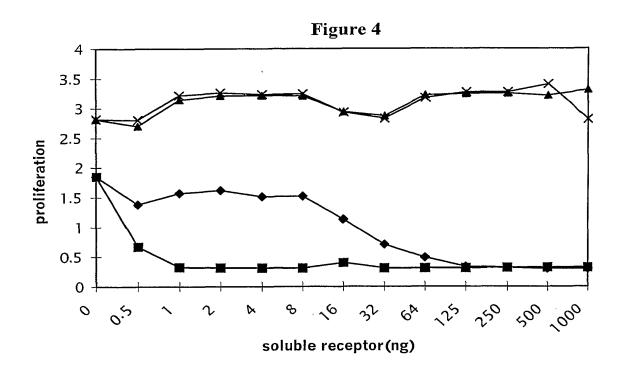


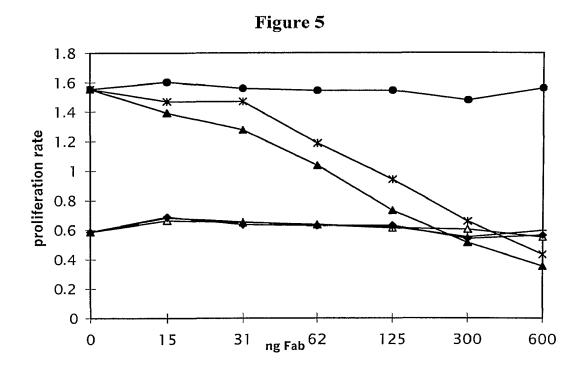


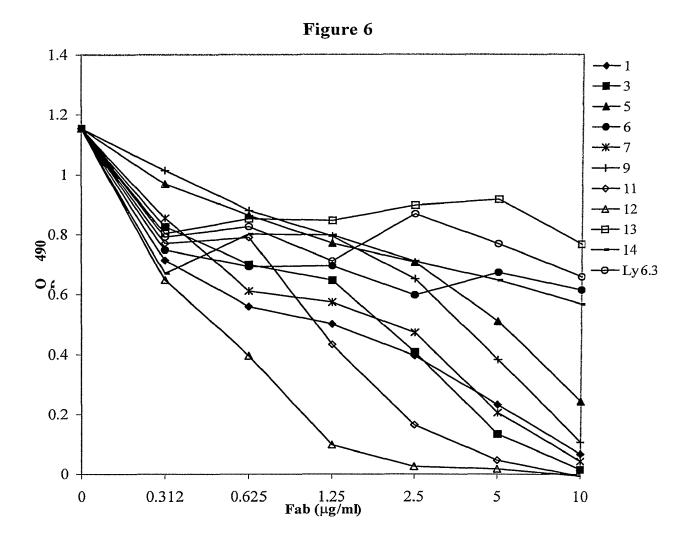


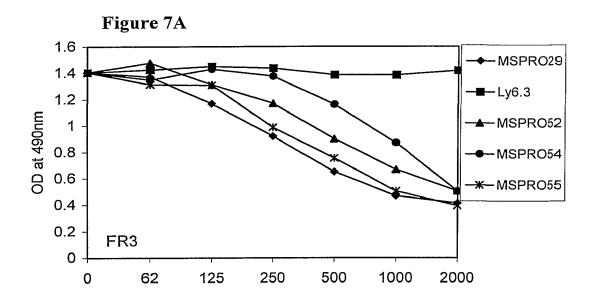












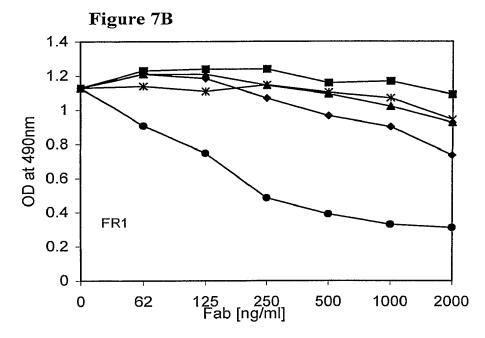


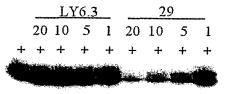
Figure 8A

Figure 8B

FIGURE 9A

<u>M14</u>

MSPRO (μg) F9



59 54 20 10 5 1 20 10 5 1 - + + + + + + + +

FIGURE 9B Wll

MSPRO (μg) F9

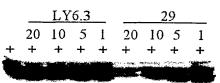


FIGURE 9C R1-1

MSPRO (μg) F9

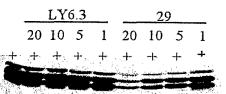


FIGURE 9D R2-2

MSPRO (μg) F9

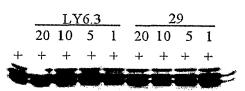
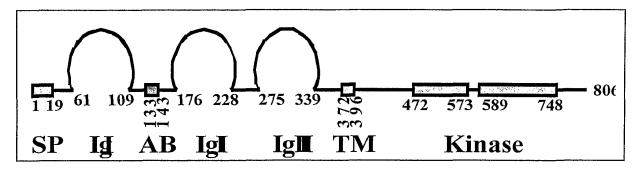


Figure 10

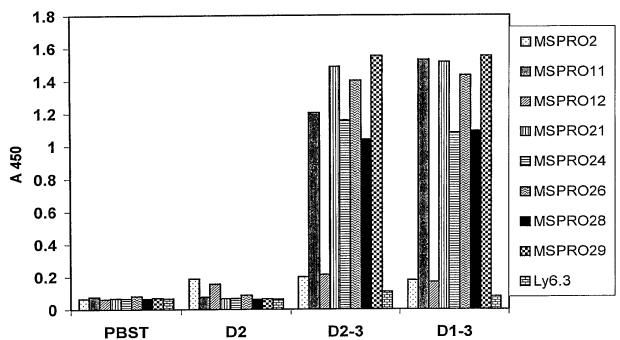
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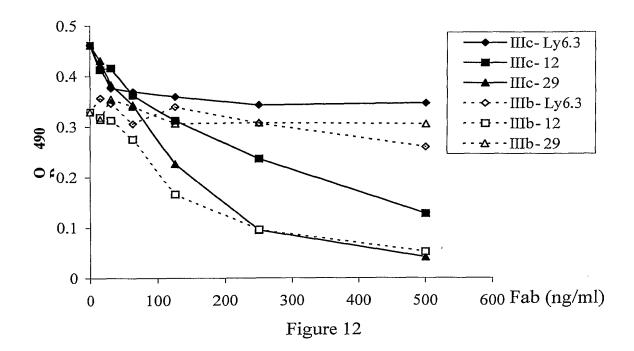
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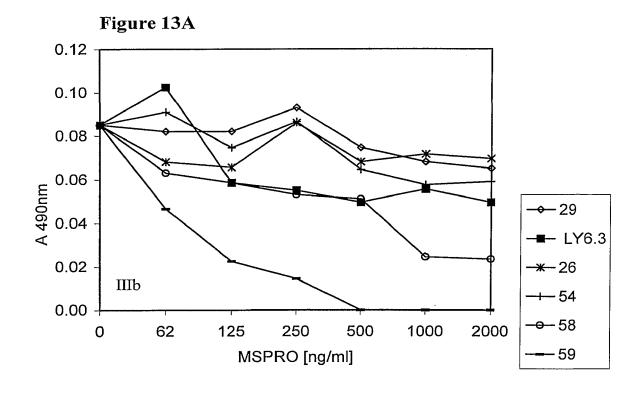
D2 _____

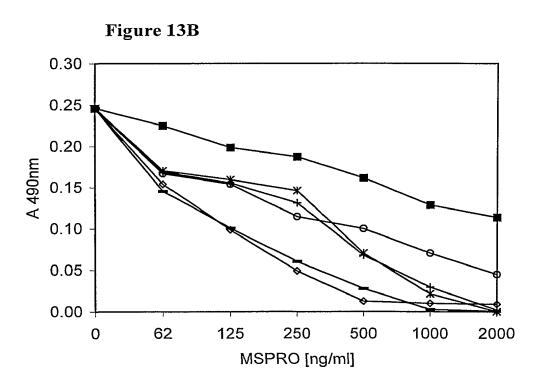






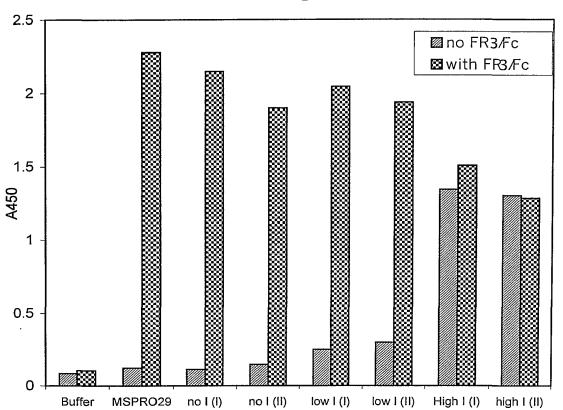




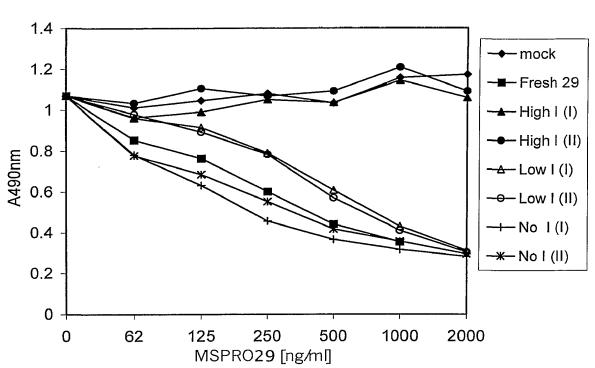


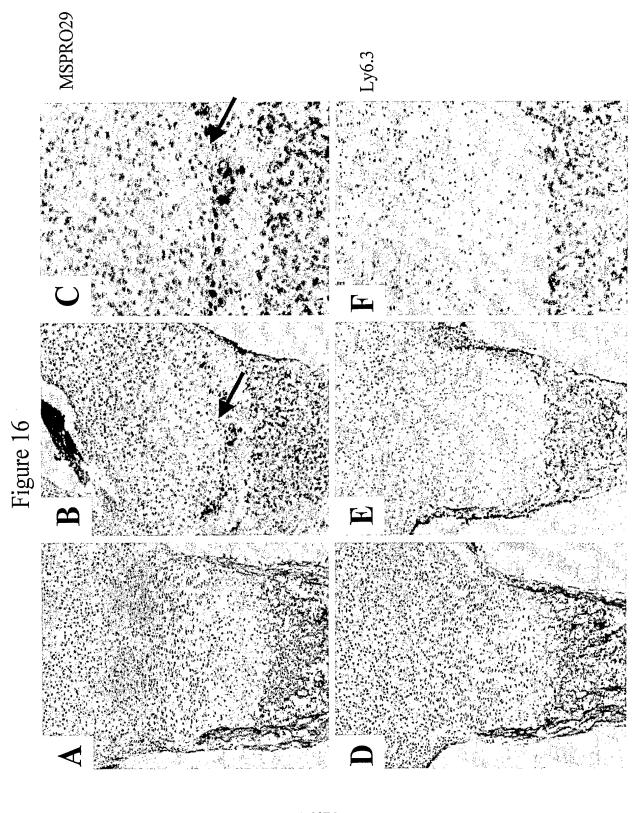
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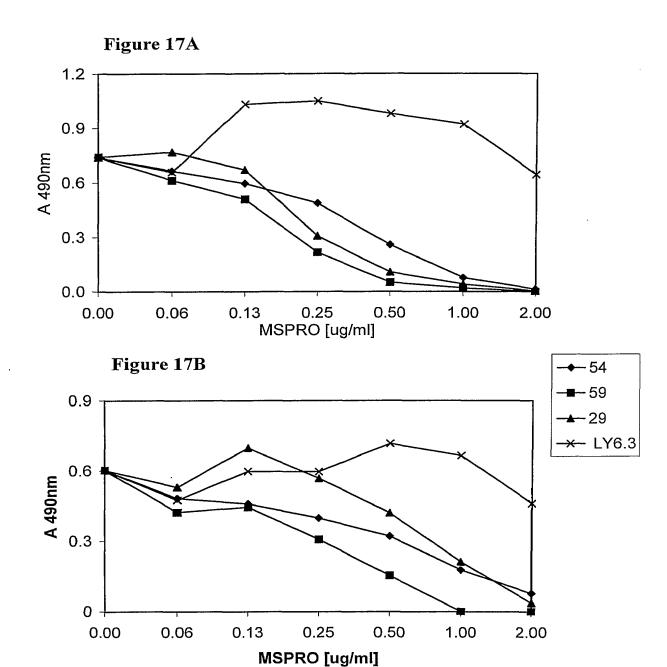




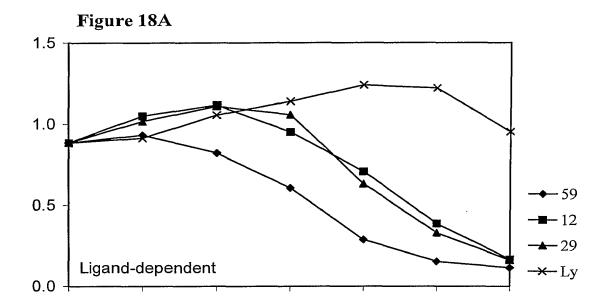


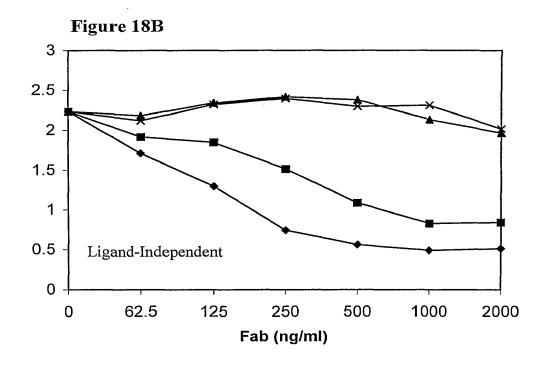


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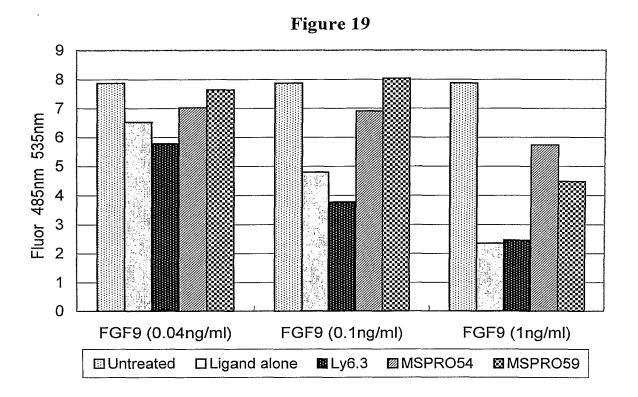


17/50
SUBSTITUTE SHEET (RULE 26)



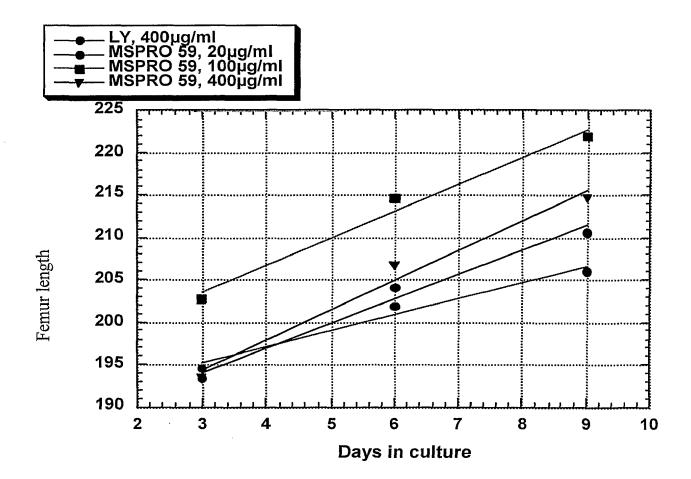


18/50 SUBSTITUTE SHEET (RULE 26)



19/50
SUBSTITUTE SHEET (RULE 26)

Figure 20



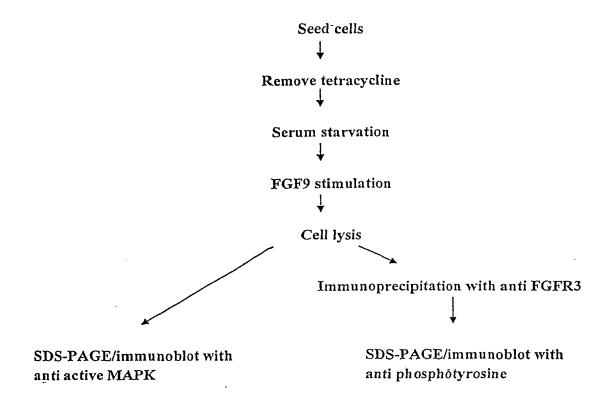


FIG.21

Figure 22

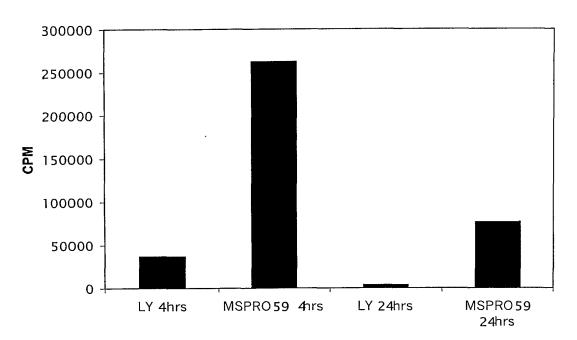
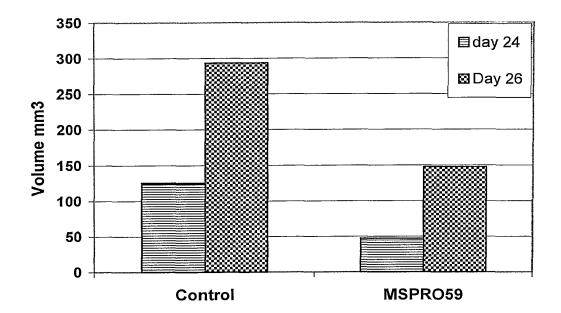
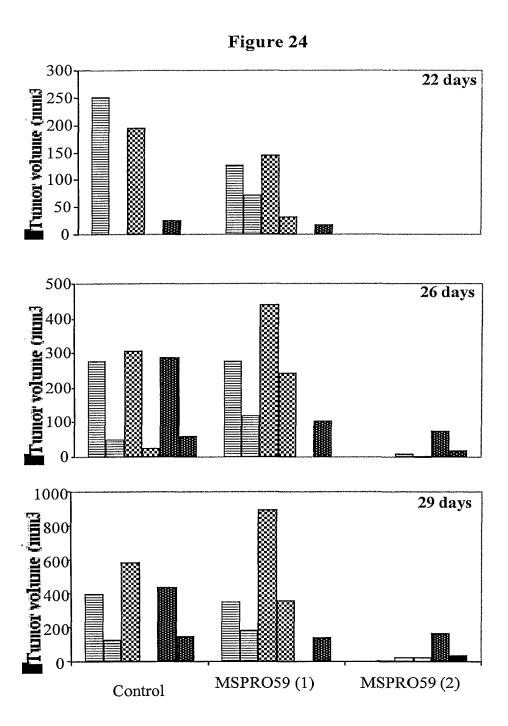


Figure 23

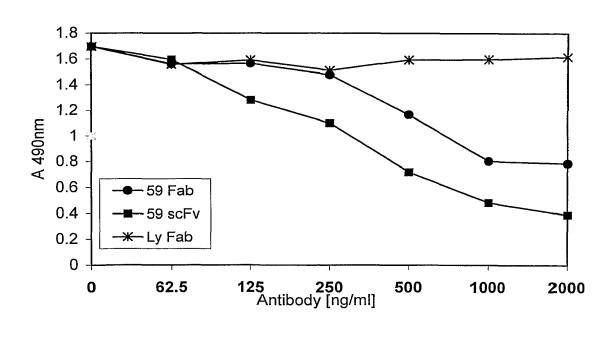




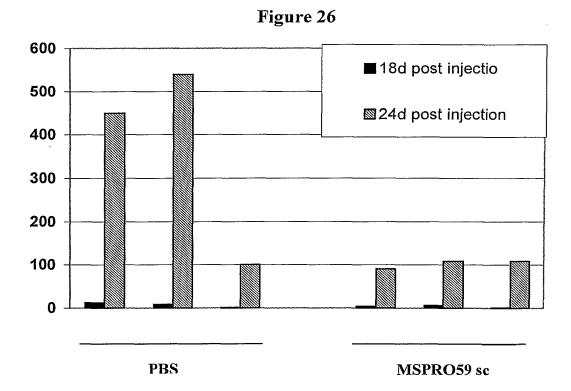
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Figure 25A 1600 ← Control 1 1400 -□- Control 2 Tumor volume mm 3 1200 1000 ---MSPRO59 1 800 ▲ MSPRO59 2 600 **●**−MSPRO59 3 400 200 0 25 Days after cell injection 31 21

Figure 25B

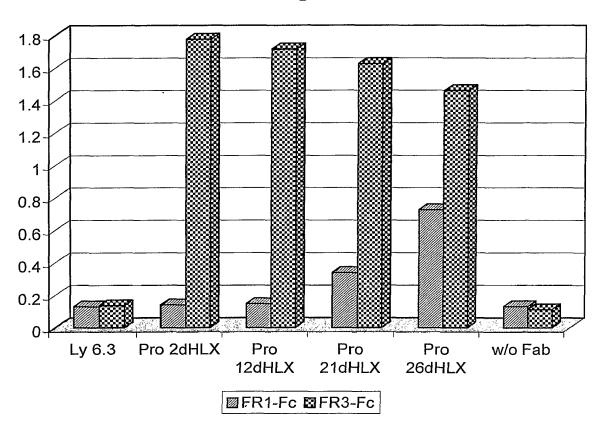


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Figure 28A

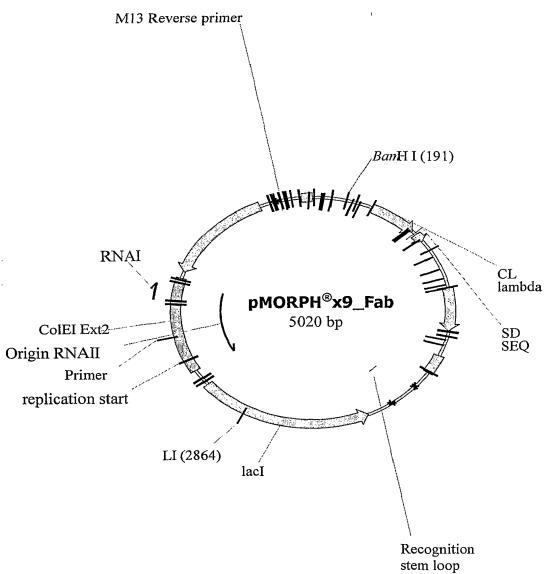


Figure 28B

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5	651 TAGGAGAAAA TAAAATGAAA CAAAGCACTA TTGCACTGGC ACTCTTACCG ATCCTCTTTT ATTTTACTTT GTTTCGTGAT AACGTGACCG TGAGAATGGC
	MfeI
10	701 TTGCTCTTCA CCCCTGTTAC CAAAGCCCAG GTGCAATTGA AAGAAAGCGG AACGAGAAGT GGGGACAATG GTTTCGGGTC CACGTTAACT TTCTTTCGCC
	BspEI
15	751 CCCGGCCCTG GTGAAACCGA CCCAAACCCT GACCCTGACC TGTACCTTTT GGGCCGGGAC CACTTTGGCT GGGTTTGGGA CTGGGACTGG ACATGGAAAA
	BspEI
20	801 CCGGATTTAG CCTGTCCACG TCTGGCGTTG GCGTGGGCTG GATTCGCCAG GGCCTAAATC GGACAGGTGC AGACCGCAAC CGCACCCGAC CTAAGCGGTC
	XhoI
25	AvaI
25	851 CCGCCTGGGA AAGCCCTCGA GTGGCTGGCT CTGATTGATT GGGATGATGA GGCGGACCCT TTCGGGAGCT CACCGACCGA GACTAACTAA CCCTACTACT
30	901 TAAGTATTAT AGCACCAGCC TGAAAACGCG TCTGACCATT AGCAAAGATA ATTCATAATA TCGTGGTCGG ACTTTTGCGC AGACTGGTAA TCGTTTCTAT
	BstBI
35	SfuI
55	NspV
40	951 CTTCGAAAAA TCAGGTGGTG CTGACTATGA CCAACATGGA CCCGGTGGAT GAAGCTTTTT AGTCCACCAC GACTGATACT GGTTGTACCT GGGCCACCTA
10	BssHII
45	1001 ACGGCCACCT ATTATTGCGC GCGTTCTCCT CGTTATCGTG GTGCTTTTGA TGCCGGTGGA TAATAACGCG CGCAAGAGGA GCAATAGCAC CACGAAAACT
.5	BlpI
	Styl CelII
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55	1101 GTCCAAGCGT GTTTCCGCTG GCTCCGAGCA GCAAAAGCAC CAGCGGCGGC CAGGTTCGCA CAAAGGCGAC CGAGGCTCGT CGTTTTCGTG GTCGCCGCCG
55	1151 ACGGCTGCCC TGGGCTGCCT GGTTAAAGAT TATTTCCCGG AACCAGTCAC TGCCGACGGG ACCCGACGGA CCAATTTCTA ATAAAGGGCC TTGGTCAGTG

WO 02/102972 PCT/IL02/00494 1201 CGTGAGCTGG AACAGCGGGG CGCTGACCAG CGGCGTGCAT ACCTTTCCGG GCACTCGACC TTGTCGCCCC GCGACTGGTC GCCGCACGTA TGGAAAGGCC 1251 CGGTGCTGCA AAGCAGCGGC CTGTATAGCC TGAGCAGCGT TGTGACCGTG 5 GCCACGACGT TTCGTCGCCG GACATATCGG ACTCGTCGCA ACACTGGCAC 1301 CCGAGCAGCA GCTTAGGCAC TCAGACCTAT ATTTGCAACG TGAACCATAA GGCTCGTCGT CGAATCCGTG AGTCTGGATA TAAACGTTGC ACTTGGTATT 10 EcoRI 1351 ACCGAGCAAC ACCAAAGTGG ATAAAAAAGT GGAACCGAAA AGCGAATTCG TGGCTCGTTG TGGTTTCACC TATTTTTCA CCTTGGCTTT TCGCTTAAGC 15 BssHII 1401 ACTATAAAGA TGACGATGAC AAAGGCGCGC CGTGGAGCCA CCCGCAGTTT TGATATTTCT ACTGCTACTG TTTCCGCGCG GCACCTCGGT GGGCGTCAAA 20 HindIII 1451 GAAAAATGAT AAGCTTGACC TGTGAAGTGA AAAATGGCGC AGATTGTGCG CTTTTTACTA TTCGAACTGG ACACTTCACT TTTTACCGCG TCTAACACGC OGIII3 100.0% 25 _____ TGTAAAAAA ACAGACGGCA AATTAATTTC CCCCCCCCC CGGCCGGACC 1551 GGGGGGGTGT ACATGAAATT GTAAACGTTA ATATTTTGTT AAAATTCGCG 30 CCCCCCACA TGTACTTTAA CATTTGCAAT TATAAAACAA TTTTAAGCGC 1601 TTAAATTTTT GTTAAATCAG CTCATTTTTT AACCAATAGG CCGAAATCGG AATTTAAAAA CAATTTAGTC GAGTAAAAAA TTGGTTATCC GGCTTTAGCC 35 1651 CAAAATCCCT TATAAATCAA AAGAATAGAC CGAGATAGGG TTGAGTGTTG GTTTTAGGGA ATATTTAGTT TTCTTATCTG GCTCTATCCC AACTCACAAC 1701 TTCCAGTTTG GAACAAGAGT CCACTATTAA AGAACGTGGA CTCCAACGTC 40 AAGGTCAAAC CTTGTTCTCA GGTGATAATT TCTTGCACCT GAGGTTGCAG 1751 AAAGGCCAA AAACCGTCTA TCAGGGCGAT GGCCCACTAC GAGAACCATC TTTCCCGCTT TTTGGCAGAT AGTCCCGCTA CCGGGTGATG CTCTTGGTAG 45 1801 ACCCTAATCA AGTTTTTTGG GGTCGAGGTG CCGTAAAGCA CTAAATCGGA TGGGATTAGT TCAAAAAACC CCAGCTCCAC GGCATTTCGT GATTTAGCCT 1851 ACCCTAAAGG GAGCCCCCGA TTTAGAGCTT GACGGGGAAA GCCGGCGAAC TGGGATTTCC CTCGGGGGCT AAATCTCGAA CTGCCCCTTT CGGCCGCTTG

31/50

1901 GTGGCGAGAA AGGAAGGGAA GAAAGCGAAA GGAGCGGGCG CTAGGGCGCT CACCGCTCTT TCCTTCCCTT CTTTCGCTTT CCTCGCCCGC GATCCCGCGA

1951 GGCAAGTGTA GCGGTCACGC TGCGCGTAAC CACCACACCC GCCGCGCTTA

CCGTTCACAT CGCCAGTGCG ACGCGCATTG GTGGTGTGGG CGGCGCGAAT

50

55

	2001 ATGCGCCGCT ACAGGGCGCG TGCTAGACTA GTGTTTAAAC CGGACCGGGG TACGCGGCGA TGTCCCGCGC ACGATCTGAT CACAAATTTG GCCTGGCCCC
5	2051 GGGGGCTTAA GTGGGCTGCA AAACAAAACG GCCTCCTGTC AGGAAGCCGC CCCCGAATT CACCCGACGT TTTGTTTTGC CGGAGGACAG TCCTTCGGCG
	2101 TTTTATCGGG TAGCCTCACT GCCCGCTTTC CAGTCGGGAA ACCTGTCGTG AAAATAGCCC ATCGGAGTGA CGGGCGAAAG GTCAGCCCTT TGGACAGCAC
10	2151 CCAGCTGCAT CAGTGAATCG GCCAACGCGC GGGGAGAGGC GGTTTGCGTA GGTCGACGTA GTCACTTAGC CGGTTGCGCG CCCCTCTCCG CCAAACGCAT
.	2201 TTGGGAGCCA GGGTGGTTTT TCTTTTCACC AGTGAGACGG GCAACAGCTG AACCCTCGGT CCCACCAAAA AGAAAAGTGG TCACTCTGCC CGTTGTCGAC
15	2251 ATTGCCCTTC ACCGCCTGGC CCTGAGAGAG TTGCAGCAAG CGGTCCACGC TAACGGGAAG TGGCGGACCG GGACTCTCTC AACGTCGTTC GCCAGGTGCG
20	2301 TGGTTTGCCC CAGCAGGCGA AAATCCTGTT TGATGGTGGT CAGCGGCGGG ACCAAACGGG GTCGTCCGCT TTTAGGACAA ACTACCACCA GTCGCCGCCC
	2351 ATATAACATG AGCTGTCCTC GGTATCGTCG TATCCCACTA CCGAGATGTC TATATTGTAC TCGACAGGAG CCATAGCAGC ATAGGGTGAT GGCTCTACAG
25	2401 CGCACCAACG CGCAGCCCGG ACTCGGTAAT GGCACGCATT GCGCCCAGCG GCGTGGTTGC GCGTCGGGCC TGAGCCATTA CCGTGCGTAA CGCGGGTCGC
	2451 CCATCTGATC GTTGGCAACC AGCATCGCAG TGGGAACGAT GCCCTCATTC GGTAGACTAG CAACCGTTGG TCGTAGCGTC ACCCTTGCTA CGGGAGTAAG
30	2501 AGCATTTGCA TGGTTTGTTG AAAACCGGAC ATGGCACTCC AGTCGCCTTC TCGTAAACGT ACCAAACAAC TTTTGGCCTG TACCGTGAGG TCAGCGGAAG
35	2551 CCGTTCCGCT ATCGGCTGAA TTTGATTGCG AGTGAGATAT TTATGCCAGC GGCAAGGCGA TAGCCGACTT AAACTAACGC TCACTCTATA AATACGGTCG
	2601 CAGCCAGACG CAGACGCGCC GAGACAGAAC TTAATGGGCC AGCTAACAGC GTCGGTCTGC GTCTGCGCGG CTCTGTCTTG AATTACCCGG TCGATTGTCG
40	2651 GCGATTTGCT GGTGGCCCAA TGCGACCAGA TGCTCCACGC CCAGTCGCGT CGCTAAACGA CCACCGGGTT ACGCTGGTCT ACGAGGTGCG GGTCAGCGCA
4.5	2701 ACCGTCCTCA TGGGAGAAAA TAATACTGTT GATGGGTGTC TGGTCAGAGA TGGCAGGAGT ACCCTCTTTT ATTATGACAA CTACCCACAG ACCAGTCTCT
45	2751 CATCAAGAAA TAACGCCGGA ACATTAGTGC AGGCAGCTTC CACAGCAATA GTAGTTCTTT ATTGCGGCCT TGTAATCACG TCCGTCGAAG GTGTCGTTAT
50	2801 GCATCCTGGT CATCCAGCGG ATAGTTAATA ATCAGCCCAC TGACACGTTG CGTAGGACCA GTAGGTCGCC TATCAATTAT TAGTCGGGTG ACTGTGCAAC
	ApaLI
55	2851 CGCGAGAAGA TTGTGCACCG CCGCTTTACA GGCTTCGACG CCGCTTCGTT GCGCTCTTCT AACACGTGGC GGCGAAATGT CCGAAGCTGC GGCGAAGCAA
	2901 CTACCATCGA CACGACCACG CTGGCACCCA GTTGATCGGC GCGAGATTTA 32/50
	Jai Ju

	GATGGTAGCT GTGCTGGTGC GACCGTGGGT CAACTAGCCG CGCTCTAAAT
5	2951 ATCGCCGCGA CAATTTGCGA CGGCGCGTGC AGGGCCAGAC TGGAGGTGG TAGCGGCGCT GTTAAACGCT GCCGCGCACG TCCCGGTCTG ACCTCCACCG
	3001 AACGCCAATC AGCAACGACT GTTTGCCCGC CAGTTGTTGT GCCACGCGGT TTGCGGTTAG TCGTTGCTGA CAAACGGGCG GTCAACAACA CGGTGCGCCA
10	3051 TAGGAATGTA ATTCAGCTCC GCCATCGCCG CTTCCACTTT TTCCCGCGTT ATCCTTACAT TAAGTCGAGG CGGTAGCGGC GAAGGTGAAA AAGGGCGCAA
	3101 TTCGCAGAAA CGTGGCTGGC CTGGTTCACC ACGCGGGAAA CGGTCTGATA AAGCGTCTTT GCACCGACCG GACCAAGTGG TGCGCCCTTT GCCAGACTAT
15	3151 AGAGACACCG GCATACTCTG CGACATCGTA TAACGTTACT GGTTTCACAT TCTCTGTGGC CGTATGAGAC GCTGTAGCAT ATTGCAATGA CCAAAGTGTA
20	3201 TCACCACCCT GAATTGACTC TCTTCCGGGC GCTATCATGC CATACCGCGA AGTGGTGGGA CTTAACTGAG AGAAGGCCCG CGATAGTACG GTATGGCGCT
	3251 AAGGTTTTGC GCCATTCGAT GCTAGCCATG TGAGCAAAAG GCCAGCAAAA TTCCAAAACG CGGTAAGCTA CGATCGGTAC ACTCGTTTTC CGGTCGTTTT
25	3301 GGCCAGGAAC CGTAAAAAGG CCGCGTTGCT GGCGTTTTTC CATAGGCTCC CCGGTCCTTG GCATTTTTCC GGCGCAACGA CCGCAAAAAG GTATCCGAGG
	3351 GCCCCCTGA CGAGCATCAC AAAAATCGAC GCTCAAGTCA GAGGTGGCGA CGGGGGGACT GCTCGTAGTG TTTTTAGCTG CGAGTTCAGT CTCCACCGCT
30	3401 AACCCGACAG GACTATAAAG ATACCAGGCG TTTCCCCCTG GAAGCTCCCT TTGGGCTGTC CTGATATTTC TATGGTCCGC AAAGGGGGAC CTTCGAGGGA
	3451 CGTGCGCTCT CCTGTTCCGA CCCTGCCGCT TACCGGATAC CTGTCCGCCT GCACGCGAGA GGACAAGGCT GGGACGGCGA ATGGCCTATG GACAGGCGGA
35	3501 TTCTCCCTTC GGGAAGCGTG GCGCTTTCTC ATAGCTCACG CTGTAGGTAT AAGAGGGAAG CCCTTCGCAC CGCGAAAGAG TATCGAGTGC GACATCCATA
40	ApaLI
40	3551 CTCAGTTCGG TGTAGGTCGT TCGCTCCAAG CTGGGCTGTG TGCACGAACC GAGTCAAGCC ACATCCAGCA AGCGAGGTTC GACCCGACAC ACGTGCTTGG
	3601 CCCCGTTCAG CCCGACCGCT GCGCCTTATC CGGTAACTAT CGTCTTGAGT

3601 CCCCGTTCAG CCCGACCGCT GCGCCTTATC CGGTAACTAT CGTCTTGAGT GGGGCAAGTC GGGCTGGCGA CGCGGAATAG GCCATTGATA GCAGAACTCA

45

50

55

3651 CCAACCGGT AAGACACGAC TTATCGCCAC TGGCAGCAGC CACTGGTAAC GGTTGGGCCA TTCTGTGCTG AATAGCGGTG ACCGTCGTCG GTGACCATTG

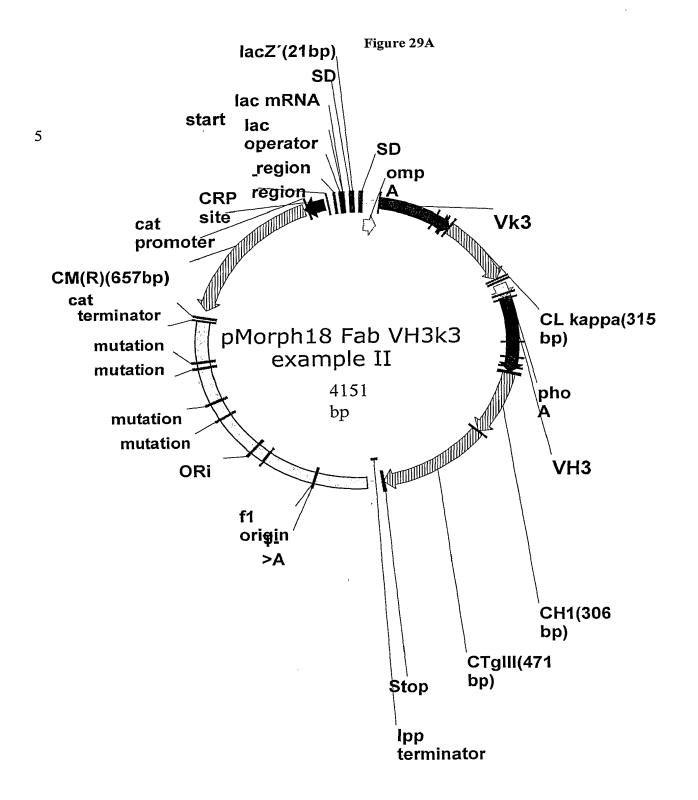
3701 AGGATTAGCA GAGCGAGGTA TGTAGGCGGT GCTACAGAGT TCTTGAAGTG TCCTAATCGT CTCGCTCCAT ACATCCGCCA CGATGTCTCA AGAACTTCAC

3751 GTGGCCTAAC TACGGCTACA CTAGAAGAAC AGTATTTGGT ATCTGCGCTC CACCGGATTG ATGCCGATGT GATCTTCTTG TCATAAACCA TAGACGCGAG

3801 TGCTGTAGCC AGTTACCTTC GGAAAAAGAG TTGGTAGCTC TTGATCCGGC ACGACATCGG TCAATGGAAG CCTTTTTCTC AACCATCGAG AACTAGGCCG

	3851 AAACAAACCA CCGCTGGTAG CGGTGGTTTT TTTGTTTGCA AGCAGCAGAT TTTGTTTGGT GGCGACCATC GCCACCAAAA AAACAAACGT TCGTCGTCTA
5	3901 TACGCGCAGA AAAAAAGGAT CTCAAGAAGA TCCTTTGATC TTTTCTACGG ATGCGCGTCT TTTTTTCCTA GAGTTCTTCT AGGAAACTAG AAAAGATGCC
10	3951 GGTCTGACGC TCAGTGGAAC GAAAACTCAC GTTAAGGGAT TTTGGTCAGA CCAGACTGCG AGTCACCTTG CTTTTGAGTG CAATTCCCTA AAACCAGTCT
10	4001 TCTAGCACCA GGCGTTTAAG GGCACCAATA ACTGCCTTAA AAAAATTACG AGATCGTGGT CCGCAAATTC CCGTGGTTAT TGACGGAATT TTTTTAATGC
15	4051 CCCCGCCCTG CCACTCATCG CAGTACTGTT GTAATTCATT AAGCATTCTG GGGCCGGGAC GGTGAGTAGC GTCATGACAA CATTAAGTAA TTCGTAAGAC
	4101 CCGACATGGA AGCCATCACA AACGGCATGA TGAACCTGAA TCGCCAGCGG GGCTGTACCT TCGGTAGTGT TTGCCGTACT ACTTGGACTT AGCGGTCGCC
20	4151 CATCAGCACC TTGTCGCCTT GCGTATAATA TTTGCCCATA GTGAAAACGG GTAGTCGTGG AACAGCGGAA CGCATATTAT AAACGGGTAT CACTTTTGCC
25	4201 GGGCGAAGAA GTTGTCCATA TTGGCTACGT TTAAATCAAA ACTGGTGAAA CCCGCTTCTT CAACAGGTAT AACCGATGCA AATTTAGTTT TGACCACTTT
23	4251 CTCACCCAGG GATTGGCTGA GACGAAAAAC ATATTCTCAA TAAACCCTTT GAGTGGGTCC CTAACCGACT CTGCTTTTTG TATAAGAGTT ATTTGGGAAA
30	4301 AGGGAAATAG GCCAGGTTTT CACCGTAACA CGCCACATCT TGCGAATATA TCCCTTTATC CGGTCCAAAA GTGGCATTGT GCGGTGTAGA ACGCTTATAT
	4351 TGTGTAGAAA CTGCCGGAAA TCGTCGTGGT ATTCACTCCA GAGCGATGAA ACACATCTTT GACGGCCTTT AGCAGCACCA TAAGTGAGGT CTCGCTACTT
35	4401 AACGTTTCAG TTTGCTCATG GAAAACGGTG TAACAAGGGT GAACACTATC TTGCAAAGTC AAACGAGTAC CTTTTGCCAC ATTGTTCCCA CTTGTGATAG
40	4451 CCATATCACC AGCTCACCGT CTTTCATTGC CATACGGAAC TCCGGGTGAG GGTATAGTGG TCGAGTGGCA GAAAGTAACG GTATGCCTTG AGGCCCACTC
TV	4501 CATTCATCAG GCGGGCAAGA ATGTGAATAA AGGCCGGATA AAACTTGTGC GTAAGTAGTC CGCCCGTTCT TACACTTATT TCCGGCCTAT TTTGAACACG
45	4551 TTATTTTCT TTACGGTCTT TAAAAAGGCC GTAATATCCA GCTGAACGGT AATAAAAAGA AATGCCAGAA ATTTTTCCGG CATTATAGGT CGACTTGCCA
	4601 CTGGTTATAG GTACATTGAG CAACTGACTG AAATGCCTCA AAATGTTCTT GACCAATATC CATGTAACTC GTTGACTGAC TTTACGGAGT TTTACAAGAA
50	4651 TACGATGCCA TTGGGATATA TCAACGGTGG TATATCCAGT GATTTTTTC ATGCTACGGT AACCCTATAT AGTTGCCACC ATATAGGTCA CTAAAAAAAG
55	4701 TCCATTTTAG CTTCCTTAGC TCCTGAAAAT CTCGATAACT CAAAAAATAC AGGTAAAATC GAAGGAATCG AGGACTTTTA GAGCTATTGA GTTTTTTATG
	4751 GCCCGGTAGT GATCTTATTT CATTATGGTG AAAGTTGGAA CCTCACCCGA CGGGCCATCA CTAGAATAAA GTAATACCAC TTTCAACCTT GGAGTGGGCT
	34/50

	4801 CGTCTAATGT GAGTTAGCTC ACTCATTAGG CACCCCAGGC TTTACACTTT GCAGATTACA CTCAATCGAG TGAGTAATCC GTGGGGTCCG AAATGTGAAA
5	4851 ATGCTTCCGG CTCGTATGTT GTGTGGAATT GTGAGCGGAT AACAATTTCA TACGAAGGCC GAGCATACAA CACACCTTAA CACTCGCCTA TTGTTAAAGT
	M13 Reverse primer 100.0% XbaI
10	4901 CACAGGAAAC AGCTATGACC ATGATTACGA ATTTCTAGAT AACGAGGGCA GTGTCCTTTG TCGATACTGG TACTAATGCT TAAAGATCTA TTGCTCCCGT
1.5	4951 AAAAATGAAA AAGACAGCTA TCGCGATTGC AGTGGCACTG GCTGGTTTCG TTTTTACTTT TTCTGTCGAT AGCGCTAACG TCACCGTGAC CGACCAAAGC
15	EcoRV
	5001 CTACCGTAGC GCAGGCCGAT GATGGCATCG CGTCCGGCTA
20	



36/50

Figure 29B lacZ' SD ompAXbaI 5 M K K T A I A I A V · SEQ ID NO:53 TCTAGATAAC GAGGGCAAAA AATGAAAAAG ACAGCTATCG CGATTGCAGT AGATCTATTG CTCCCGTTTT TTACTTTTTC TGTCGATAGC GCTAACGTCA Vk3 10 ompA **EcoRV** 15 ALA GFAT VAQ ADI VLTQ. GGCACTGGCT GGTTTCGCTA CCGTAGCGCA GGCCGATATC GTGCTGACCC CCGTGACCGA CCAAAGCGAT GGCATCGCGT CCGGCTATAG CACGACTGGG Vk3 20 · S P A T L S L S P G E R A T L S AGAGCCCGGC GACCCTGAGC CTGTCTCCGG GCGAACGTGC GACCCTGAGC 101 TCTCGGGCCG CTGGGACTCG GACAGAGGCC CGCTTGCACG CTGGGACTCG Vk3 CRASQSVSSSYLAWYQQ. 25 TGCAGAGCGA GCCAGAGCGT GAGCAGCAGC TATCTGGCGT GGTACCAGCA 151 ACGTCTCGCT CGGTCTCGCA CTCGTCGTCG ATAGACCGCA CCATGGTCGT Vk3 ·KPG QAPR LLI YGA SSRA· 30 GAAACCAGGT CAAGCACCGC GTCTATTAAT TTATGGCGCG AGCAGCCGTG 201 CTTTGGTCCA GTTCGTGGCG CAGATAATTA AATACCGCGC TCGTCGGCAC Vk3 · T G V P A R F S G S G S G T D F 35 CAACTGGGGT CCCGGCGCGT TTTAGCGGCT CTGGATCCGG CACGGATTTT 251 GTTGACCCCA GGGCCGCGCA AAATCGCCGA GACCTAGGCC GTGCCTAAAA Vk3 40 BbsI TLTISSLEPE DFAV YYC. ACCCTGACCA TTAGCAGCCT GGAACCTGAA GACTTTGCGG TGTATTATTG 301 TGGGACTGGT AATCGTCGGA CCTTGGACTT CTGAAACGCC ACATAATAAC 45 Vk3 MscI · Q Q H Y T T P P T F G Q G T K V E · CCAGCAGCAT TATACCACCC CGCCGACCTT TGGCCAGGGT ACGAAAGTTG 50 351 GGTCGTCGTA ATATGGTGGG GCGGCTGGAA ACCGGTCCCA TGCTTTCAAC CL kappa Vk3 55 **BsiWI** · I K R T V A A P S V F I F P P S AAATTAAACG TACGGTGGCT GCTCCGAGCG TGTTTATTTT TCCGCCGAGC 401

37/50

60

TTTAATTTGC ATGCCACCGA CGAGGCTCGC ACAAATAAAA AGGCGGCTCG

CL kappa DEQLKSGTAS VVCLLNN. 451 GATGAACAAC TGAAAAGCGG CACGGCGAGC GTGGTGTGCC TGCTGAACAA 5 CTACTTGTTG ACTTTTCGCC GTGCCGCTCG CACCACACGG ACGACTTGTT CL kappa ·FYPREAK VQW KVD NALQ· 501 CTTTTATCCG CGTGAAGCGA AAGTTCAGTG GAAAGTAGAC AACGCGCTGC GAAAATAGGC GCACTTCGCT TTCAAGTCAC CTTTCATCTG TTGCGCGACG 10 CL kappa \cdot S G N S Q E S V T E Q D S K D S 551 AAAGCGGCAA CAGCCAGGAA AGCGTGACCG AACAGGATAG CAAAGATAGC 15 TTTCGCCGTT GTCGGTCCTT TCGCACTGGC TTGTCCTATC GTTTCTATCG CL kappa TYSL SST LTL SKAD YEK. 601 ACCTATTCTC TGAGCAGCAC CCTGACCCTG AGCAAAGCGG ATTATGAAAA 20 TGGATAAGAG ACTCGTCGTG GGACTGGGAC TCGTTTCGCC TAATACTTTT CL kappa ·HKV YACE VTH QGL SSP V· ACATAAAGTG TATGCGTGCG AAGTGACCCA TCAAGGTCTG AGCAGCCCGG 651 25 TGTATTTCAC ATACGCACGC TTCACTGGGT AGTTCCAGAC TCGTCGGGCC CL kappa SphI StuI 30 · TKSFNRGEA 701 TGACTAAATC TTTTAATCGT GGCGAGGCCT GATAAGCATG CGTAGGAGAA ACTGATTTAG AAAATTAGCA CCGCTCCGGA CTATTCGTAC GCATCCTCTT phoA 35 SapI MK QSTIAL ALLP LLF. AATAAAATGA AACAAAGCAC TATTGCACTG GCACTCTTAC CGTTGCTCTT 751 TTATTTTACT TTGTTTCGTG ATAACGTGAC CGTGAGAATG GCAACGAGAA 40 VH3 phoA MfeI SapI 45 ·TPV TKAQ VQL VES GGGL· 801 CACCCCTGTT ACCAAAGCCG AAGTGCAATT GGTGGAAAGC GGCGGCGGCC GTGGGGACAA TGGTTTCGGC TTCACGTTAA CCACCTTTCG CCGCCGCGG VH3 50 · V Q P G G S L R L S C A A S G F TGGTGCAACC GGGCGGCAGC CTGCGTCTGA GCTGCGCGGC CTCCGGATTT 851 ACCACGTTGG CCCGCCGTCG GACGCAGACT CGACGCGCCG GAGGCCTAAA VH3 55 TFSS YAM SWV RQAP GKG. 901 ACCTTTAGCA GCTATGCGAT GAGCTGGGTG CGCCAAGCCC CTGGGAAGGG TGGAAATCGT CGATACGCTA CTCGACCCAC GCGGTTCGGG GACCCTTCCC VH₃ 60 38/50

E	951	·LEW VSAISGSGGSTYYA· TCTCGAGTGGGTGAGCGCGA TTAGCGGTAG CGGCGGCAGC ACCTATTATG AGAGCTCACC CACTCGCGCT AATCGCCATC GCCGCCGTCG TGGATAATAC VH3
5		PmlI
10	1001	D S V K G R F T I S R D N S K N CGGATAGCGT GAAAGGCCGT TTTACCATTT CACGTGATAA TTCGAAAAAC GCCTATCGCA CTTTCCGGCA AAATGGTAAA GTGCACTATT AAGCTTTTTG VH3
15	1051	T L Y L Q M N S L R A E D T A V Y · ACCCTGTATC TGCAAATGAA CAGCCTGCGT GCGGAAGATA CGGCCGTGTA TGGGACATAG ACGTTTACTT GTCGGACGCA CGCCTTCTAT GCCGGCACAT VH3
		BssHII
20	1101	· Y C A R W G G D G F Y A M D Y W G · TTATTGCGCG CGTTGGGGCG GCGATGGCTT TTATGCGATG GATTATTGGG AATAACGCGC GCAACCCCGC CGCTACCGAA AATACGCTAC CTAATAACCC CH1
25		VH3
		SalI
30		StyI BlpI
	1151	· Q G T L V T V S S A S T K G P S GCCAAGGCAC CCTGGTGACG GTTAGCTCAG CGTCGACCAA AGGTCCAAGC CGGTTCCGTG GGACCACTGC CAATCGAGTC GCAGCTGGTT TCCAGGTTCG CH1
35	1201	V F P L A P S S K S T S G G T A A · GTGTTTCCGC TGGCTCCGAG CAGCAAAAGC ACCAGCGGCG GCACGGCTGC CACAAAGGCG ACCGAGGCTC GTCGTTTTCG TGGTCGCCGC CGTGCCGACG CH1
40	1251	·L G C L V K D Y F P E P V T V S W · CCTGGGCTGC CTGGTTAAAG ATTATTTCCC GGAACCAGTC ACCGTGAGCT GGACCCGACG GACCAATTTC TAATAAAGGG CCTTGGTCAG TGGCACTCGA CH1
45	1301	N S G A L T S G V H T F P A V L GGAACAGCGG GGCGCTGACC AGCGGCGTGC ATACCTTTCC GGCGGTGCTG CCTTGTCGCC CCGCGACTGG TCGCCGCACG TATGGAAAGG CCGCCACGAC CH1
50	1351	Q S S G L Y S L S S V V T V P S S · CAAAGCAGCG GCCTGTATAG CCTGAGCAGC GTTGTGACCG TGCCGAGCAG GTTTCGTCGC CGGACATATC GGACTCGTCG CAACACTGGC ACGGCTCGTC CH1
55	1401	·S L G T Q T Y I C N V N H K P S N · CAGCTTAGGC ACTCAGACCT ATATTTGCAA CGTGAACCAT AAACCGAGCA GTCGAATCCG TGAGTCTGGA TATAAACGTT GCACTTGGTA TTTGGCTCGT CH1 CTgIII
60		39/50

EcoRI

00		40/50
60	1951	N K E S AATAAGGAGT CTTGATAAGC TTGACCTGTG AAGTGAAAAA TGGCGCAGAT TTATTCCTCA GAACTATTCG AACTGGACAC TTCACTTTTT ACCGCGTCTA lpp terminator
55		HindIII
		Stop lpp terminator
50	1901	· V A T F M Y V F S T F A N I L R ATGTTGCCAC CTTTATGTAT GTATTTTCTA CGTTTGCTAA CATACTGCGT TACAACGGTG GAAATACATA CATAAAAGAT GCAAACGATT GTATGACGCA CTgIII
45	1851	·D C D K I N L F R G V F A F L L Y · TGATTGTGAC AAAATAAACT TATTCCGTGG TGTCTTTGCG TTTCTTTAT ACTAACACTG TTTTATTTGA ATAAGGCACC ACAGAAACGC AAAGAAAATA CTgIII
40	1801	E C R P F V F G A G K P Y E F S I · GAATGTCGCC CTTTTGTCTT TGGCGCTGGT AAACCATATG AATTTTCTAT CTTACAGCGG GAAAACAGAA ACCGCGACCA TTTGGTATAC TTAAAAGATA CTgIII
35	1751	· L M N N F R Q Y L P S L P Q S V CTTTAATGAA TAATTTCCGT CAATATTTAC CTTCCCTCCC TCAATCGGTT GAAATTACTT ATTAAAGGCA GTTATAAATG GAAGGGAGGG AGTTAGCCAA CTgIII
30	1701	· A G S N S Q M A Q V G D G D N S P · TGCTGGCTCT AATTCCCAAA TGGCTCAAGT CGGTGACGGT GATAATTCAC ACGACCGAGA TTAAGGGTTT ACCGAGTTCA GCCACTGCCA CTATTAAGTG CTgIII
25	1651	I G D V S G L A N G N G A T G D F. ATTGGTGACG TTTCCGGCCT TGCTAATGGT AATGGTGCTA CTGGTGATTT TAACCACTGC AAAGGCCGGA ACGATTACCA TTACCACGAT GACCACTAAA CTgIII
20	1601	· K L D S V A T D Y G A A I D G F GCAAACTTGA TTCTGTCGCT ACTGATTACG GTGCTGCTAT CGATGGTTTC CGTTTGAACT AAGACAGCGA TGACTAATGC CACGACGATA GCTACCAAAG CTgIII
15	1551	· A M T E N A D E N A L Q S D A K G · GGCTATGACC GAAAATGCCG ATGAAAACGC GCTACAGTCT GACGCTAAAG CCGATACTGG CTTTTACGGC TACTTTTGCG CGATGTCAGA CTGCGATTTC CTgIII
10	1501	S G S G D F D Y E K M A N A N K G · AGCGGGAGCG GTGATTTTGA TTATGAAAAG ATGGCAAACG CTAATAAGGG TCGCCCTCGC CACTAAAACT AATACTTTTC TACCGTTTGC GATTATTCCC CTgIII
5	1451	T K V D K K V E P K S E F G G G ACACCAAAGT GGATAAAAAA GTGGAACCGA AAAGCGAATT CGGGGGAGGG TGTGGTTTCA CCTATTTTTT CACCTTGGCT TTTCGCTTAA GCCCCCTCCC CTgIII

	2001	TGTGCGACAT TTTTTTTGTC TGCCGTTTAA TGAAATTGTA AACGTTAATA ACACGCTGTA AAAAAAACAG ACGGCAAATT ACTTTAACAT TTGCAATTAT
5		fl origin
5	2051	TTTTGTTAAA ATTCGCGTTA AATTTTTGTT AAATCAGCTC ATTTTTTAAC AAAACAATTT TAAGCGCAAT TTAAAAAACAA TTTAGTCGAG TAAAAAATTG
		f1 origin
10	2101	CAATAGGCCG AAATCGGCAA AATCCCTTAT AAATCAAAAG AATAGACCGA GTTATCCGGC TTTAGCCGTT TTAGGGAATA TTTAGTTTTC TTATCTGGCT
		f1 origin
15	2151	GATAGGGTTG AGTGTTGTTC CAGTTTGGAA CAAGAGTCCA CTATTAAAGA CTATCCCAAC TCACAACAAG GTCAAACCTT GTTCTCAGGT GATAATTTCT
		fl origin
20	2201	ACGTGGACTC CAACGTCAAA GGGCGAAAAA CCGTCTATCA GGGCGATGGC TGCACCTGAG GTTGCAGTTT CCCGCTTTTT GGCAGATAGT CCCGCTACCG
20		fl origin
		T->A
	20.51	~
25	2251	CCACTACGAG AACCATCACC CTAATCAAGT TTTTTGGGGT CGAGGTGCCG GGTGATGCTC TTGGTAGTGG GATTAGTTCA AAAAACCCCA GCTCCACGGC
		fl origin
	2301	TAAAGCACTA AATCGGAACC CTAAAGGGAG CCCCCGATTT AGAGCTTGAC
30		ATTTCGTGAT TTAGCCTTGG GATTTCCCTC GGGGGCTAAA TCTCGAACTG
50		f1 origin
	2351	GGGGAAAGCC GGCGAACGTG GCGAGAAAGG AAGGGAAGAA AGCGAAAGGA CCCCTTTCGG CCGCTTGCAC CGCTCTTTCC TTCCCTTCTT TCGCTTTCCT
35		fl origin
	2401	GCGGGCGCTA GGGCGCTGGC AAGTGTAGCG GTCACGCTGC GCGTAACCAC CGCCGCGAT CCCGCGACCG TTCACATCGC CAGTGCGACG CGCATTGGTG
		fl origin
40		NheI
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	2451	CACACCGCC GCGCTTAATG CGCCGCTACA GGGCGCGTGC TAGCCATGTG GTGTGGGCGG CGCGAATTAC GCGGCGATGT CCCGCGCACG ATCGGTACAC
45		fl origin ColEI
	2501	AGCAAAAGGC CAGCAAAAGG CCAGGAACCG TAAAAAAGGCC GCGTTGCTGG TCGTTTTCCG GTCGTTTTCC GGTCCTTGGC ATTTTTCCGG CGCAACGACC
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50		ORI ~
	2551	CGTTTTTCCA TAGGCTCCGC CCCCCTGACG AGCATCACAA AAATCGACGC GCAAAAAGGT ATCCGAGGCG GGGGGACTGC TCGTAGTGTT TTTAGCTGCG
55		ColEI
	2601	TCAAGTCAGA GGTGGCGAAA CCCGACAGGA CTATAAAGAT ACCAGGCGTT AGTTCAGTCT CCACCGCTTT GGGCTGTCCT GATATTTCTA TGGTCCGCAA
		ColEI
60	2651	TCCCCCTGGA AGCTCCCTCG TGCGCTCTCC TGTTCCGACC CTGCCGCTTA
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AGGGGGACCT TCGAGGGAGC ACGCGAGAGG ACAAGGCTGG GACGGCGAAT ColEI mutation 5 CCGGATACCT GTCCGCCTTT CTCCCTTCGG GAAGCGTGGC GCTTTCTCAT 2701 GGCCTATGGA CAGGCGGAAA GAGGGAAGCC CTTCGCACCG CGAAAGAGTA ColEI 10 mutation AGCTCACGCT GTAGGTATCT CAGTTCGGTG TAGGTCGTTC GCTCCAAGCT 2751 TCGAGTGCGA CATCCATAGA GTCAAGCCAC ATCCAGCAAG CGAGGTTCGA 15 ColEI mutation GGGCTGTGTG CACGAACCCC CCGTTCAGTC CGACCGCTGC GCCTTATCCG 2801 CCCGACACAC GTGCTTGGGG GGCAAGTCAG GCTGGCGACG CGGAATAGGC 20 ColEI GTAACTATCG TCTTGAGTCC AACCCGGTAA GACACGACTT ATCGCCACTG 2851 CATTGATAGC AGAACTCAGG TTGGGCCATT CTGTGCTGAA TAGCGGTGAC 25 ColEI GCAGCAGCCA CTGGTAACAG GATTAGCAGA GCGAGGTATG TAGGCGGTGC 2901 CGTCGTCGGT GACCATTGTC CTAATCGTCT CGCTCCATAC ATCCGCCACG ColEI 30 mutation TACAGAGTTC TTGAAGTGGT GGCCTAACTA CGGCTACACT AGAAGAACAG 2951 ATGTCTCAAG AACTTCACCA CCGGATTGAT GCCGATGTGA TCTTCTTGTC 35 ColEI mutation 3001 TATTTGGTAT CTGCGCTCTG CTGTAGCCAG TTACCTTCGG AAAAAGAGTT ATAAACCATA GACGCGAGAC GACATCGGTC AATGGAAGCC TTTTTCTCAA 40 ColEI GGTAGCTCTT GATCCGGCAA ACAAACCACC GCTGGTAGCG GTGGTTTTTT 3051 CCATCGAGAA CTAGGCCGTT TGTTTGGTGG CGACCATCGC CACCAAAAAA 45 ColEI 3101 TGTTTGCAAG CAGCAGATTA CGCGCAGAAA AAAAGGATCT CAAGAAGATC ACAAACGTTC GTCGTCTAAT GCGCGTCTTT TTTTCCTAGA GTTCTTCTAG ColEI 50 3151 CTTTGATCTT TTCTACGGGG TCTGACGCTC AGTGGAACGA AAACTCACGT GAAACTAGAA AAGATGCCCC AGACTGCGAG TCACCTTGCT TTTGAGTGCA ColEI cat terminator 55 BglII TAAGGGATTT TGGTCAGATC TAGCACCAGG CGTTTAAGGG CACCAATAAC 3201 ATTCCCTAAA ACCAGTCTAG ATCGTGGTCC GCAAATTCCC GTGGTTATTG 60 42/50

ColEI cat terminator TGCCTTAAAA AAATTACGCC CCGCCCTGCC ACTCATCGCA GTACTGTTGT 3251 5 ACGGAATTTT TTTAATGCGG GGCGGGACGG TGAGTAGCGT CATGACAACA CM(R) 3301 AATTCATTAA GCATTCTGCC GACATGGAAG CCATCACAAA CGGCATGATG TTAAGTAATT CGTAAGACGG CTGTACCTTC GGTAGTGTTT GCCGTACTAC 10 CM(R)AACCTGAATC GCCAGCGGCA TCAGCACCTT GTCGCCTTGC GTATAATATT 3351 TTGGACTTAG CGGTCGCCGT AGTCGTGGAA CAGCGGAACG CATATTATAA 15 CM(R) TGCCCATAGT GAAAACGGGG GCGAAGAAGT TGTCCATATT GGCTACGTTT 3401 ACGGGTATCA CTTTTGCCCC CGCTTCTTCA ACAGGTATAA CCGATGCAAA CM(R) AAATCAAAAC TGGTGAAACT CACCCAGGGA TTGGCTGAGA CGAAAAACAT 20 3451 TTTAGTTTTG ACCACTTTGA GTGGGTCCCT AACCGACTCT GCTTTTTGTA CM(R) ATTCTCAATA AACCCTTTAG GGAAATAGGC CAGGTTTTCA CCGTAACACG 3501 25 TAAGAGTTAT TTGGGAAATC CCTTTATCCG GTCCAAAAGT GGCATTGTGC CM(R) CCACATCTTG CGAATATATG TGTAGAAACT GCCGGAAATC GTCGTGGTAT 3551 GGTGTAGAAC GCTTATATAC ACATCTTTGA CGGCCTTTAG CAGCACCATA 30 CM(R)TCACTCCAGA GCGATGAAAA CGTTTCAGTT TGCTCATGGA AAACGGTGTA 3601 AGTGAGGTCT CGCTACTTTT GCAAAGTCAA ACGAGTACCT TTTGCCACAT 35 CM(R)ACAAGGGTGA ACACTATCCC ATATCACCAG CTCACCGTCT TTCATTGCCA 3651 TGTTCCCACT TGTGATAGGG TATAGTGGTC GAGTGGCAGA AAGTAACGGT CM(R) 40 TACGGAACTC CGGGTGAGCA TTCATCAGGC GGGCAAGAAT GTGAATAAAG 3701 ATGCCTTGAG GCCCACTCGT AAGTAGTCCG CCCGTTCTTA CACTTATTTC CM(R) GCCGGATAAA ACTTGTGCTT ATTTTTCTTT ACGGTCTTTA AAAAGGCCGT 3751 45 CGGCCTATTT TGAACACGAA TAAAAAGAAA TGCCAGAAAT TTTTCCGGCA CM(R)AATATCCAGC TGAACGGTCT GGTTATAGGT ACATTGAGCA ACTGACTGAA 3801 TTATAGGTCG ACTTGCCAGA CCAATATCCA TGTAACTCGT TGACTGACTT 50 CM(R) ATGCCTCAAA ATGTTCTTTA CGATGCCATT GGGATATATC AACGGTGGTA 3851 TACGGAGTTT TACAAGAAAT GCTACGGTAA CCCTATATAG TTGCCACCAT 55 CM(R) TATCCAGTGA TTTTTTTCTC CATTTTAGCT TCCTTAGCTC CTGAAAATCT 3901 ATAGGTCACT AAAAAAAGAG GTAAAATCGA AGGAATCGAG GACTTTTAGA SDCM(R) 60 43/50

	3951	cat promoter CGATAACTCA AAAAATACGC CCGGTAGTGA TCTTATTTCA TTATGGTGAA GCTATTGAGT TTTTTATGCG GGCCATCACT AGAATAAAGT AATACCACTT
5		cat promoter CRP site
10	4001	AGTTGGAACC TCACCCGACG TCTAATGTGA GTTAGCTCAC TCATTAGGCA TCAACCTTGG AGTGGGCTGC AGATTACACT CAATCGAGTG AGTAATCCGT
10		cat promoter lac mRNA start
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20	4051	CCCCAGGCTT TACACTTTAT GCTTCCGGCT CGTATGTTGT GTGGAATTGT GGGGTCCGAA ATGTGAAATA CGAAGGCCGA GCATACAACA CACCTTAACA lac operator SD lacZ'
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MSPro55 (SEQ ID NO:69)	GAT	ATC	GCA	CIG	ACC	CAG	CCA	GCT	-	TCA		AGC (						AGC
MSPro11 (SEQ ID NO:70)	GAT	ATC	GCA					GCT	٠.									AGC
MSPro26 (SEQ ID NO:71)	GAT	ATC	GCA					EJ					GGC					AGC
VL13 (SEQ ID NO:72)	GAT	ATC	GAA		ACC		_	EJ	•				_					ACC
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TAT	CIL	TLL	GAT	CLL	TGG	ည္ဟမ္မာ	CAA	ည္ဟ	ACC	CIG	GTG	ACG	GIL	AGC	TCA	<del>- ပ</del> ္ပ
×	×	×	GAT	×	TGG	ည္သမ္မ	CCAA	<u> </u>	ACC	CIG	GTG	ACG	GTT	AGC	TCA	ပ္ပ
E C C	CAT	TII	GAT	TAT	TGG	ည္ဟမ	CCAA	ပ္ပ	ACC	CIG	GTG	ACG	GTT	AGC	TCA	ပ္ပ
GTT	CIL	TII	GAT	CAT		ည္သမ္မ		ည္ဟ	ACC	CIG	GTG	ACG	GTT	AGC	TCA	ပ္ပ
TAT	GAG	TTT	GAT		TGG	ည္ဟ	CCAA	ည	ACC	CTG	GIG		_	AGC	TCA	ပ္ပ
GGT	TLL	ATT	GAT	. *		255	CCAA	ည္ပ	ACC	CTG	GTG	_	GTT	AGC	TCA	ပ္ပ
GGT	TAT		,	AAT	TGG	ည္သမ္	CCAA	ည	ACC	CTG	GTG	ACG	GTT	AGC	TCA	ပ္ပ
TAT	TAT	LLL	GAT	ATT	TGG	ည္သမ္မ	CCAA	ည္ဟ	ACC	CIG	GTG	ACG	GTI	AGC	TCA	ပ္ပ
×	×		GAT	×	TGG	ည္ဟမ္	CCAA	ည္ဟ	ACC	CIG	GTG	ACG	GTT	AGC	TCA	ပ္ပ
TAT	ATG	TII	GAT	TAT	TGG	ည္ပမ္	CCAA	ည္ဟ	ACC	CIG	GIG	ACG	GIT	AGC	TCA	ပ္ပ
×	×	×	GAT	×	TGG	ည္ဟ	CCAA	ည	ACC	CTG	GIG	ACG	GTT	AGC	TCA	ည
CCI	GAT	TLL	GAT	TAT	TGG	ບູ	CAA	<u> </u>	ACC	CIG	GTG	ACG	GTT	AGC	TCA	ည